

Polyphenolic Characterisation of Vranac, Kratosija and Cabernet Sauvignon (*Vitis vinifera* L. cv.) Grapes and Wines from Different Vineyard Locations in Montenegro

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In Montenegro, red wines are produced predominantly, and Vranac accounts for nearly 80% of these wines, followed by Kratosija and Cabernet Sauvignon. In order to characterise polyphenols in red varieties, grapes were sampled from representative vineyards at harvest time during 2011 and 2012. The content and distribution of extractable anthocyanins, low-molecular mass proanthocyanidins (LMP) and high-molecular mass proanthocyanidins (HMP) in the seeds and skins of the grape berries were evaluated by applying a five-day extraction method using ethanol:water (12:88) as extraction solvent. On average, the highest content of LMP (2 006 and 1 690 mg/kg of grape fresh mass in years 2011 and 2012 respectively), HMP (2 705 and 2 805 mg/kg in years 2011 and 2012 respectively) and anthocyanins (1 035 mg/kg in the year 2011) was found in the Cabernet Sauvignon grapes. The highest content of anthocyanins (1 113 mg/kg in the year 2012) and the lowest content of LMP (1 103 and 846 mg/kg in years 2011 and 2012 respectively) was found in Vranac grapes. Kratosija grapes had the lowest anthocyanin content (456 and 517 mg/kg in years 2011 and 2012 respectively), and levels of LMP were similar to Vranac. The percentage distributions of LMP between skins and seeds were 34:66, 39:61 and 49:51, whereas the distributions of HMP between skins and seeds were 67:33, 62:38 and 64:36 for Vranac, Kratosija and Cabernet Sauvignon respectively. All varieties had more LMP in the seeds and more HMP in the skins of the grapes. The results obtained are important to better understand the polyphenolic potential of Montenegrin red grape varieties.

INTRODUCTION

Grape growing and winemaking in Montenegro started to develop in ancient times due to the geographical position and warm climate of the area. Winemaking is traditionally based on local varieties such as Vranac and Kratosija. Kratosija dominated until the outbreak of phylloxera, although Vranac became the preferred grape variety due to its lower heterogeneity and improved skin coloration (Ulicevic, 1966; Pejovic, 1988). Vranac and Kratosija are also cultivated in Macedonia, Bosnia and Herzegovina, Croatia and Serbia and are considered as indigenous varieties of the Western Balkan countries (Bozinovic, 2005). The winegrowing areas in Montenegro are increasing due to renewed investments in wine production, and are producing both local and international grape varieties. Vranac is the dominant grape among red grapes, whereas Kratosija is decreasing and Cabernet Sauvignon is increasing (Pajovic *et al.*, 2011).

Vranac grapes are considered to have a strong polyphenol potential (Ivanova *et al.*, 2011), as well as a high colour potential (Avramov, 1991). In contrast, Pajovic *et al.* (2009) reported about 300 to 400 mg/L of anthocyanins in young Vranac wines.

Flavonoids, i.e. anthocyanins and proanthocyanidins (also called grape condensed tannins), account for the major part of red wine polyphenols that have an impact on the sensorial quality of red wines; they are wine preservatives and the basis for ageing. Anthocyanins are responsible for the red colour (Ribéreau-Gayon, 1982) and proanthocyanidins for the colour stability (Somers, 1971), the taste of bitterness and the mouthfeel of astringency (Robichaud & Noble, 1990). Generally, the astringency of proanthocyanidins increases with chain length and the bitterness decreases (Peleg *et al.*, 1999; Chira *et al.*, 2009). Other studies, however, report that differences in astringency are mainly due to the total

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content of proanthocyanidins in wines (Brossaud *et al.*, 2001; Preys *et al.*, 2006). Flavonoids, besides having technological importance, are also strong antioxidants that are important for human health (Rodrigo *et al.*, 2011). The levels of anthocyanins and proanthocyanidins and the distribution of proanthocyanidins between the skin and seeds of grape berries are determined by grape variety, and by climatic and pedological conditions (Mattivi *et al.*, 2002a, 2009). The grape variety also has an impact on the degree of proanthocyanidin polymerisation (Chira *et al.*, 2009; Mattivi *et al.*, 2009), which affects the extractability from skins and seeds during the winemaking process (Gambuti *et al.*, 2009). Generally, skin proanthocyanidins are extracted earlier during the fermentation process and, as the maceration time increases, the extraction of seed proanthocyanidins increases (Peyrot des Gachons & Kennedy, 2003).

The estimation of the polyphenolic potential allows the identification of differences in the polyphenol composition and provides factors with which to evaluate the oenological potential of the grape. Not many studies have been conducted on the polyphenol content of *Vitis vinifera* red grape varieties in the Western Balkan region, and the polyphenol content was obtained by the extraction of grape seeds and skin in organic solvents (Ivanova *et al.*, 2010, 2011). The content and structure of the polyphenols extracted by the maceration process can differ from those extracted by means of strong organic solvents, since high-molecular mass proanthocyanidins are unlikely to be extracted to any great extent in a wine-like solution (Mattivi *et al.*, 2009). The aim of this study therefore was to characterise Vranac, Kratosija and Cabernet Sauvignon grapes from representative Montenegrin vineyard locations according to the extractable polyphenol content and polyphenol distribution between the skin and the seeds of the grape berries. By using an extraction method that simulates the process of maceration (Mattivi

et al., 2002b), the content of extractable total polyphenols, total anthocyanins, low-molecular mass proanthocyanidins (LMP) and high-molecular mass proanthocyanidins (HMP) have been determined separately in the skin and seeds of grape berries. The same polyphenol groups were evaluated in microvinified wines produced from the same grape.

MATERIALS AND METHODS

Chemicals and reagents

Methanol, ethanol, hydrochloric acid, sodium hydroxide, sodium bisulphite and L(+)-tartaric acid were purchased from Sigma Aldrich (St. Louis, MO, USA) and Merck (Darmstadt, Germany). Ultra pure water was of Milli Q grade (Millipore Corporation, Billerica, MA, USA). The reagents Folin-Ciocalteu and vanillin were from Merck.

Montenegrin wine region

The Montenegrin wine region consists of two principal regions, one lying around the basin of Lake Skadar and the other along the coastal area on the Adriatic Sea. The majority of Montenegrin vineyards (almost 90%) are located in the Podgorica district, in the basin of Lake Skadar (Fig. 1). The Podgorica district is characterised by a high Winkler index, making it favourable for growing red grape varieties, being > 2800 and > 2300 in 2011 and 2012 respectively. The Huglin index also proved to be high, at > 3700 and > 3400 in the same seasons. The average vegetation temperature was 23.1°C and 20.7°C , whereas the precipitation in the same vegetation period was 311 mm and 902 mm in 2011 and 2012 respectively (MONSTAT, 2013).

Grape sampling

Grapes were sampled at the time of their technological maturity, between 3 and 25 September in 2011 and 2012. Grape samples of Vranac ($n = 6$), Kratosija ($n = 5$) and

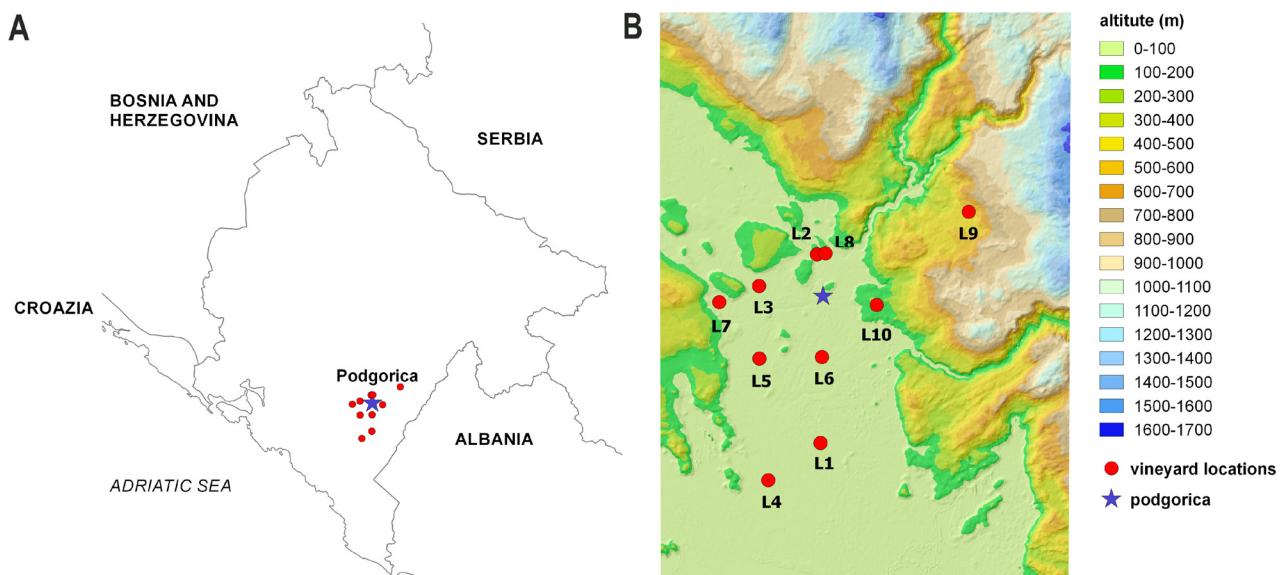


FIGURE 1

Geographical position of Montenegro and location of vineyards where grapes were sampled (A). Vineyard locations at different altitudes (B): 1 – Sipcanik, 2 – Rogami, 3 – Ljeskopolje, 4 – Zeta, 5 – Kokoti, 6 – Nikolj Crkva, 7 – Beri, 8 – Piperi, 9 – Kuci, 10 – Kakaricka Gora.

TABLE 1

The characteristics of the vineyards from which grapes were sampled.

Locality	Rootstock	Planting year	Plant density	Training system	Row orientation
VRANAC					
L1	Kober 5BB	1978	4 274 (2.6 x 0.9 m)	Double Guyot	north - south
L2	Teleki SO4	2006	6 211 (2.3 x 0.7m)	Single Guyot	north - south
L3	Kober 5BB	2005	4 000 (2.5 x 1.0 m)	Double Guyot	north - south
L4	Kober 5BB	2008	4 167 (2.4 x 1.0 m)	Double Guyot	north - south
L5	Kober 5BB	2008	5 714 (2.5 x 0.7 m)	Single Guyot	north - south
L7	Kober 5BB	1989	6 944 (1.6 x 0.9 m)	Double Guyot	north - south
KRATOSIJA					
L1	1103 Paulsen	2006	5 495 (2.6 x 0.7 m)	Single Guyot	north - south
L2	Teleki SO4	2006	6 211 (2.3 x 0.7m)	Single Guyot	north - south
L3	Kober 5BB	2005	4 000 (2.5 x 1.0 m)	Double Guyot	north - south
L6	Kober 5BB	1996	4 808 (2.6 x 0.8 m)	Double Guyot	north - south
L9	Kober 5BB	1983	2 500 (2.0 x 2.0 m)	Pergola	north - south
CABERNET SAUVIGNON					
L1	1103 Paulsen	2006	5 495 (2.6 x 0.7 m)	Single Guyot	north - south
L2	Teleki SO4	2006	6 211 (2.3 x 0.7m)	Single Guyot	north - south
L5	Kober 5BB	2008	5 714 (2.5 x 0.7 m)	Single Guyot	north - south
L6	Kober 5BB	1996	4 808 (2.6 x 0.8 m)	Single Guyot	north - south
L8	140 Ruggeri	2006	7 937 (1.8 x 0.7 m)	Single Guyot	north - south
L10	Kober 5BB	2000	5 714 (2.5 x 0.7 m)	Single Guyot	east - west

Cabernet Sauvignon (n = 6) were collected from vineyards located in the Podgorica wine-growing district (Fig. 1A). Vineyard characteristics are presented in Table 1. The yield of the vineyards was not reduced and water was applied by drip irrigation, thus water status in the field was not checked. Approximately 20 kg of grapes from each vineyard were representatively sampled. The vineyard locations were as follows: Sipcanik (1), Rogami (2), Ljeskopolje (3), Zeta (4), Kokoti (5), Nikolj Crkva (6), Beri (7), Piperi (8), Kuci (9) and Kakaricka Gora (10). Location 9 and location 10 lie at 250 and 400 m above sea level, while the others lie between 25 and 50 m above sea level (Fig. 1B).

Determination of grape physicochemical characteristics
Grape samples of 100 berries each were representatively collected from the 20 kg sample and weighed. Total soluble solids (TSS), titratable acidity and the pH of the berry juice were determined following International Organisation of Vine and Wine procedures (OIV, 2011).

Polyphenol extraction from grapes

A selective extraction of polyphenols from the skins and seeds of grape berries that simulates the maceration process of red wines was used (Mattivi *et al.*, 2002b; Vacca *et al.*, 2009). Skins and seeds of 200 g of randomly sampled grape berries were manually separated and separately extracted for five days at 30°C in a 200 mL solution consisting of ethanol:water (12:88 v/v), 100 mg/L of SO₂, 5 g/L tartaric acid and a pH value adjusted to 3.2 (with NaOH). The extracts were shaken by hand once a day. Skins and seeds were removed from the hydro-alcoholic solution after five days and the skin extract was centrifuged for 10 min at 3500 × g. Extracts were poured

into dark glass bottles, flushed with nitrogen and stored at 4°C until required for spectrophotometric analyses. Analyses were conducted four months later.

Vinification

The harvested grapes (20 kg), which originated from different localities, were microvinified at the winery of the Biotechnical Faculty in Podgorica during the 2011 and 2012 seasons. The grapes were destemmed and crushed and sodium metabisulphite was added (5 g of SO₂ to 100 kg of grapes). Vinification was conducted by spontaneous fermentation without repetitions due to the limited grapes available for vinification. Maceration lasted for seven days at a temperature between 25 and 28°C. The cap was punched down twice a day throughout the skin-contact period. Wines were decanted (sugar level < 2 g/L) without pressing at the end of alcoholic fermentation. Free-run wines were stored at room temperature. After the spontaneous malolactic fermentation, the wines were decanted again and SO₂ was added. Polyphenols in the wines were analysed four months after the fermentation.

Spectrophotometric analyses

Analyses were performed using a Varian Cary 100 spectrophotometer (Bio Tech, Maryland, USA), as described by Di Stefano and Guidoni (1989) and Di Stefano *et al.* (1989), under optimising conditions for red wine analysis (Rigo *et al.*, 2000). Polar compounds such as sugars, organic acids, amino acids and free SO₂, which could interfere with the assays, were removed by clean-up of grape extracts and wine using Sep-Pak classic (0.35 g) C-18 columns supplied by Waters (Milford, MA, USA).

Total polyphenols

Total polyphenols (TP) were assessed by the reduction of Folin-Ciocalteu reagent to blue pigments caused by polyphenols in alkaline solution. A realistic estimation of total polyphenols can be obtained only after preliminary cleaning of samples from other compounds (free SO₂, sugars, etc.) that interfere with the assay (Di Stefano & Guidoni, 1989). When the absorbance was between 0.3 and 0.6 AU (the linear response range), the results were expressed against the corresponding blank as (+)-catechin = $186.5 \times A \times d$ in mg/kg grape fresh mass (FM) or in mg/L of wine; A = absorbance and d = sample dilution.

High-molecular mass proanthocyanidins

HMP were evaluated by transformation into cyanidin (Di Stefano *et al.*, 1989). When the absorbance was between 0.3 and 0.6 AU the results were expressed against the corresponding blank as cyanidin chloride = $1162.5 \times \Delta A \times d$ in mg/kg grape FM or in mg/L of wine; ΔA = difference in absorbance between sample and blank, and d = sample dilution. The method provides a good estimation for the evaluation of HMP (Vrhovsek *et al.*, 2001).

Low-molecular mass proanthocyanidins – index of vanillin

The catechins and proanthocyanidins reactive to vanillin were analysed according to the optimised and controlled vanillin-HCl method of Broadhurst and Jones (1978), following the conditions described by Di Stefano *et al.* (1989). The method provides an estimation of the free carbon 6 and carbon 8 of the A-ring of both catechins and proanthocyanidins. This index decreases with the increase in polymerisation, because mainly carbon 6 and carbon 8 are involved in polymerisation bonds. The method provides a good estimation of free flavanols and a low degree of polymerised flavanols. When the absorbance was between 0.2 and 0.4 AU, the LMP were evaluated as (+)-catechin = $290.8 \times \Delta A \times d$ in mg/kg grape FM or in mg/L of wine; ΔA = difference in absorbance between sample and blank and d = sample dilutions.

Total anthocyanins

Total anthocyanins (TA) were determined on the basis of maximal absorbance in the visible range (536 to 542 nm). When the absorbance was between 0.3 and 0.6 AU, the results were expressed against the corresponding blank as TA = $A \times 26.6 \times 4 \times d$ in mg/kg grape FM or in mg/L of wine (d = sample dilution) by assuming an average absorbance of the mixture of anthocyanins extracted from Cabernet Sauvignon grapes (average MW = 500 Da, $\epsilon = 18800 \text{ M}^{-1} \text{ cm}^{-1}$ in 70:30:1 ethanol:water:HCl solution) (Di Stefano *et al.*, 1989).

Statistical analysis

Data was processed by ANOVA (p indicated) and, when significant, the means were separated using Tukey's honest significant difference (HSD) test ($p < 0.05$). Statistical analysis was performed using the Statgraphics Centurion XVI program (Manugistics Inc., Rockville, MD, USA).

RESULTS AND DISCUSSION

Physicochemical characteristics of Vranac, Kratosija and Cabernet Sauvignon grapes at the time of harvest

The physicochemical characteristics of Vranac, Kratosija and Cabernet Sauvignon grapes at the time of harvest from different vineyard locations in the Podgorica district in 2011 and 2012 are listed in Table 2. The average mass of 100 berries did not differ significantly between the Vranac and Kratosija varieties in 2011 (240 g and 242 g, respectively) and 2012 (217 and 222 g, respectively) (Table 2). The average berry mass of Cabernet Sauvignon was significantly lower (113 g in 2011 and 2012) than that of the Vranac and Kratosija varieties in both years. Grape berries of Vranac and Kratosija are almost twice as heavy as those of Cabernet Sauvignon and have a significantly lower percentage of skins by berry weight compared to Cabernet Sauvignon grapes. Average TSS content (Table 2) at the time of sampling was the highest in the grape juice of Kratosija (23.8°Brix in 2011 and 2012), followed by Cabernet Sauvignon (23.7 and 22.5°Brix). Vranac grape juice reported the lowest TSS values (21.7 and 20.7°Brix).

The acidity of the grape juice was low, as is typical for warm climates such as the Montenegro region. Significant lower titratable acidity (TA) was assessed in the Vranac grapes (5.4 g/L and 4.9 g/L for 2011 and 2012, respectively) compared to the Kratosija grapes (6.4 g/L and 6.5 g/L), even if not significantly different from Cabernet Sauvignon (5.8 g/L and 6.0 g/L). At the time of harvest, Vranac grapes proved to be higher in pH value compared to Kratosija and Cabernet Sauvignon in both years, although differences among the varieties were not statistically significant.

Contents of extractable total polyphenols, low- and high-molecular mass proanthocyanidins and total anthocyanins in grapes of Vranac, Kratosija and Cabernet Sauvignon

The content of extractable polyphenols (in mg/kg grape FM) in Vranac, Kratosija and Cabernet Sauvignon at the time of harvest from different vineyard locations in Podgorica district in 2011 and 2012 is shown in Table 3. The concentration levels represent the sum of extractable polyphenols in the skin and in the seeds of grape berries evaluated as the content in mg/kg of grape FM.

The mean content of total extractable polyphenols in the skin and seeds of grape berries was the highest in Cabernet Sauvignon in both years (2 705 mg/kg and 2 017 mg/kg in 2011 and 2012, respectively) and was significantly higher compared to Kratosija grapes (1 699 mg/kg and 1 097 mg/kg). The content of extractable polyphenols in Vranac grapes was intermediate (1 908 mg/kg and 1 598 mg/kg in 2011 and 2012, respectively), but not significantly different compared to Kratosija and Cabernet Sauvignon. The results presented here are in compliance with the extractable total polyphenol content in red *Vitis vinifera* grape varieties grown in Slovenia. Vrhovsek *et al.* (2002) showed that the average extractable total polyphenol content in grapes of red *Vitis vinifera* varieties in a two-year study was between 1 100 and 2 100 mg/kg grape FM, and the highest content was reported in Cabernet Sauvignon grapes (2 000 to

TABLE 2

Physicochemical characteristics (MB - Mass of 100 berries, TSS - Total soluble solids, TA - Titratable acidity, SKIN/BERRY - percent of skin by berry weight) of Vranac, Kratosija and Cabernet Sauvignon grapes at the time of harvest.

Locality	2011					2012				
	MB (g)	TSS (°Brix)	TA (g/L tartaric acid)	pH	SKIN/BERRY (%)	MB (g)	TSS (°Brix)	TA (g/L tartaric acid)	pH	SKIN/BERRY (%)
VRANAC										
L1	257	22.9	5.0	3.53	8.8	250	19.2	4.7	3.56	9.8
L2	219	22.7	5.5	3.48	9.8	207	21.6	5.1	3.45	10.4
L3	210	22.3	5.3	3.46	10.2	200	23.5	5.3	3.45	10.6
L4	258	19.6	5.6	3.50	9.3	214	17.6	5.6	3.40	10.9
L5	224	23.1	5.1	3.60	9.9	212	23.0	4.0	3.59	10.2
L7	271	19.6	5.6	3.69	9.1	217	19.5	4.6	3.58	10.7
Mean	240 a	21.7	5.4 b	3.54	9.5 b	217 a	20.7 b	4.9 b	3.51	10.4 b
KRATOSIJA										
L1	238	22.0	6.3	3.57	10.6	225	22.5	9.0	3.36	11.2
L2	242	24.4	6.2	3.47	10.4	224	24.5	5.9	3.30	10.9
L3	233	28.0	6.7	3.53	10.1	211	25.4	6.6	3.58	11.4
L6	244	22.6	6.1	3.49	10.7	195	24.6	4.9	3.36	11.9
L9	255	22.2	6.6	3.48	9.7	253	21.8	6.0	3.46	9.8
Mean	242 a	23.8	6.4 a	3.51	10.3 b	222 a	23.8 a	6.5 a	3.41	11.0 b
CABERNET SAUVIGNON										
L1	131	22.3	5.9	3.41	12.5	95	22.9	7.1	3.20	16.2
L2	128	22.9	5.9	3.48	13.1	112	22.9	5.6	3.44	14.4
L5	83	23.1	6.5	3.64	15.8	133	22.1	5.8	3.40	12.6
L6	117	24.1	5.9	3.41	13.6	110	22.5	5.8	3.40	15.4
L8	107	25.7	5.6	3.45	15.9	111	21.2	5.6	3.35	15.1
L110	110	24.0	5.1	3.58	15.4	118	23.2	6.0	3.68	13.9
Mean	113 b	23.7	5.8 b	3.50	14.4 a	113 b	22.5 ab	6.0 ab	3.41	14.6 a
sign. F	***	n.s.	***	n.s.	***	***	*	*	n.s.	***

ANOVA was used to compare data (n.s. not significant, * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$). Different lower-case letters indicate significant differences of means between varieties using Tukey's HSD test ($p \leq 0.05$).

2 100 mg/kg). The extractable anthocyanin content of the grapes did not differ significantly between Vranac and Cabernet Sauvignon in 2011 (1 035 mg/kg and 960 mg/kg, respectively), whereas the extractable anthocyanin content of grapes harvested during 2012 was significantly higher in Vranac grapes compared to Cabernet Sauvignon grapes (1 113 mg/kg and 861 mg/kg respectively). Kratosija grapes proved to have a significantly lower content of extractable anthocyanins (Table 3) compared to Vranac and Cabernet Sauvignon in both years (456 mg/kg and 517 mg/kg in 2011 and 2012, respectively). The extractable anthocyanin content (determined by the same method) in Slovenian red grapes was 300 to 800 mg/kg in Pinot Noir, 700 to 900 mg/kg in Blaufränkisch, 900 to 1100 mg/kg in Barbera and Syrah and 1 100 to 1 300 mg/kg in Merlot, Refosk and Cabernet Sauvignon (Vrhovsek *et al.*, 2002), whereas in 14 Sardinian red grape varieties the total extractable anthocyanin content ranged from 800 to 2 000 mg/kg (Vacca *et al.*, 2009). In this study, Vranac and Cabernet Sauvignon grapes proved to be highest in anthocyanins compared to Kratosija. In *Vitis vinifera* grape varieties, anthocyanins are located only in the skin of the grape berries, and their content in grapes

and wines is influenced by the size of the berry (Romero-Cascales *et al.*, 2005). Vranac and Kratosija grapes are almost twice the size of Cabernet Sauvignon grapes. The highest content of anthocyanins (Fig. 2) in terms of mg/kg skin FM was found in Vranac grapes (8 759 and 6 929 mg/kg skin FM in 2011 and 2012 respectively), followed by Cabernet Sauvignon (5 985 and 4 621 mg/kg skin FM) and Kratosija (3428 and 3 479 mg/kg skin FM) (Fig. 2). Such a result is a further confirmation that Vranac is a variety synthesising very high levels of anthocyanins.

The average extractable LMP content was the highest in Cabernet Sauvignon grapes (2 006 and 1 690 mg/kg in 2011 and 2012, respectively). Statistically lower contents of LMP were found in Vranac grapes (1 103 and 846 mg/kg in 2011 and 2012 respectively) and in the Kratosija grapes from 2012 (976 mg/kg). The LMP content in Kratosija grapes from 2011 (1 413 mg/kg) was not significantly different compared to the Cabernet Sauvignon grapes from 2011. Similarly, the average extractable HMP content was significantly higher in the Cabernet Sauvignon grapes (2 705 and 2 805 mg/kg in 2011 and 2012 respectively) compared to Vranac and Kratosija. The average extractable HMP content did not

TABLE 3

Content of extractable polyphenols (TP - total polyphenols, TA - total anthocyanins, LMP - low-molecular mass proanthocyanidins, HMP - high-molecular mass proanthocyanidins) in Vranac, Kratosija and Cabernet Sauvignon fresh grape berries.

Locality	2011				2012			
	TP (mg/kg (+) catechin)	TA (mg/kg)	LMP (mg/kg (+) catechin)	HMP (mg/kg cyanidin chloride)	TP (mg/kg (+) catechin)	TA (mg/kg)	LMP (mg/kg (+) catechin)	HMP (mg/kg cyanidin chloride)
VRANAC								
L1	1 505	1 149	774	785	1 854	1 075	946	1 778
L2	2 174	881	1 392	1 403	1 550	1 408	1 096	1 926
L3	2 353	1 136	1 785	1 716	1 843	1 166	874	1 791
L4	1 741	721	874	995	1 325	935	689	1 442
L5	2 139	999	1 067	1 564	1 480	1 177	642	1 395
L7	1 534	873	727	1 199	1 534	917	830	1 527
Mean	1 908 ab	960 a	1 103 b	1 277 b	1 598 ab	1 113 a	846 b	1 643 b
KRATOSIJA								
L1	1 988	450	1 726	2 072	852	417	901	1 197
L2	1 983	497	1 653	2 069	1 065	568	961	1 532
L3	2 038	505	1 862	1 368	1 049	565	878	1 469
L6	1 356	436	936	1 091	1 675	529	1 400	2 426
L9	1 128	394	888	2 148	843	506	740	1 114
Mean	1 699 b	456 b	1 413 ab	1 750 b	1 097 b	517 c	976 b	1 548 b
CABERNET SAUVIGNON								
L1	2 991	1 064	2 177	2 112	2 380	995	1 932	3 402
L2	2 241	852	1 611	2 585	1 875	981	1 860	3 356
L5	3 861	1 078	3 449	3 501	1 845	710	1 430	2 646
L6	3 241	1 411	1 972	3 562	1 841	825	2 000	3 550
L8	2 076	808	1 581	2 323	2 614	816	1 389	2 143
L10	1 819	994	1 244	2 146	1 544	836	1 528	1 734
Mean	2 705 a	1 035 a	2 006 a	2 705 a	2 017 a	861 b	1 690 a	2 805 a
sign. F	*	***	*	***	**	***	***	**

ANOVA was used to compare data (n.s. not significant, * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$). Different lower-case letters indicate significant differences of means between varieties using Tukey's HSD test ($p \leq 0.05$).

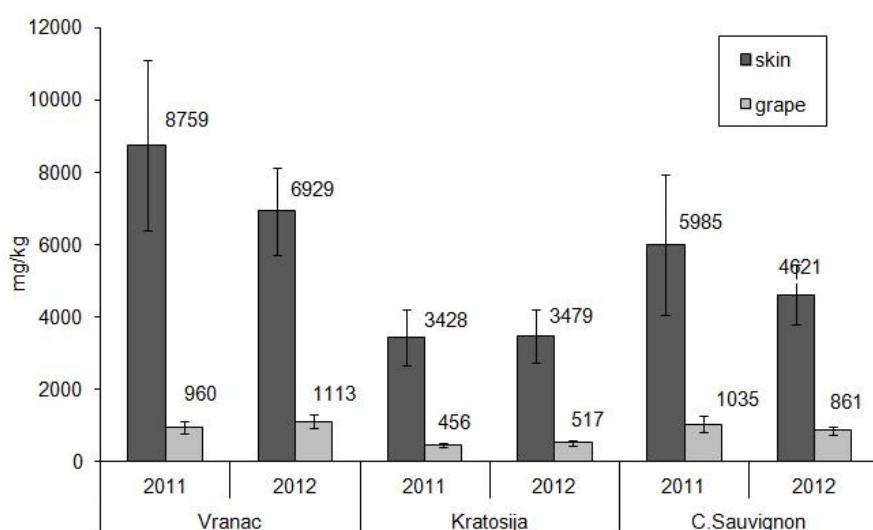


FIGURE 2

Content of extractable anthocyanins in the skin of grape berries (mg/kg skin fresh mass) and in the grape berries (mg/kg grape fresh mass) of the different varieties: Vranac ($n = 6$), Kratosija ($n = 5$) and Cabernet Sauvignon ($n = 6$). The error bars represent the standard deviation of the content from different locations.

differ significantly between the Vranac grapes (1 277 mg/kg and 1 643 mg/kg in 2011 and 2012 respectively) and the Kratosija grapes (1 750 mg/kg and 1 548 mg/kg in 2011 and 2012 respectively). Extractable HMP contents in Cabernet Sauvignon grapes were in agreement with results obtained by Mattivi *et al.* (2002a) and Vrhovsek *et al.* (2002), who reported contents of HMP ranging between 2 500 and 2 800 mg/kg and 2 300 to 2 700 mg/kg of grape FM, respectively.

The impact of the vineyard location on the polyphenol content in grapes

Differences in grape composition regarding extractable total polyphenols, anthocyanins, LMP and HMP contents were also found among vineyard locations in the Podgorica winegrowing area (Table 3), and similar outcomes were observed for the microvinified wines (Table 4). In the Podgorica winegrowing area, vineyards at locations L1 to L8 lie between 25 and 50 m above sea level (m.a.s.l.), whereas vineyards L9 (Kratosija) and L10 (Cabernet Sauvignon) lie between 250 and 400 m.a.s.l. (Fig. 1B). The

altitude of vineyards was found to have an impact on the levels of anthocyanins in the grapes, i.e. higher vineyard sites were reported to be advantageous to the biosynthesis of anthocyanins (Mateus *et al.*, 2002). Catechin monomers, procyanidin dimers, trimer C1 and total extractable proanthocyanidins proved to be higher in both the skins and seeds of grapes growing at lower altitudes (Mateus *et al.*, 2001). It was indeed found in both years that total polyphenols and LMP and HMP contents in Kratosija and Cabernet Sauvignon grapes from L9 and L10 respectively were the lowest among all investigated locations (except the LMP content in Kratosija in 2011), and similar outcomes were obtained in microvinified wines of Kratosija (Table 4). Vranac grapes originating from L2 and L3 proved to be high in the sum of extractable LMP and HMP contents for both 2011 and 2012 (Table 3). The extractable LMP and HMP contents in Kratosija grapes varied between the two years, i.e. from the highest content in L1 and the lowest content in L6 in 2011 to the lowest content in L1 and the highest content in L6 in 2012. The sum of extractable LMP and HMP

TABLE 4

Content of polyphenols (TP - Total polyphenols, TA - Total anthocyanins, LMP – low-molecular mass proanthocyanidins, HMP – high-molecular mass proanthocyanidins) in four-month-old wines from Vranac, Kratosija and Cabernet Sauvignon from the 2011 and 2012 vintages.

Locality	2011				2012			
	TP (mg/L (+) catechin)	TA (mg/L)	LMP (mg/L (+) catechin)	HMP (mg/L cyanidin chloride)	TP (mg/L (+) catechin)	TA (mg/L)	LMP (mg/L (+) catechin)	HMP (mg/L cyanidin chloride)
VRANAC								
L1	1 107	641	176	703	1 515	945	657	1 802
L2	1 000	474	173	1 405	1 650	1 107	581	2 093
L3	1 485	720	322	1 717	1 727	900	868	2 090
L4	1 146	734	281	995	885	617	350	913
L5	1 602	813	400	1 565	1 101	758	290	913
L7	891	559	246	896	859	664	363	921
Mean	1 205	657 b	266	1 214	1 290	832 a	518	1 455
KRATOSIJA								
L1	877	241	251	988	944	500	551	1 049
L2	937	268	292	860	1 015	458	579	994
L3	1 515	366	622	1 360	1 147	370	681	654
L9	837	394	272	889	736	387	414	686
Mean	1 042	317 c	359	1 024	961	429 b	556	846
CABERNET SAUVIGNON								
L1	1 085	964	350	1 020	1 461	514	963	1 924
L2	1 109	570	297	703	1 679	572	1 059	2 587
L5	1 387	1 018	349	1 302	1 367	484	516	788
L6	1 538	1 111	747	1 889	1 393	521	790	1 607
L8	1 388	1 063	670	1 471	1 065	512	607	916
Mean	1 301	945 a	483	1 277	1 393	521 b	787	1 564
sign. F	n.s.	***	n.s.	n.s.	n.s.	***	n.s.	n.s.

ANOVA was used to compare data (n.s. not significant, * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$). Different lower-case letters indicate significant differences of means between varieties using Tukey's HSD test ($p \leq 0.05$).

contents in Cabernet Sauvignon grapes was higher in L1, L2, L5 and L6 compared to L8 and L10 in both investigated years (Table 3). Grapes obtained from locations with high tannin levels can be used for a desired wine style, taking into consideration the winemaking protocol. However, at this stage it is too early to characterise vineyard locations based on polyphenol potential. Instead, a multiyear monitoring approach is needed for statistically relevant vineyard mapping.

The impact of seasonal variation (2011 and 2012) on the extractable polyphenols in grapes at different vineyard locations is evident. Climatic conditions play an important role in polyphenol biosynthesis in grapes, resulting in markedly higher levels of polyphenol contents in grapes grown in locations or areas with higher average daytime temperatures and decreased precipitation (Lee *et al.*, 2009). The total polyphenol content in grapes proved to be higher in 2011, which was characterised by high vegetation temperatures and lower precipitation compared to 2012 for all three investigated varieties.

Distribution of low- and high-molecular mass proanthocyanidins between the skin and seeds of the grape berries

The distribution of proanthocyanidins between the seeds and skins of grape berries is known to be affected by grape variety (Mattivi *et al.*, 2002b, 2009), although the genetic control of the qualitative and quantitative proanthocyanidin composition between berry skin and seeds is complex and still poorly understood (Huang *et al.*, 2012). Fig. 3 represents the two-year average distribution of extractable LMP and HMP between grape berry skins and seeds for the varieties Vranac, Kratosija and Cabernet Sauvignon. The two-year average distribution of LMP between skins and seeds was 34:66, 39:61 and 49:51, whereas the distribution of HMP between skins and seeds was 67:33, 62:38 and 64:36 for Vranac, Kratosija and Cabernet Sauvignon respectively

(Fig. 3). Vrhovsek *et al.* (2002), Mattivi *et al.* (2002a) and Vacca *et al.* (2009) reported similar results regarding extractable proanthocyanidin distribution in different red *Vitis vinifera* grapes, with increased levels of LMP in the seeds and increased levels of HMP in the skins of the grape berries.

Contents of total polyphenols, low- and high-molecular mass proanthocyanidins and total anthocyanins in Vranac, Kratosija and Cabernet Sauvignon wines

Contents of total polyphenols, low- and high-molecular mass proanthocyanidins and total anthocyanins in four-months-old Vranac, Kratosija and Cabernet Sauvignon wines from different vineyard locations in Podgorica district in 2011 and 2012 are shown in Table 4.

On average, the Cabernet Sauvignon wines had a higher content of total polyphenols in both years investigated (1 301 and 1 393 mg/L in 2011 and 2012, respectively) compared to the Vranac wines (1 205 and 1 290 mg/L) and Kratosija wines (1 042 and 961 mg/L), although the differences were not statistically significant. The anthocyanin content was significantly higher in the Cabernet Sauvignon wines from 2011 (945 mg/L) compared to the Vranac (657 mg/L) and Kratosija wines (317 mg/L). Vranac wines made during 2012 proved to be significantly higher in anthocyanin content (832 mg/L) compared to the Cabernet Sauvignon (521 mg/L) and Kratosija wines (429 mg/L). Kratosija wines showed the lowest anthocyanin content in both 2011 and 2012. The mean LMP content was also the highest for the Cabernet Sauvignon wines (483 and 787 mg/L in 2011 and 2012, respectively). Lower contents, although not statistically significant, were found in Kratosija wines (359 and 556 mg/L in 2011 and 2012, respectively) and in Vranac wines (266 and 518 mg/L in 2011 and 2012 respectively). The average HMP content was the highest in Cabernet Sauvignon wines (1 277 and 1 564 mg/L in 2011 and 2012 respectively), followed by Vranac wines (1 214 mg/L and 1 455 mg/L in 2011 and 2012,

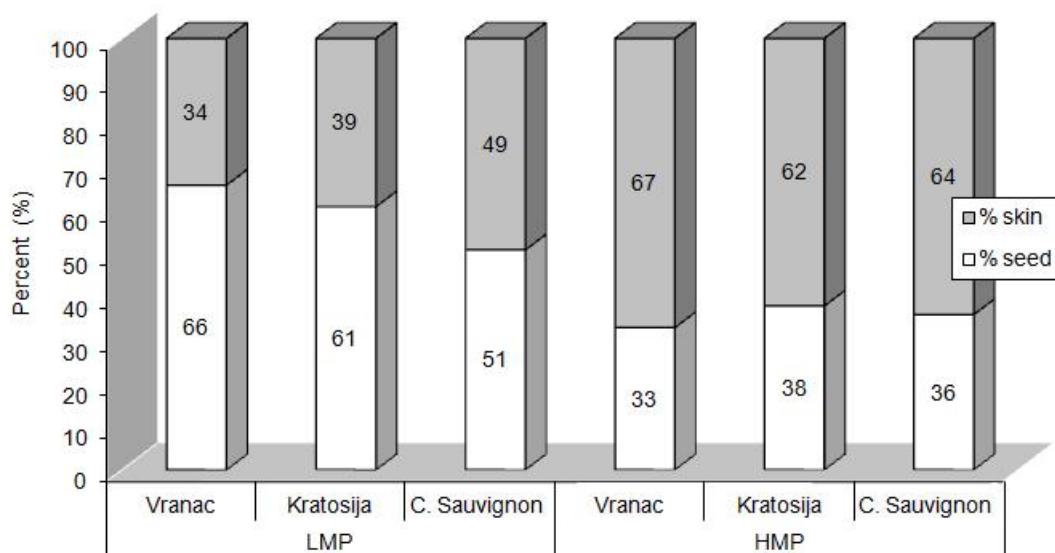


FIGURE 3

Distribution of extractable low-molecular mass (LMP) and high-molecular mass (HMP) proanthocyanidins between skins and seeds of Vranac, Kratosija and Cabernet Sauvignon grapes.

respectively) and Kratosija wines (1 024 and 846 mg/L in 2011 and 2012, respectively). In both years, the trend in the polyphenol contents determined in wines was correlated well with the contents found in the grapes (Table 3). However, lower contents of LMP were found in wines than in grape extracts, thus concluding that the extraction of LMP was less efficient in microvinification than in the grape extraction process. The reason for this could be that the yeasts used were not standardised, the press fraction was missing or the maceration time was too short to allow for a proper extraction of polyphenols from the grape seeds.

CONCLUSIONS

Polyphenol compounds in red wine are directly linked to eventual wine flavour, colour and ageing characteristics. The evaluation of polyphenol compounds in grapes and wines of Montenegrin red grape varieties provided a technological characterisation that can be used by both winegrowers and winemakers to develop proper programmes for phenolic management. Cabernet Sauvignon grapes proved to be highest in extractable total polyphenols, anthocyanins (Vranac grapes showed increased levels), and low- and high-molecular mass proanthocyanidins during 2011 and 2012. The same trend was also found in the microvinified wines. Vranac grapes showed lower polyphenol potential in comparison to Cabernet Sauvignon, but a high anthocyanin content that gave the wines an intense red colour. The grapes of Vranac showed the highest anthocyanin content in the grape skins in comparison to Cabernet Sauvignon and Kratosija. However, the larger berry size (almost twice) of Vranac and Kratosija in comparison to Cabernet Sauvignon dilutes the content of polyphenols in the grapes. Vranac could be considered as a variety with medium to long ageing potential, although the optimal ripeness of the grapes and suitable winemaking procedures have to be considered. Kratosija grapes had the highest TSS content at the time of sampling and a similar content of extractable total polyphenols, LMP and HMP as for Vranac. The main difference between Vranac and Kratosija was the lower content of anthocyanins in both the grapes and wines of Kratosija compared to Vranac. Kratosija can be considered a variety with lower polyphenol potential compared to Vranac and Cabernet Sauvignon.

As regards Vranac and Kratosija grapes, a strange relationship was found between sugar and polyphenol accumulation. Further studies (cluster thinning, canopy to crop rate) therefore are needed in order to better understand the best equilibrium for these varieties. As regards Cabernet Sauvignon grapes, it would be good to perform cluster thinning in colder seasons in order to obtain better maturation. It would also be profitable to take care of water stress in hot seasons to get better maturation.

Considering the vineyard locations, it could be postulated that increased levels of polyphenols were present in grapes originating from locations between 25 m and 50 m. a. s. l. compared to those at 250 m and 400 m. a. s. l.. However, only two locations out of the nine lie at higher altitudes. Furthermore, it was found that climatic conditions during 2011 and 2012 had an impact on polyphenol biosynthesis in grapes from different vineyard locations. Two-year monitoring of total extractable tannins in grapes from

different locations led to the identification of locations with improved grape tannin potential: L2 and L3 for Vranac and L5 and L6 for Cabernet Sauvignon. No conclusion can be drawn regarding the grape polyphenol potential of the locations for the Kratosija variety. A multiyear study therefore will be carried out in the future to evaluate the polyphenol potential of grapes from different vineyard locations.

LITERATURE CITED

- Avramov, L., 1991. Vinogradarstvo. Nolit, Beograd.
- Bozinovic, Z., 2005. Ampelografija. Akademik. Skopje.
- Broadhurst, R.B. & Jones, W.T., 1978. Analysis of condensed tannins using acidified vanillin. *J. Sci. Food Agric.* 29, 788-794.
- Brossaud, F., Cheynier, V. & Noble, A.C., 2001. Bitterness and astringency of grape and wine polyphenols. *Aus. J. Grape Wine Res.* 7, 33-39.
- Chira, K., Schmauch, G., Saucier, C., Fabre, S. & Teissedre, P.L., 2009. Grape variety effect on proanthocyanidin composition and sensory perception of skin and seed tannin extracts from Bordeaux wine grapes (Cabernet Sauvignon and Merlot) for two consecutive vintages (2006 and 2007). *J. Agric. Food Chem.* 57 (2), 545-553.
- Di Stefano, R. & Guidoni, S., 1989. The analysis of total polyphenols in musts and wines. *Vigne Vini* 1/2, 47-52.
- Di Stefano, R., Cravero, M.C. & Gentilini, N., 1989. Methods for the study of wine polyphenols. *L'Enotecnico* 5, 83-89.
- Gambuti, A., Capuano, R., Lecce, L., Fragasso, M.G. & Moio, L., 2009. Extraction of phenolic compounds from 'Aglianico' And 'Uva di Troia' grape skin and seeds in model solutions: Influence of ethanol and maceration time. *Vitis* 48, (4), 193-200.
- Huang, Y.-F., Doligez, A., Fournier-Level, A., Le Cunff, L., Bertrand, Y., Canaguier, A., Morel, C., Miralles, V., Veran, F., Souquet, J.-M., Cheynier, V., Terrier, N. & This, P., 2012. Dissecting genetic architecture of grape proanthocyanidin composition through quantitative trait locus mapping. *BMC Plant Biology* 12, 30.
- Ivanova, V., Stefova, M. & Chinnici, F., 2010. Determination of the polyphenol contents in Macedonian grapes and wines by standardized spectrophotometric methods. *J. Serb. Chem. Soc.* 75, 45-59.
- Ivanova, V., Stefova, M., Vojnoski, B., Dörnyei, Á., Márk, L., Dimovska, V., Stafilov, T. & Kilár, F., 2011. Identification of polyphenolic compounds in red and white grape varieties grown in R. Macedonia and changes of their content during ripening. *Food Res. Int.* 44, 2851-2860.
- Lee, J.-E., Hwang, G.-S., Van den Berg, F., Lee, C.-H. & Hong, Y.-S., 2009. Evidence of vintage effects on grape wines using ¹H NMR-based metabolomic study. *Anal. Chim. Acta* 648, 71-76.
- Mateus, N., Machado, J.M. & De Freitas, V., 2002. Development changes of anthocyanins in *Vitis vinifera* grapes grown in the Douro Valley and concentration in respective wines. *J. Sci. Food Agric.* 82, 1689-1695.
- Mateus, N., Marques, S., Gonçalves, A.C., Machado, J.-M. & De Freitas, V., 2001. Proanthocyanidin composition of red *Vitis vinifera* varieties from the Douro Valley during ripening: Influence of cultivation altitude. *Am. J. Enol. Vitic.* 52, 115-121.
- Mattivi, F., Prast, A., Nicolini, G. & Valenti, L., 2002b. Validazione di un nuovo metodo per la misura del potenziale polifenolico delle uve rosse e discussione del suo campo di applicazione in enologia. *Riv. Vitic. Enol.* 2/3, 55-74.
- Mattivi, F., Vrhovsek, U., Masuero, D. & Trainotti, D., 2009. Differences in the amount and structure of extractable skin and seed tannins amongst red grape varieties. *Aus. J. Grape Wine Res.* 15, 27-35.

- Mattivi, F., Julian, C., Nicolini, G. & Valenti, L., 2002a. Wine, biodiversity, technology, and antioxidants. *Ann. N.Y. Acad. Sci.* 957, 37-56.
- MONSTAT - Statistical Office of Montenegro, 2013. Statistical Yearbook. MONSTAT, Podgorica.
- OIV (International Organisation of Vine and Wine), 2011. Compendium of international methods of wine and must analysis, vol 1. International Organisation of Vine and Wine, Paris.
- Pajovic, R., Popovic, T., Boskov, K. & Beleski, K., 2009. Technological characteristics of introducing varieties Merlot and Cabernet Sauvignon in the Podgorica wine growing region (Montenegro) and Skopje wine growing region (Macedonia). *Agroznanje* 10(1), 89-96.
- Pajovic, R., Popovic, T., Mijovic, S. & Vukoslavljevic, V., 2011. Economic, technological and polyphenolic characteristics of Vranac, Kratosija and Merlot grapes varieties and wines in Montenegro and Macedonia. In: Blesic, M. (ed). Proc. 22nd International Scientific-Expert Conference on Agriculture and Food Industry, 2011, Neum, Bosnia and Herzegovina. pp. 341 – 345.
- Pejovic, L., 1988. Ampelografska proučavanja varijeteta Kratosije. [Ampelographic investigations of Kratosija biotypes]. Jugoslovensko vinogradarstvo i vinarstvo, Beograd, 34, 21-23.
- Peleg, H., Gacon, K., Schlich, P. & Noble, A.C., 1999. Bitterness and astringency of flavan 3-ol monomers, dimers and trimers. *J. Sci. Food Agric.* 79, 1123-1128.
- Peyrot des Gachons, C. & Kennedy, J.A., 2003. Direct method for determining seed and skin proanthocyanidin extraction in red wine. *J. Agric. Food Chem.* 51, 5877-5881.
- Preys, S., Mazerolles, G., Courcoux, P., Samson, A., Fischer, U., Hanafi, M., Bertrand, D. & Cheynier, V., 2006. Relationship between polyphenolic composition and some sensory properties in red wine using multi way analyses. *Anal. Chim. Acta* 563, 126-136.
- Ribéreau-Gayon, P., 1982. The anthocyanins of grapes and wine. In: Markakis P. (ed). Anthocyanins as food colors. Academic Press, New York. pp. 112 – 118.
- Rigo, A., Vianello, F., Clementi, G., Rossetto, M., Scarpa, M., Vrhovsek, U. & Mattivi, F., 2000. Contribution of proanthocyanidins to the peroxy radical scavenging capacity of some Italian red wines. *J. Agric. Food Chem.* 48, 1996-2002.
- Robichaud, J.L. & Noble, A.C., 1990. Astringency and bitterness of the selected phenolics in wine. *J. Sci. Food Agric.* 53, 343-353.
- Rodrigo, R., Miranda, A. & Vergara, L., 2011. Modulation of endogenous antioxidant system by wine polyphenols in human disease. *Clin. Chim. Acta* 412, 410-424.
- Romero-Cascales, I., Ortega-Regules, A., López-Roca, J., Fernández-Fernández, J. & Gómez-Plaza, E., 2005. Differences in anthocyanin extractability from grapes to wines according to variety. *Am. J. Enol. Vitic.* 56, 212-219.
- Somers, T.C., 1971. The polymeric nature of wine pigments. *Phytochemistry* 10, 2175-2186.
- Ulicevic, M., 1966. Prilog proučavanju osobina najvažnijih sorata vinove loze gajenih u SR Crnoj Gori. [Contribution to characterization of the most important grapevine varieties grown in Republic of Montenegro]. *Arhiv za poljoprivredne nauke* 23, 1-100.
- Vacca, V., Del Caro, A., Millela, G.G. & Nieddu, G., 2009. Preliminary characterisation of Sardinian red grape cultivars (*Vitis vinifera* L.) according to their phenolic potential. *S. Afr. J. Enol. Vitic.* 30, (2), 93-100.
- Vrhovsek, U., Mattivi, F. & Waterhouse, A.L., 2001. Analysis of red wine phenolics: comparison of HPLC and spectrophotometric methods. *Vitis* 40(2), 87-91.
- Vrhovsek, U., Vanzo, A., Koruza, B. & Korosec-Koruza, Z., 2002. Polifenolni potencial slovenskega rdečega grozja. [Polyphenol potential of Slovenian red grapes]. In: Puconja, M. (ed). *Vinogradi in vina za tretje tisočletje*. Otocec, Slovenia. pp. 359 – 367.