

The abundance of *Cryptosporidium* and *Giardia* spp. in treated effluents produced by four wastewater treatment plants in the Gauteng Province of South Africa

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Abstract

This study aimed at assessing the effectiveness of 4 wastewater treatment plants of the Gauteng Province, namely Zeekoegat, Bavianspoort, Rayton and Refilwe Water Care Works (WCW), in the removal of *Cryptosporidium* and *Giardia* (oo)cysts. Wastewater influent and treated effluent samples were taken weekly between January and April 2008. *Cryptosporidium* and *Giardia* (oo)cysts were detected by immunofluorescence and immunomagnetic separation, according to a modified US EPA 1623 method. Effluent samples were subjected to a molecular study for the identification of *Cryptosporidium parvum* Genotype I and *Giardia lamblia* Assemblage A. The 18S rRNA gene for restriction digests was therefore used to characterise these (oo)cysts. *Cryptosporidium* oocysts were repeatedly detected in effluent samples collected from all wastewater treatments at lower concentration (range <1 to 40 oocysts/L) levels than *Giardia* cysts (range <1 to 175 cysts/l). The mean removal efficiencies of *Cryptosporidium* and *Giardia* at the 4 wastewater treatment plants ranged from 67.40% to 98.26% and from 86.81% to 99.96%, respectively. For all effluent samples, except Zeekoegat WCW, 29% and 41% contained oocysts of *Cryptosporidium parvum* Genotype I and cysts of *Giardia lamblia* Assemblage A, respectively. Both *C. Parvum* and *G. lamblia* are human pathogens. This stresses the potential risk of discharging these parasites into receiving water bodies.

Keywords: wastewater, removal, *Cryptosporidium parvum*, *Giardia lamblia*, genotype

Introduction

The world's freshwater reserves are limited and freshwater is obtained through the water cycle. The limited water reserves are declining year after year and freshwater is becoming a scarce commodity. Therefore, the recycling of treated wastewater is essential for reducing the negative impact of pollution on freshwater reserves. As South Africa is a water-scarce country, return flows (sewage and effluent purification) occupy second position in terms of the size of their contribution (14%) to the total supply of freshwater in South Africa, with 77% sourced from surface water (dams and rivers) and 9% from groundwater (boreholes) (Van Vuuren, 2009).

Uncontrolled sewage discharges and poorly managed wastewater treatment plants have been identified as two of the major sources of water pollution in South Africa (Momba and Mfenyana, 2005; Momba and Sibewu, 2009). Treated sewage is often discharged directly into watercourses that also serve as sources of drinking water or which have recreational sites downstream. River water has been identified as a significant source of *Cryptosporidium* and other pathogens (Momba et al., 2006; Clarke et al., 2008; Pennil et al., 2008).

Parasites belonging to the genera *Cryptosporidium* and *Giardia* are an important and widespread cause of enteric diseases in humans and in many other vertebrates (WHO, 2004). The transmission of protozoan parasites occurs mainly through

the faecal-oral route by exposure to contaminated water and food (Rose et al., 2002). Several studies have shown that water is a source of various waterborne infectious diseases affecting numerous communities, particularly those in rural and indigenous areas (Venter, 2000; Murcott, 2006; Momba et al., 2008), and consequently an estimated 5 million people lose their lives due to water-related diseases each year (Pritchard et al., 2009; Baumgartner et al., 2007). Outbreaks of cryptosporidiosis in many communities in the past have been reported even when the bacterial quality of the water complied with the set standards after treatment (Insulander et al., 2005).

Giardiasis and cryptosporidiosis are common infections of domestic and wild animals, which shed a large number of cysts and oocysts into the environment (WHO, 2004). A study by Jarmy-Swan and co-workers on the prevalence of diarrhoeal disease caused by waterborne pathogens indicates that *Cryptosporium* and *Giardia* are endemic in the KwaZulu-Natal population, with prevalence rates of 39.3% in the < 1-year age group and 38.5% in the 3- to 4-year age group (Jarmey-Swan et al., 2001). The occurrence of cryptosporidiosis and giardiasis is probably higher than recorded in this province. In 1998, Pegram and co-workers reported that only 1 in 14 people with diarrhoea in South Africa seeks formal treatment from a health practitioner, clinic or hospital every year, while approximately 43 000 South Africans have been estimated to die from diarrhoea every year. Thus, producing effluent of a high microbiological quality remains a matter of great concern in developing countries such as South Africa.

It has been reported that bacteria such as *E. coli*, *Salmonella typhimurium* and *Vibrio cholera* and viral indicators, once entrapped in suspended flocs, appeared to be able to survive in the effluent despite high residual chlorine

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concentrations. However, the removal of these microorganisms by sedimentation and filtration is a major prerequisite for plants that have demonstrated successful disinfection efficiency (Momba et al., 2006).

A serious concern is that the receiving waterbodies, which can also be contaminated with pathogenic protozoa, are used by communities for drinking, recreational and agricultural purposes directly or indirectly without prior treatment. The (oo)cysts are insensitive to disinfectants at concentrations commonly used in water treatment plants to reduce bacterial contamination (Mahdy et al., 2007). As the recent study revealed the failure of South African wastewater treatment plants to remove pathogenic bacteria and viral indicators, it was also crucial to assess the effectiveness of Zeekoegat, Bavianspoort, Rayton and Refilwe wastewater treatment plants in the removal of *Cryptosporidium* and *Giardia* (oo)cysts from their influents. Monitoring the quality of wastewater effluents before their discharge into the receiving water body might therefore assist water services providers in tracing the origin of water source contaminations which can result in a diarrhoeal epidemic in the Gauteng Province.

Materials and methods

Study sites

The study was conducted at 4 wastewater treatment plants in the Gauteng Province of South Africa. All these plants treat municipal wastewaters using the activated sludge system. Zeekoegat and Bavianspoort water care works (WCW) are 2 of several wastewater treatment plants that serve the City of Tshwane Metropolitan Municipality (hereafter referred to simply as Tshwane). Both plants are located approximately 40 km east of Pretoria (which forms part of the Tshwane municipal area). These 2 plants were studied because they drain into the Roodeplaat Dam, which serves as a source of water for several drinking water treatment plants in the area. It is also a recreational area.

The Zeekoegat WCW is located immediately to the west of Roodeplaat Dam. This plant has a design capacity of 35 Ml/day, which is upgradeable to 120 Ml/day. At the Zeekoegat WCW, the influent flows into primary settling tanks and then to a division tank that enables the process controllers to maintain a constant flow rate to the 2 biological reactors despite changes in influent flow rates. The biological reactors are characterised by automated submerged diffusion-air aeration systems. The effluent from the biological reactors flows to the secondary settling tanks where clarification takes place. The overflow from the settlers flows through sand filters and finally to an emergency dam where a prolonged contact time with chlorine is achieved. The treated effluent flows into the Roodeplaat Dam via a short earth canal that passes through the Roodeplaat Dam Nature Reserve.

Bavianspoort WCW has a capacity of approximately 35 Ml/day. The set-up of Bavianspoort WCW is similar to that of Zeekoegat WCW. The main difference is in the design of the plant, which is characterised by the lack of a division tank, the presence of a mechanical aeration system in the 2 biological reactors and the absence of sand filtration of the final treated effluent. This plant is located 10 km upstream of the Roodeplaat Dam on the eastern bank of the Pienaars River.

The Rayton and Refilwe WCWs are located in the small towns of Rayton and Cullinan, respectively, and serve their respective towns. These towns are also approximately 45 km

east of Pretoria. The populations, according to the Statistics South Africa 2001 census of Cullinan and Rayton, were 7 683 and 2 961, respectively (STATS SA, 2001). The design of these plants is simpler than those of the first 2 plants. The influent flows through a grit screen and then into a single biological reactor tank. The effluent of the biological reactor flows to the settling tank and the overflow of the settler flows to the contact tank. After the contact tank, no further treatment is done. The effluent is discharged into the receiving water bodies. The design capacities of the Rayton and Refilwe WCWs are 0.6 and 1.1 Ml/day, respectively. Because of their small size, they were included in this study to compare their performance with that of the larger plants.

Sample collection and preparation

Samples of untreated wastewater influents (2 l) and treated wastewater final effluents (10 l) were collected weekly between January and April 2008. The water samples were filtered through a 300 µm 50-mesh sieve to remove large particles and then concentrated by filtration on cellulose-acetate filters of 0.8 µm pore size and 142 mm diameter (Sartorius, RSA). The filters were scraped and washed using 50 ml 0.1% Tween 80 followed by centrifugation at 2000 x g to pellet the (oo)cysts. The supernatants were aspirated to 10 ml above the pellet according to the US EPA 1623 method (US EPA, 2001).

Immunomagnetic separation and staining

The cysts and oocysts were captured from the remaining 10 ml of the supernatant using Dynalbead anti-*Giardia* and anti-*Cryptosporidium* immunomagnetic antibodies (DEHTEQ, RSA). A 50 µl aliquot of the purified suspension containing the captured oocysts was air-dried on a well-slide and stained with anti-*G. lamblia* and anti-*C. parvum* monoclonal antibodies conjugated to fluorescein isothiocyanate (FITC) (Aqua-Glo G/C Kit, Invitrogen USA). The slides were examined at 1000 x magnification using an Axio Carl Zeiss epifluorescence microscope (Carl Zeiss, RSA). *Giardia* cysts were identified based on their size, shape, and the pattern and intensity of immunofluorescent assay staining (bright green fluorescence of the cyst wall). *Cryptosporidium* oocysts were identified based on their size, shape, and the presence of a suture on the oocyst wall at 1000 x magnification. The number of (oo)cysts was counted in duplicate for each sample according to the US EPA 1623 method (2001).

DNA extraction and PCR amplification

The polymerase chain reaction (PCR) was used to confirm the identities of the presumptive oocysts. DNA extraction was performed according to the method of Sturbaum and co-workers (2001). A 50 µl suspension was incubated with 1 mg of proteinase K per ml at 56°C for at least 1 h and subjected to 6 freeze-thaw cycles (2 min in liquid nitrogen, followed by 2 min in a 98°C water bath), with the centrifugation (16 000 x g) performed between the third and fourth cycles for 30 s. Lastly, the 30 µl suspension was centrifuged at 16000 x g for 3 min. One 10 µl aliquot of the freeze-thaw supernatant was used directly as the PCR DNA template.

The PCR primers and conditions used in this study for *Cryptosporidium* and *Giardia* have been fully described by Sturbaum et al. (2001) and Cacció (2003) (Table 1). The nested PCR primers for *C. parvum* amplified a

Target organism	Primers name	Sequence 5'-3'	Target	References
<i>C. parvum</i>	ExCry 1 ExCry 2 NesCry 3 NesCry 4	GCC AGT AGT CAT ATG CTT GTC TC ACT GTT AAA TAG AAA TGC CCC C GCG AAA AAA CTC GAC TTT ATG GAA GGG GGA GTA TTC AAG GCA TAT GCC TGC	590-bp of 18S rRNA	Sturbaum et al., (2001) Reed et al., (2001)
<i>G. lamblia</i>	G7 G759	AAG CCC GAC GAC CTC ACC CGC AGT GC GAG GCC GCC CTG GAT CTT CGA GAC GAC	753-bp of β -giardin gene	Cacciò et al., (2003)

Restriction enzyme	Target organism	Conditions	Products	Reference
<i>VspI</i>	<i>C. parvum</i> genotype I and II	In 20- μ l reaction volumes, 15 μ l of amplicon is digested with 5 U of <i>VspI</i> or 4 U of <i>DraII</i> for 6 h at 37°C	Digestion of 590 bp product for only genotype I.	Sturbaum et al., 2001 Reed et al., 2001
<i>DraII</i>	<i>C. baileyi</i> and <i>C. serpentis</i>			
<i>HaeIII</i>	<i>Giardia lamblia</i> Assemblage A and B	4 h digestion at 37°C with 10 U of <i>HaeIII</i> in a final volume of 20 μ l.	Fragments of 202, 201, 150, 126, and 74 bp for Assemblage A, and fragments of 202, 176, 150, 117, 84, and 24 bp for Assemblage B	Cacciò et al., 2003 Cacciò et al., 2002

590-bp region of the 18S rRNA gene of Genotype I and II. An initial denaturation step was performed at 95°C for 5 min and a 10 min holding step at 80°C; this was followed by 40 cycles at 94°C for 45 s, 53°C for 1 min 16 s, and 72°C for 45 s. Final extension was carried out at 72°C for 7 min. The nested-reaction parameters were the same except that the 35 cycles were performed at an annealing temperature of 65°C and denaturation, annealing, and extension time periods were 25 s each. (Sturbaum et al., 2001). The PCR primers for *G. lamblia* amplified a 753-bp region of the β -giardin gene of Assemblage A and B. An initial denaturation of 5 min at 94°C was used, followed by a set of 35 to 40 cycles, each consisting of 30 s at 94°C, 30 s at 65°C, and 1 min at 72°C. A final extension of 5 min at 72°C was performed (Cacciò et al., 2003). The PCR reactions were performed in a Chromo 4 thermal cyler (BIORAD, RSA). Following PCR, 15 μ l of the *Cryptosporidium* nested amplicon was digested with 5 U of *VspI* (Promega) for 6 h at 37°C (Table 2) (Sturbaum et al., 2001). Similarly, the *Giardia* PCR products were treated with 10 U of *HaeIII* (Promega) for 4 h at 37°C as shown in Table 2 (Cacciò et al., 2002; 2003). The digested products were visualised on 2% agarose gels stained with ethidium bromide (0.5 μ g/ μ l).

Statistical analysis

To show the efficiency of the 4 wastewater treatment plants in removing pathogenic protozoa, the data were analysed statistically using the SPSS computer software, Version 11.0. A descriptive analysis was performed with interest in the kurtosis and skewness of the data. With 75% of the kurtosis and skewness between -2 and +2, the data proved to be of normal distribution hence parametric tests, which have assumptions of normality, such as comparison of means using the one-way analysis of variance (ANOVA) and test of relationship were carried out using the Pearson correlation index at 95% confidence interval.

Results

In general, *Cryptosporidium* oocysts and *Giardia* cysts were detected in influents and effluents from all the plants during the study period. Compared to the counts of *Giardia* cysts, lower counts for *Cryptosporidium* oocysts were observed in both the influent and effluent samples. While these parasites were always present in the effluents collected from the Refilwe WCW, they could also be detected from time to time at the Baviaanspoort, Zeekoegat and Rayton WCWs.

Table 3 summarises the number of oocysts and cysts detected from influent and effluent samples of the Zeekoegat and Baviaanspoort WCWs. At the Zeekoegat WCW, the counts of oocysts ranged between 0 and 150 oocysts/l in the influents and between 0 and 2 oocysts/l in the effluent samples. In the same plant, the counts for cysts ranged between 0 and 4 500 cysts/l in the influents and between 0 and 1 cysts/l in the effluents. *Cryptosporidium* oocysts were isolated in 50% of the influent samples and in 43% of the effluent samples, while *Giardia* was observed in 78% of all influent samples and 36% of effluent samples (Table 3). At the Baviaanspoort WCW, the counts of oocysts ranged between 0 and 76 oocysts/l in the influent samples and between 0 and 35 oocysts/l in the effluent samples. For the same plant, the cyst counts ranged between 0 and 4 300 cysts/l in the influents and between 0 and 27 cysts/l in the effluent samples. In this plant, *Cryptosporidium* occurred in 57% of the influent samples and in 62% of the effluent samples, while *Giardia* was observed in 93% of all influent samples and 86% of effluent samples (Table 3).

The number of oocysts and cysts observed per litre in the influent and effluent samples collected at the Rayton and Refilwe WCWs is shown in Table 4. At the Rayton WCW, the counts of oocysts ranged between 0 and 130 oocysts/l in the influents and between 0 and 15 oocysts/l in the effluent samples. The cyst counts for the same plant ranged between 0 and 210 cysts/l in the influents and between 0 and 24 cysts/l in the effluents. *Cryptosporidium* oocysts were observed in 57% of

Date in 2008	Zeekoegat				Baviaanspoort			
	Influent		Effluent		Influent		Effluent	
	Oocysts	Cysts	Oocysts	Cysts	Oocysts	Cysts	Oocysts	Cysts
29 Jan	145	450	2	1	42	3 400	10	10
7 Feb	0	123	0	0	0	3 200	0	15
13 Feb	58	450	0	0	45	2 400	12	0
20 Feb	46	25	1	1	0	4 300	13	12
27 Feb	0	0	0	0	51	820	12	13
5 Mar	0	45	1	0	0	560	0	0
12 Mar	48	2 100	0	1	26	1 200	0	2
19 Mar	0	0	0	1	0	4 200	0	4
26 Mar	53	510	2	0	0	3 100	2	16
2 Apr	0	0	0	1	76	570	10	1
11 Apr	16	4 500	2	0	0	0	6	4
16 Apr	0	3 200	1	0	45	1 500	35	5
23 Apr	150	15	0	0	10	43	0	27
30 Apr	0	7	0	0	67	163	1	9
Mean	37	816	1	0	26	1 818	7	8
Removal efficiency (%)			98.26	99.96			67.40	99.60

Date in 2008	Rayton				Refilwe			
	Influent		Effluent		Influent		Effluent	
	Oocysts	Cysts	Oocysts	Cysts	Oocysts	Cysts	Oocysts	Cysts
29 Jan	5	120	2	24	74	130	12	26
7 Feb	43	3	3	3	46	410	9	35
13 Feb	0	57	5	18	53	2 900	13	52
20 Feb	48	0	0	0	0	370	1	26
27 Feb	41	0	3	0	54	150	11	41
5 Mar	0	63	2	12	42	400	25	11
12 Mar	0	150	6	0	0	2 300	40	35
19 Mar	130	0	4	13	101	0	12	31
26 Mar	0	210	0	0	52	520	0	3
2 Apr	0	0	1	2	46	1 200	24	175
11 Apr	17	0	5	0	0	46	0	15
16 Apr	5	45	0	6	86	45	2	4
23 Apr	2	3	15	8	53	2	23	7
30 Apr	0	1	2	0	1	16	5	33
Mean	21	47	3	6	43	606	13	35
Removal efficiency (%)			83.51	86.81			70.89	94.18

the influent samples and in 79% of the effluent samples, while *Giardia* cysts were observed in 64% of the influent samples and 57% of the effluent samples. At the Refilwe WCW, the counts of oocysts ranged between 0 and 101 oocysts/l in the influent samples and between 0 and 40 oocysts/l in the effluent samples. In the same plant, the counts for cysts ranged between 0 and 2 900 cysts/l in the influent samples and between 3 and 175 cysts/l in the effluent samples. *Cryptosporidium* oocysts were observed in 79% of the influent samples and in 86% of the

effluent sample, whilst *Giardia* cysts were observed in 93% of all influent samples and in all of the effluent samples (Table 4).

Comparing the larger plants (Zeekoegat and Baviaanspoort) with the smaller plants (Refilwe and Rayton), it was noted that there was no significant differences in the number of oocysts and cysts isolated from the influent samples ($p \geq 0.05$). However, with respect to the number of cysts in the influents and effluents, the Zeekoegat WCW had significantly lower counts than the Baviaanspoort WCW ($p \leq 0.05$). Significant differences

Plant	<i>Cryptosporidium parvum</i>		<i>Giardia lamblia</i>	
	Genotype I	Genotype II	Assemblage A	Assemblage B
Baviaanspoort	4/14 (29%)	3/14 (22%)	5/14 (36%)	1/14 (7%)
Zeekoegat	0	0	0	0
Rayton	1/14 (7%)	0	4/14 (29%)	2/14 (14%)
Refilwe	7/14 (50%)	4/14 (29%)	9/14 (62%)	8/14 (57%)

in the number of oocysts and cysts isolated from the effluent samples ($p \leq 0.05$) were also noted between the Rayton and Refilwe WCWs. The counts for *Cryptosporidium* oocysts at the Refilwe WCW's effluents were found to be significantly higher ($p \leq 0.05$) than those of Baviaanspoort and Zeekoegat. Furthermore, it was observed that the counts of *Giardia* cysts in the effluent samples of the Refilwe WCW were significantly higher ($p \leq 0.05$) than those at the rest of the plants.

With the exception of the effluent samples collected from the Zeekoegat WCW, which showed the absence of the targeted genotypes of the parasites from all samples collected during the study period, the PCR reactions resulted in the positive identification of *Cryptosporidium parvum* **Genotype I** and *Giardia lamblia* **Assemblage A** at 29% and 36% for Baviaanspoort WCW, 7% and 29% for Rayton WRC, and 50% and 62% for Refilwe WCW, respectively (Table 5). Overall, pathogens capable of interspecies infection, *Cryptosporidium parvum* **Genotype II** and *Giardia lamblia* **Assemblage B**, were observed at lower occurrences than those that are infectious to humans only (Table 5).

Discussion

The transmission of *Giardia* and *Cryptosporidium* spp. through contaminated water, food and mainly the oral-faecal route to humans is well documented. The cycling of these pathogens can be between humans and also between animals and humans (Thompson, 2000; Cacció et al., 2003). Because the infected individuals pass very large numbers of oocysts and cysts with their faeces, this increases the environmental contamination. The situation is worsened by the fact that these pathogens can withstand normal water disinfection processes. The results of this study revealed that *Giardia* cysts were ubiquitous in the influents collected from Zeekoegat, Baviaanspoort, Rayton and Refilwe WCWs, whereas *Cryptosporidium* oocysts occurred less frequently. Similar occurrences of cysts and oocysts were reported by previous investigators (Brandonisio et al., 2000; Cacció et al., 2003). Even after the treatment of the influents, the (oo)cysts were still found in considerable concentrations in the final effluents of the Baviaanspoort, Rayton and Refilwe WCWs, as indicated in Tables 3 and 4. Despite the high removal efficiencies of *Cryptosporidium* and *Giardia* recorded at the Zeekoegat, Baviaanspoort, Rayton and Refilwe WCWs during the study period (Tables 3 and 4), the collective range of (oo)cysts detected in final effluents discharged into the receiving water bodies ranged from <1 to 215 cysts/l. The highest removal of oocysts and cysts (98.36% and 99.96%, respectively) noted at the Zeekoegat WCW could be attributed to the presence of the rapid sand filtration process, which resulted in the physical removal of the protozoan parasites from the water, as the operational chlorine disinfection dosing levels (between 1.5 and 2.0 mg/l) employed in all plants are considered ineffective for inactivating oocysts and cysts (Clancy et al., 2004).

The findings of the present study concur with those obtained by Casson and co-workers (1990) and Robertson and co-workers (2000). Casson and co-workers indicated that primary settlement had an (oo)cyst removal efficiency of between 0 and 53% at one wastewater plant. Robertson and co-workers (2000) estimated the removal efficiency of *Giardia* cysts to range between 60 and 90% in wastewater treatment plants incorporating a primary and secondary process, and removal efficiency varied from under 10% to over 90% for *Cryptosporidium* oocysts. A comparison of these 2 studies demonstrated that secondary treatments were of more importance in removal of these parasites than primary settlement.

Using the US EPA Method 1623, Castro-Hermida and co-workers (2008) also found both parasites in influent and effluent samples from 12 Galicia (northwest Spain) wastewater treatment plants throughout the year, and *Giardia* cysts (*G. duodenalis*) always outnumbered *Cryptosporidium* spp. The authors indicated that the mean concentration of *Giardia* cysts per litre of influent/effluent was significantly higher ($p < 0.05$) than the mean concentration of *Cryptosporidium* spp. per litre of influent/effluent. Similar observations were also noted in the present study (Tables 3 and 4). These findings, therefore, confirm the sensitivity level of US EPA Method 1623 in the detection of *Cryptosporidium* oocysts and species, and *Giardia* cysts, in wastewater sources.

Giardia cysts and *Cryptosporidium* oocysts have been found in a number of different waters and in treated drinking water in South Africa, and are commonly found in surface waters. In a study conducted in the former Natal and Transvaal provinces over a period of 3 years (1992-1994), samples were taken from raw sewage, treated effluents, surface water and drinking water. All of the samples collected from 10 surface water sites were contaminated with *Giardia* cysts and *Cryptosporidium* oocysts at levels ranging between 50 and 1 000 counts/100l (Gericke et al., 1995). The authors also indicated that conventional wastewater treatment works could not significantly reduce cyst or oocyst numbers, from raw wastewater to treated effluents discharged in the receiving water body (Gericke et al., 1995). These findings corroborate with those of the present investigation in terms of (oo)cyst removal, especially by Refilwe, Rayton and Baviaanspoort WCWs. Since 1995, Stellenbosch Municipality had been testing quarterly for parasites at the town's 3 untreated raw water sources. The presence of both *Giardia* cysts and *Cryptosporidium* oocysts was detected for the first time in March 1998 (Mackintosh et al., 2000).

Giardia cysts and *Cryptosporidium* oocysts are extremely resistant to environmental stress and to treatment and are minimally affected by chlorination, as stated above. They may also survive for days or months in the environmental water, and animals may act as reservoirs of infection (DWAf, 1996). This clearly explained their persistence in the effluents collected from the Refilwe WCW, while these parasites could

be detected from time to time at other plants, especially at the Baviaanspoort and Rayton WCWs. In some cases, their absence or lower counts in the influents were coupled with higher counts in the effluents (Tables 3 and 4). This demonstrated the ability of the (oo)cysts to survive for days or months in the environmental water even after their discharge into receiving water bodies.

The infective dose for *Cryptosporidium* and *Giardia* is as low as 1 cyst or oocyst, which is quite sufficient to cause infection (DWAF, 1996; Cacció et al., 2003). As the amount of (oo)cysts isolated from the effluents produced by the 4 wastewater treatment plants under this study was greater than 1 per litre (Tables 3 and 4), there might be a risk of protozoan parasite infection for continuous, short or occasional exposures to these water sources. The microbiological quality of the effluents discharged by all these plants is therefore a matter of concern, especially for the surrounding communities who use these water sources, including their respective receiving water bodies, for drinking, recreational and agricultural purposes. Consequently, more consideration should be given to the management of the drinking water plants in order to comply with the suggested safe levels of less than 1 (oo)cyst/l, as set nationally by water services authorities (DWAF, 1996).

Immunomagnetic separation-based epifluorescence microscopy, a conventional diagnostic method, is capable of identifying the genera *Cryptosporidium* and *Giardia* based on morphological features. However this technique does not allow a reliable identification of species. Hence, the identification of genotypes requires the use of molecular assays. The most widely-used markers (the small subunit ribosomal DNA and a set of nuclear genes) enable the 2 major subtypes in the *C. parvum* and *G. lamblia* species to be distinguished. This is possible through the selection of sequences that display a higher level of polymorphism, as proved by previous investigators (Johnson et al., 1995; Spano et al., 1997; Cacció et al., 2003). Therefore, in this study, the occurrence of *Cryptosporidium* and *Giardia* (oo)cysts in wastewater treatment plants was investigated up to the species subtype level. This is of importance, given that *G. lamblia* and *C. parvum* are associated with both animal and human infections (Cacció et al., 2002; 2003).

The most commonly identified etiological agents of human cryptosporidiosis and giardiasis have been *Cryptosporidium parvum* and *G. lamblia*. Based on the molecular characterisation of oocysts and cysts, these parasites can be divided into 2 groups of genetically distinct subtypes: *C. parvum* Genotype I and *G. lamblia* Assemblage A. These 2 groups are associated wholly with human infection while Genotype II and Assemblage B are linked to both human and animal infections (Xiao et al., 2000). During the study period it was noted that the Refilwe WCW discharged significantly higher loads of oocysts and cysts per litre compared to other wastewater treatment plants. This was confirmed at a molecular level, which resulted in the identification of *C. Parvum Genotype I* and *G. lamblia Assemblage A* in 50% and 62%, respectively, of samples collected from this plant. The molecular study also revealed the presence of *C. parvum Genotype II* and *G. lamblia Assemblage B* in 29% and 57%, respectively, of the effluent samples taken from this plant. Although viability studies on oocysts and cysts and epidemiological studies in the community served by this plant were not carried out, these findings indicate that the effluents processed at this wastewater treatment plant could be a potential source of human infection with *C. parvum* and *G. lamblia*. No species identification was possible for samples taken from the effluents of Zeekoegat WCW.

This could be due to the fact that the (oo)cysts, as suggested by observation through immunomagnetic methods, could be of a different subspecies, and thus amplification could not be achieved with the primers used in this study (Cacció et al., 2003; Sturbaum et al., 2001). In view of this observation, the presence of oocysts and cysts, in the absence of species data, does not imply a risk of transmission of protozoan parasites of this genera that are of health risk to humans. In general, this study revealed that the risk of contamination of water courses by *Cryptosporidium* ssp. and *Giardia* spp. is considerable. It is important that wastewater treatment authorities consider the relevance of the levels of contamination by both parasites in wastewater, and develop adequate countermeasures.

Conclusions and recommendations

The results of this study provide insight into wastewater treatment plants as a source of pollution of rivers with respect to *Cryptosporidium* and *Giardia* species and also highlight the potential health risk associated with the use of the effluents or the receiving water bodies for drinking, recreational and agricultural purposes. Despite the high removal efficiencies of these protozoan parasites demonstrated by all of the wastewater treatment plants sampled, the results of this study revealed that *Cryptosporidium* and *Giardia* persisted in the effluents collected from the Refilwe WCW. At the other plants, especially the Baviaanspoort and Rayton WCWs, these parasites could also be detected from time to time. In other cases, their absence in the influents and presence in the effluents, or lower counts in the influents coupled with higher counts in the effluents, was noted. This demonstrated the ability of the (oo)cysts to survive for days or months in the environmental water even after their discharge into the receiving water bodies.

The study also revealed that the high removal efficiency of *Cryptosporidium* and *Giardia* noted in Zeekoegat WCW was linked to the physical process incorporated into this plant. To curb disease outbreaks related to *Cryptosporidium* and *Giardia*, this study therefore suggests an upgrading of the wastewater treatment plants by including processes such as sand filters and UV light disinfection, which have proved to be effective in the removal and inactivation of these protozoan parasites.

We thus conclude that additional water purification procedures are necessary if the health risk posed by this organism is to be effectively eliminated. These findings may provide further insight into the possible sources of *Cryptosporidium* and *Giardia* in the natural environment, and stress the potential risk associated with the use of waters for recreational and agricultural purposes.

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