Synthesis and Characterization of 5-Substituted 1H-Tetrazoles in the Presence of Nano-TiCl₄·SiO₂

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ABSTRACT
Nano-TiCl₄·SiO₂ was found to be an extremely efficient catalyst for the preparation of 5-substituted 1H-tetrazole derivatives. Nano-TiCl₄·SiO₂ is a solid Lewis-acid was synthesized by the reaction of nano-SiO₂ and TiCl₄. The structure characterization of this acid was achieved with X-ray diffraction, thermogravimetric analysis and electron microscopy. The synthesis of the catalyst is simple and environmentally benign with a good yield. Furthermore, the catalyst is conveniently recoverable and was reused for at least three times. The antimicrobial activities of the synthetic compounds were also determined by both microdilution methods as recommended by the Clinical Laboratory Standard Institute, but unfortunately did not exhibit antibacterial activities at the highest concentration (256 µL mL⁻¹). Further studies are still needed to investigate the potential biological activities of these compounds against other diseases.

KEYWORDS
Nano-TiCl₄·SiO₂, heterogeneous catalyst, 5-substituted 1H-tetrazoles, antibacterial.

1. Introduction
Tetrazoles are an increasingly popular functionality with wide ranging applications. There is considerable interest in the medicinal and biological applications of tetrazoles, due to their reported anti-allergic and anti-asthmatic, antiviral and anti-inflammatory, anti-neoplastic, and cognition disorder activities. Tetrazoles are also applied as ligands in coordination chemistry as explosives and rocket propellants and they are used as isosteric replacements for carboxylic acids in drug design.

Tetrazole derivatives are used as antibiotics and optically active tetrazole-containing antifungal preparations of azole type was reported. There is always a need for new and effective antifungal and antibacterial agents with broad-spectrum activities. It was decided to develop this interest by ascertaining the molecules features essential for activity and utilizing them to develop a new class of potential drugs.

The conventional method of synthesizing tetrazoles is via the addition of azide ions to organic nitriles or cyanamides. Later Sharpless and co-workers reported an innovative and safe procedure for the synthesis of tetrazoles by the addition of sodium azide to nitriles using stoichiometric amounts of Zn(II) salts in water. Pizzo and co-workers efficiently synthesized tetrazoles by the addition of TMSN₃ to organic nitriles using 10 mol% TBAF as catalyst. Several syntheses of 5-substituted tetrazoles were reported through the [2+3] cycloaddition of nitriles to Na₃N or TMSN₃, in the presence of catalysts such as Montmorillonite K-10 Clay, CuCl₂, FeCl₃·SiO₂, Zeolite and sulfated zirconia, BaWO₄·18 Nano-TiO₂, MoO₃·SiO₂.

However, each method has certain restrictions with regard to scope and reaction conditions; for example, costs of synthesis, unrecoverable catalysts, strong acidic conditions, long reaction times, low yields, difficult work up and harsh reaction conditions. To avoid these limitations, our studies attempted the development of more efficient methods accompanied with higher yields for the synthesis of tetrazoles in the presence of nano-TiCl₄·SiO₂ and to evaluate their antibacterial and antifungal properties.

2. Experimental
2.1. General
The chemicals were purchased from Merck and used without any additional purification. The products were characterized by FT-IR (ATR), 1H-NMR, and a comparison of their physical properties with those reported in the literature. FT-IR (ATR) spectra were acquired on a Bruker, Equinox 55 spectrometer. A Bruker (DRX-400 Avance) nmr was used to record the 1H NMR spectra. The X-ray diffraction (XRD) patterns of materials were obtained by employing a Philips Xpert MPD diffractometer (The instrument is a PAN alytical X'Pert Pro MPD, powered by a Philips PW3040/60 X-ray generator and fitted with an X'Celerator detector). Diffraction data is acquired by exposing powder samples to Cu-Kα X-ray radiation, which has a characteristic wavelength of 1.5418 Å. X-rays were generated from a Cu anode supplied with 40 kV and a current of 40 mA), equipped with a Cu Kz anode (λ = 1.54 Å) in the 2θ range from 10 to 80 °. The SEM of nanoparticles determined with VEGA/TESCAN scanning electron microscope (model: Mira 3-XMU) and TEM photograph was prepared by Leo 912AB OMEGA microscope. The thermal gravimetric analysis (TGA) was performed with a ‘NETZSCH TG 209 F1 Iris’ instrument, Spectrophotometer (UV/Vis biotek model UVIKONXL), Vortex mixer (Heidolf, d-91126 schwabach), Retsch Mixer Mill MM400, 100-240 VAC, 50/60 Hz, Sonication, Bandelin SONOPULS, HD3220 homogenizer, TT13 probe, Microwave, KENWOOD K25MW11 Microwave Oven, Melting points were determined with a Thermo Scientific Electrothermal digital apparatus (Thermo Fisher Scientific Inc.).
2.2. General Method for the Synthesis of Tetrazole Derivatives:
Nano-TiCl₄·SiO₂ (0.1 g) was added to a mixture of benzonitrile (1 mmol), sodium azide (2 mmol) in DMF (5 mL) at reflux for 2 h. After completion of reaction (as monitored by TLC), the mixture was allowed to cool to room temperature, the catalyst was removed by filtration. Then by adding ice water and 4N HCl (5 mL) to the residue, a white solid was obtained. This was then washed with cold chloroform. This simple procedure yielded pure tetrazole with good yields.

The products are known and were identified by comparison of their physical and spectral data with those of authentic samples.

2.3. Determination of Antifungal Activities

2.3.1. Microorganisms
The antifungal activities of the synthetic compounds against eighteen American Type Culture Collection (ATCC) strains of fungi, including Candida albicans (ATCC 10261, 562, 1949, 1912, 5982, 2730), C. tropicalis (ATCC 750), C. krusei (ATCC 6258), C. glabrata (ATCC 90030, 6144, 2175, 863), C. parapsilosis (ATCC 4344), C. dubliniensis (ATCC 8500, 7987, 8501), Cryptococcus neoformans (ATCC 9011), Aspergillus flavus (ATCC 64025), A. fumigatus (ATCC 14110) and A. clavatus (CBS 514.65) were determined.

The antibacterial activities of the EO against standard species of S. aureus (ATCC 700698), Escherichia coli (ATCC 25912), E. faecalis (ATCC1700), P. aeruginosa (ATCC 27853), and B. cereus (11778) were also determined in this study. The susceptibility of all clinical isolates of bacteria and fungi against select antibiotics was examined by microdilution and disk diffusion methods. The susceptibility of all isolates of fungi against select antibiotics was examined by microdilution and disk diffusion methods.

2.3.2. Determination of Minimum Inhibitory Concentration
MICs were determined using broth microdilution method recommended by the CLSI with some modifications. Briefly, for determination of antifungal activities against fungi, serial dilutions of the synthetic compounds (0.25 to 256.0 µg mL⁻¹) were prepared in 96-well microtitre plates using RPMI-1640 media (Sigma, St. Louis, USA) buffered with MOPS (Sigma, St. Louis, USA). To determine the antibacterial activities, serial dilutions of the synthetic compounds (0.25 to 256.0 µg mL⁻¹) were prepared in Mueller-Hinton Broth media (Merck, Darmstadt, Germany). Test fungi or bacteria strains were suspended in the media and the cell densities were adjusted to 0.5 McFarland standards at 530 nm wavelength using a spectrophotometric method (this yields stock suspension of 1.5 × 10⁶ cells mL⁻¹ for yeast and 1–1.5 × 10⁸ cells mL⁻¹ for bacteria). One hundred microlitre of the working inoculums was added to the micolitre plates which were incubated in a humid atmosphere at 30 °C for 24–48 h (fungi) or at 37 °C for 24 h (bacteria). Two hundred microlitre of the uninoculated medium was included as a sterility control (blank). In addition, growth controls (medium with inoculums but without drugs) were also included. The growth in each well was compared with that of the growth control well. MICs were visually determined and defined as the lowest concentration of the synthetic compounds or drugs produced no visible growth. Each experiment was performed in triplicate. Each experiment was performed in triplicate. In addition, media from wells with fungi showing no visible growth were further cultured on Sabouraud Dextrose Agar (Merck, Darmstadt, Germany) and from wells with bacteria showing no visible growth on Muller-Hinton agar (Merck, Darmstadt, Germany) to determine the minimum fungicidal concentration (MFC) and minimum bactericidal concentration (MBC). MBCs and MFCs were determined as the lowest concentration yielding no more than four colonies, which corresponds to a mortality of 99.9 % of the microbes in the initial inoculums.

3. Results and Discussion

3.1. Optimization of the Reaction Conditions
We investigated the synthesis of 5-substituted 1H-tetrazole in the presence of nano-TiCl₄·SiO₂. However our study is the first to characterize and report the IR spectra of nano-SiO₂, nano-TiCl₄, SiO₂ and TiCl₄. In literature, the XRD pattern of nano-SiO₂ has a strong peak in the 2θ value of 21.8024° with full width at half maximum (FWHM) equal to 0.1771.

By contrast, our data showed that the particle size of nano-TiCl₄·SiO₂ in TEM and SEM patterns are calculated as 14–20 nm and 37–41 nm, respectively (Fig. 1). Thermal gravimetric analysis (TGA) pattern of nano-TiCl₄·SiO₂ was detected from 25.43 to 513.43 °C. The catalyst is stable below 173.43 °C and only 2.98 % of its weight was reduced at 173.43 °C, which is related to removal of catalyst moisture.

Initially, in an effort to develop better reaction conditions, different solvents and conditions were screened for the preparation of 5-substituted 1H-tetrazole from the reaction of benzonitrile with sodium azide in the presence of nano-TiCl₄·SiO₂ and the results are summarized in Table 1. Among the different solvents screened, DMF gave the product in good yield at reflux tempera-
ture (Table 1, entry 1). Other solvents such as toluene gave the desired products in low yield (Table 1, entry 3). On the other hand, the product was formed in 60 % when the reaction was performed under solvent-free conditions (Table 1, entry 5).

According to the obtained data, the best conditions were at reflux temperature in DMF using 0.2 g of TiCl₄·SiO₂ or 0.1 g of nano-TiCl₄·SiO₂ (Table 1, entries 1, 13).

Once the scope of the reaction condition was established, the reusability of the catalyst was examined. After performing the reaction, the catalyst was separated, washed with acetone, dried and re-used up to three times in reaction (Table 1, entries 15, 16). The catalyst was reusable although a gradual decline was observed in its activity.

Also, we tried to reaction of benzonitrile with sodium azide using mixer mill, ultrasonic and microwave, but these conditions didn’t give the product in good yield (Table 1, entries 6, 7 and 8).

Under the optimized reaction conditions, we chose a variety of structurally divergent benzonitriles to explore the scope and generality of the nano-TiCl₄·SiO₂ promoted [2+3] cycloaddition reaction to form 5-substituted 1H-tetrazoles and the results are presented in Table 2. It seems that the nature of the substituents on the aromatic ring of benzonitriles exert different influences. It is of interest to note that electron-withdrawing groups that increase the polarity of the cyanide group inductively (Table 2, entry 4-nitro) give higher yields of products compared to the electron-donating groups (Table 2, entry 3-hydroxy). Different halogen substituted benzonitriles, such as 4-chlorobenzonitrile and 4-bromobenzonitrile reacted smoothly and gave the desired products in decent yields (Table 2, entries 5, 6).

Heteroaromatic nitriles such as 2-pyridinecarbonitrile gave the corresponding tetrazoles in shorter reaction times with excellent yields (Table 2, entry 2). Unfortunately the present method is not amenable for aliphatic nitriles.

Some of the previous studies reported promising antimicrobial activities for tetrazole derivatives, 31,32 none of the synthetic compounds in this study exhibited antifungal activity against the examined fungi at the tested concentrations which is similar to the study of Bekhit et al. 33 Moreover, the examined compounds failed to inhibit the growth of the tested Gram-positive and Gram-negative bacteria at the concentration up to 256 µg mL⁻¹.

Besides to antimicrobial activities, 31,32 some of the tetrazole derivatives exhibit biological properties including anti-inflammatory, inhibition of cyclo-oxygenase, antidiabetic, anticonvulsant, and anticancer activities. 34–36 Hence, further studies are required to investigate the other potential biological activities of these synthetic compounds.

**4. Conclusion**

We have demonstrated a simple method for the preparation of 5-substituted 1H-tetrazole derivatives using nano-TiCl₄·SiO₂ as eco-friendly and efficient catalyst in a one-pot procedure. Short reaction times, high yields, a clean process, simple methodology, easy work-up and green conditions are advantages of this protocol. Since some of the tetrazole derivatives previously showed antibacterial 35 and antifungal 31,32 activities, further studies are still required for the design and synthesis of more novel tetrazole derivatives with better antimicrobial activities by this simple green method.

**Supplementary material**

The IR, ¹³C and ¹H NMR spectra of the novel 5-substituted
Table 2  Synthesis of 5-substituted 1H-tetrazole derivatives at reflux/DMF in the presence of nano-TiCl₄·SiO₂.

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<thead>
<tr>
<th>Entry R (Ar)</th>
<th>Products</th>
<th>Yield/%b</th>
<th>Time/h</th>
<th>MP/C</th>
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<td>214–216</td>
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<tr>
<td>2 4-CH₂Ph</td>
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<td>251–152</td>
<td>250–251[15]</td>
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<tr>
<td>3 4-OPh</td>
<td><img src="image3" alt="Product" /></td>
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<td>2.5</td>
<td>231–233</td>
<td>234–235[16]</td>
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<tr>
<td>4 3-NO₂Ph</td>
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<td>1.5</td>
<td>217–220</td>
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</tr>
<tr>
<td>5 4-CIPh</td>
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<tr>
<td>7 PHCH₂</td>
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ArCN + NaN₃ → TiCl₄·SiO₂ → Reflux/DMF

[a] Reaction conditions: nitrile (1 mmol), NaN₃ (2 mmol), (0.1 g), nano-TiCl₄·SiO₂, DMF (5 ml) at reflux.
[b] Isolated yields.

1H-tetrazole derivatives (Table 2, compounds 9, 11 and 12) are presented in the online supplement.

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Here, we describe the IR, 13C and 1H NMR spectra of the novel 5-substituted 1*H*-tetrazolatederivatives (Table 2, compounds 9, 11 and 12).

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5-Phenyl-1H-tetrazole

Yield: 95%, White crystal.

Figure S1. FT-IR: $\tilde{\nu}$ (KBr) = 2600-300, 1608, 1563, 1485, 1465, 1409, 1480, 726, 687 cm$^{-1}$.

Figure S2. $^1$H NMR (400 MHz, DMSO-d$_6$): 8.01 (brs, 2H), 7.51 (brs, 3H) ppm.
5-(4-Methylphenyl)-1H-tetrazole

Yield: 84%, White crystal.

**Figure S3.** FT-IR: $\tilde{\nu}$ (KBr) = 2600-300, 1608, 1563, 1485, 1465, 1409, 1480, 726, 687 cm$^{-1}$.

**Figure S4, S5.** $^1$H NMR (500 MHz, CDCl$_3$): 8.81 (d, $J = 5.8$, 2H), 8.09 (d, $J = 5.8$, 2H), 2.5 (s, 3H) ppm.
Figure S4: $^1$H NMR (500 MHz, CDCl$_3$) 5-(4-Methylphenyl)-1H-tetrazole

Figure S5: $^1$H NMR (500 MHz, CDCl$_3$) 5-(4-Methylphenyl)-1H-tetrazole (expand)
5-(4-Hydroxyphenyl)-1H-tetrazole

Yield: 82%, White crystal.

**Figure S6.** FT-IR: $\tilde{\nu}$ (KBr) = 2500-3400, 1648, 1600, 1515, 1470, 1435, 1080, 842 cm$^{-1}$.

**Figure S7, S8.** $^1$H NMR (400 MHz, DMSO-d6): 16.5 (brs, NH), 10.17 (s, 1H), 7.84 (d, $J$ = 8, 2H), 6.93 (d, $J$ = 7.6, 2H) ppm.
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Figure S8. $^1$H NMR (500 MHz, DMSO-d$_6$) 5-(4-Hydroxyphenyl)-1H-tetrazole (expand)
5-(3-Nitrophenyl)-1H-tetrazole
Yield: 92%, White crystal.

**Figure S9.** FT-IR: $\tilde{\nu}$ (KBr) = 2600-3300, 1626, 1569, 1528, 1349, 872, 743, 973 cm$^{-1}$.

**Figure S10, S11.** $^1$H NMR (400 MHz, DMSO-d6): 8.82 (s, 1H), 8.43 (dd, $J = 7.6$ and 8 Hz, 2H), 7.89 (t, $J = 8.4$, 1H) ppm.
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Yield: 91%, White crystal.

Figure S12. FT-IR: $\tilde{\nu}$ (KBr) = 2500-3000, 1654, 1610, 1561, 1487, 1434, 831 cm$^{-1}$.

Figure S13, S14. $^1$H NMR (500 MHz, DMSO-d6): 8.10 (d, $J = 10.53$, 2H), 7.69 (d, $J = 8.41$, 2H) ppm.
Figure S13. $^1$H NMR (500 MHz, DMSO-d$_6$) 5-(4-Chlorophenyl)-1H-tetrazole

Figure S14. $^1$H NMR (500 MHz, DMSO-d$_6$) 5-(4-Chlorophenyl)-1H-tetrazole (expand)
5-(4-Bromophenyl)-1H-tetrazole

Yield: 90%, White crystal

**Figure S15.** FT-IR: $\tilde{\nu}$ (KBr) = 2600-300, 1649, 1604, 1560, 1482, 1431, 1053, 829 cm$^{-1}$.

**Figure S16, S17.** $^1$H NMR (500 MHz, CDCl$_3$): 8.06 (d, $J$= 7.2, 2H), 7.69 (d, $J$= 7.2, 2H) ppm.

*Figure S15: FT-IR (KBr) 5-(4-Bromophenyl)-1H-tetrazole*
Figure S16. $^1$H NMR ((500 MHz, CDCl$_3$): 5-(4- Bromophenyl)-1H-tetrazole

Figure S17. $^1$H NMR ((500 MHz, CDCl$_3$): 5-(4- Bromophenyl)-1H-tetrazole
(expand)
5-Benzyltetrazole

Yield: 78%, White crystal.

**Figure S18.** FT-IR: $\ddot{\nu}$ (KBr) = 2400-300, 1592, 1549, 1496, 1248, 775 cm$^{-1}$.

**Figure S19, S20.** $^1$H NMR (400 MHz, DMSO-d$_6$): 4.26 (s, 2H, CH$_2$), 7.25-1.31 (m, 5H) ppm.
Figure S19. $^1$H NMR (500 MHz, DMSO-$d_6$) 5-Benzyltetrazole

Figure S20. $^1$H NMR (500 MHz, DMSO-$d_6$) 5-Benzyltetrazole (expand)
5-((4-Methoxyphenyl)methyl)tetrazole
Yield: 81%, White crystal.

**Figure S21.** FT-IR: $\nu$ (KBr) = 2600-3400, 1636, 1514, 1124, 848 cm$^{-1}$.

**Figure S22, S23.** $^1$H NMR (400 MHz, DMSO-d6): 7.19 (brs, 2H), 6.86 (brs, 2H), 4.18 (s, 2H), 3.69 (s, 3H) ppm.
Figure S22. $^1$H NMR (500 MHz, DMSO-$d_6$) 5-((4-Methoxyphenyl)methyl)tetrazole

Figure S23. $^1$H NMR (500 MHz, DMSO-$d_6$) 5-((4-Methoxyphenyl)methyl)tetrazole (expand)
5-((4-Chlorophenyl)methyl)tetrazole

Yield: 84%, White crystal.

**Figure S24.** FT-IR: $\tilde{\nu}$ (KBr) = 2600-300, 1538, 1492, 1407, 1263, 1207, 834 cm$^{-1}$.

**Figure S25, S26.** $^1$H NMR (400 MHz, DMSO-d6): 7.40 (m, 2H), 7.31 (d, $J$ = 8, 2H), 4.30 (s, 2H) ppm.

**Figure S27.** $^{13}$C-NMR (125 MHz, DMSO) $\delta$ = 155.0, 132.5, 128.9, 124.7, 123.6, 28.8 ppm.
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Figure S26. $^1$H NMR (500 MHz, DMSO-d$_6$) 5-((4-Chlorophenyl)methyl)tetrazole
(expand)
Figure S27. $^{13}\text{C}$ NMR (500 MHz, DMSO) of 5-((4-Chlorophenyl)methyl)tetrazole
5-Benzhydryltetrazole

Yield: 88%, White crystal.

**Figure S28.** FT-IR: $\tilde{\nu}$ (KBr) = 2600-300, 1567, 1496, 1245, 745 cm$^{-1}$.

**Figure S29, S30.** $^1$H NMR (500 MHz, CDCl$_3$): 5.82 (s, 1H), 8.128 (brs, 1H), 7.25-7.41 (m, 10H) ppm.

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**Figure S28.** FT-IR (KBr) of 5-Benzhydryltetrazole
Figure S29. $^1$H NMR (500 MHz, CDCl$_3$) of 5-Benzyhydryltetrazole

Figure S30. $^1$H NMR (500 MHz, CDCl$_3$) of 5-Benzyhydryltetrazole (expand)
5-((3,4-dichlorophenyl)methyl)tetrazole

Yield: 84%, White crystal.

**Figure S31.** FT-IR: $\tilde{\nu}$ (KBr) = 2500-3300, 1560, 1472, 1440, 1260, 1210, 827, 766, 706, 674 cm$^{-1}$.

**Figure S32, S33.** $^1$H NMR (500 MHz, CDCl$_3$): 7.61 (d, J= 8.45, 2H, 2H), 7.28 (d, J= 8.2, 1H), 4.32 (s, 2H) ppm.

**Figure S34.** $^{13}$C-NMR (125 MHz, DMSO) $\delta$ = 28.8, 130.2, 131.6, 131.8, 174.8 ppm

![Figure S31: FT-IR (KBr) 5-((3,4-dichlorophenyl)methyl)tetrazole](image-url)
Figure S32. $^1$H NMR (500 MHz, CDCl$_3$) 5-((3,4-dichlorophenyl)methyl)tetrazole

Figure S33. $^1$H NMR (500 MHz, CDCl$_3$) 5-((3,4-dichlorophenyl)methyl)tetrazole (expand)
**Figure S34.** $^{13}$C NMR (500 MHz, DMSO) 5-((3,4-dichlorophenyl)methyl)tetrazole
4-(1H-tetrazole-5-yl)pyridine (table 2, entry 12)
Yield: 92%, White crystal.

**Figure S35.** FT-IR: $\tilde{\nu}$ (KBr) = 2500-3000, 1631, 1529, 1440, 1338, 1292, 1042, 990, 846, 751 cm$^{-1}$.

**Figure S36, S37.** $^1$H NMR (500 MHz, CDCl$_3$): 8.00 (d, $J$ = 7.89, 2H), 7.40 (d, $J$ = 7.86, 2H) ppm.

**Figure S38.** $^{13}$C-NMR (125 MHz, DMSO) $\delta$ = 127.9, 130.7 ppm.
**Figure S36.** $^1$H NMR (500 MHz, CDCl$_3$) 4-(1$H$-tetrazole-5-yl)pyridine

**Figure S37.** $^1$H NMR (500 MHz, CDCl$_3$) 4-(1$H$-tetrazole-5-yl)pyridine (expand)
**Figure S38.** $^{13}$C NMR (500 MHz, DMSO) 4-(1$H$-tetrazole-5-yl)pyridine
Characterization of 5-((3,4-dichlorophenyl)methyl)tetrazole was completed using FT-IR, $^1$H NMR and $^{13}$C NMR. The marked structure of 5-((3, 4-dichlorophenyl) methyl) tetrazole is showed in Figure 1.

As can be seen in the IR spectrum, the stretching frequencies of C-H and N-H groups are indicated at 2500-3300 cm$^{-1}$. The stretching frequency of C=C group is demonstrated at 1560 cm$^{-1}$. The frequency absorption of tetrazole ring is specified at 1472 cm$^{-1}$. The stretching frequency of C-H benzyl group at 1260 cm$^{-1}$ and the bending frequency of C-H phenyl ring is appeared at 827, 766, 706 cm$^{-1}$ (Figure 1).

In the $^1$H NMR spectrum, the appearance of the methylene protons (Hc, figure 1) as singlet at 4.32 ppm, integrating to two. The signal related to the one aromatic proton (Hh, figure 1) appear as a doublet at 7.28 ppm, integrating to one. In the appearance of a doublet at 7.61 ppm due to the two aromatic protons (Hg and Hd, figure 1), integrating to two, This signal appears at up filed due to the deshielding nature of the neighbouring chlorine atoms.

The $^{13}$CNMR spectrum for 5-((3,4-dichlorophenyl)methyl) tetrazole displays signals characteristic various carbones (Cb, Cd, Cg, Ch, Cc, figure 1) at 28.8, 130.2, 131.6, 131.8, 174.8 ppm, respectively.
References