Synthesis and Characterization of 5-Substituted 1*H*-Tetrazoles in the Presence of Nano-TiCl₄·SiO₂

Leila Zamani^{a,*}, Bi Bi Fatemeh Mirjalili^a, Kamiar Zomorodian^b and Sogol Zomorodian^b

^aDepartment of Chemistry, College of Science, Yazd University, Yazd, P.O.Box 8915813149, Iran.

^bCenter of Basic Researches in Infectious Diseases and Department of Medical Mycology and Parasitology School of Medicine, Shiraz University of Medical Sciences, Post code 71348-45794, Shiraz, Iran.

Received 22 April 2014, revised 17 March 2015, accepted 18 March 2014.

ABSTRACT

Nano-TiCl₄·SiO₂ was found to be an extremely efficient catalyst for the preparation of 5-substituted 1*H*-tetrazole derivatives. Nano-TiCl₄·SiO₂ is a solid Lewis-acid was synthesized by the reaction of nano-SiO₂ and TiCl₄. The structure characterization of this acid was achieved with X-ray diffraction, thermogravimetric analysis and electron microscopy. The synthesis of the catalyst is simple and environmentally benign with a good yield. Furthermore, the catalyst is conveniently recoverable and was reused for at least three times. The antimicrobial activities of the synthetic compounds were also determined by both micro dilution methods as recommended by the Clinical Laboratory Standard Institute, but unfortunately did not exhibit antibacterial activities at the highest concentration (256 μL mL⁻¹). Further studies are still needed to investigate the potential biological activities of these compounds against other diseases.

KEYWORDS

Nano-TiCl₄·SiO₂, heterogeneous catalyst, 5-substituted 1*H*-tetrazoles, antibacterial.

1. Introduction

Tetrazoles are an increasingly popular functionality with wide ranging applications. There is considerable interest in the medicinal and biological applications of tetrazoles,^{1,2} due to their reported anti-allergic and anti-asthmatic,^{3,4} antiviral and anti-inflammatory, anti-neoplastic, and cognition disorder activities.^{5,6,7} Tetrazoles are also applied as ligands in coordination chemistry⁸ as explosives and rocket propellants and they are used as isosteric replacements for carboxylic acids in drug design.⁹

Tetrazole derivatives are used as antibiotics and optically active tetrazole-containing antifungal preparations of azole type was reported. There is always a need for new and effective antifungal and antibacterial agents with broad-spectrum activities. It was decided to develop this interest by ascertaining the molecules features essential for activity and utilizing them to develop a new class of potential drugs.

The conventional method of synthesizing tetrazoles is *via* the addition of azide ions to organic nitriles or cyanamides.¹¹ Later Sharpless and co-workers reported an innovative and safe procedure for the synthesis of tetrazoles by the addition of sodium azide to nitriles using stoichiometric amounts of Zn(II) salts in water.¹² Pizzo and co-workers efficiently synthesized tetrazoles by the addition of TMSN₃ to organic nitriles using 10 mol% TBAF as catalyst.¹³ Several syntheses of 5-substituted tetrazoles were reported through the [2+3] cyclo-addition of nitriles to NaN₃ or TMSN₃ in the presence of catalysts such as Montmorillonite K-10 Clay,¹⁴ Cu₂O,¹⁵ FeCl₃·SiO₂,¹⁶ Zeolite and sulfated zirconia,¹⁷ BaWO₄,¹⁸ Nano-TiO₂,¹⁹ MoO₃·SiO₂.²⁰

However, each method has certain restrictions with regard to scope and reaction conditions; for example, costs of synthesis, unrecoverable catalysts, strong acidic conditions, long reaction times, low yields, difficult work up and harsh reaction conditions. To avoid these limitations, our studies attempted the develop-

ment of more efficient methods accompanied with higher yields for the synthesis of tetrazoles in the presence of nano-TiCl₄·SiO₂ and to evaluate their antibacterial and antifungal properties.

2. Experimental

2.1. General

The chemicals were purchased from Merck and used without any additional purification. The products were characterized by FT-IR (ATR), ¹H-NMR, and a comparison of their physical properties with those reported in the literature. FT-IR (ATR) spectra were acquired on a Bruker, Equinox 55 spectrometer. A Bruker (DRX-400 Avance) nmR was used to record the ¹H NMR spectra. The X-ray diffraction (XRD) patterns of materials were obtained by employing a Philips Xpert MPD diffractometer (The instrument is a PAN alytical X'Pert Pro MPD, powered by a Philips PW3040/60 X-ray generator and fitted with an X'Celerator detector. Diffraction data is acquired by exposing powder samples to Cu-Kα X-ray radiation, which has a characteristic wavelength of 1.5418 Å. X-rays were generated from a Cu anode supplied with 40 kV and a current of 40 mA), equipped with a Cu Kα anode (λ = 1.54 Å) in the 2θ range from 10 to 80°. The SEM of nanoparticles determined with VEGA/TESCAN scanning electron microscope (model: Mira 3-XMU) and TEM photograph was prepared by Leo 912AB OMEGA microscope. The thermal gravimetric analysis (TGA) was performed with a 'NETZSCH TG 209 F1 Iris' instrument, Spectrophotometer (UV/Vis biotek model UVIKONXL), Vortex mixer (Heidolf, d-91126 schwabach), Retsch Mixer Mill MM400, 100-240 VAC, 50/60 Hz, Sonication, Bandelin SONOPULS, HD3220 homogenizer, TT13 probe, Microwave, KENWOOD K25MW11 Microwave Oven, Melting points were determined with a Thermo Scientific Electrothermal digital apparatus (Thermo Fisher Scientific Inc.).

* To whom correspondence should be addressed. E-mail: l.zamani@stu.yazd.ac.ir

2.2. General Method for the Synthesis of Tetrazole Derivatives:

Nano- $\text{TiCl}_4 \cdot \text{SiO}_2$ (0.1 g) was added to a mixture of benzonitrile (1 mmol), sodium azide (2 mmol) in DMF (5 mL) at reflux for 2 h. After completion of reaction (as monitored by TLC), the mixture was allowed to cool to room temperature, the catalyst was removed by filtration. Then by adding ice water and 4N HCl (5 mL) to the residue, a white solid was obtained. This was then washed with cold chloroform. This simple procedure yielded pure tetrazole with good yields.

All the products are known and were identified by comparison of their physical and spectral data with those of authentic samples.

2.3. Determination of Antifungal Activities

2.3.1. Microorganisms

The antifungal activities of the synthetic compounds against eighteen American Type Culture Collection (ATCC) strains of fungi, including *Candida albicans* (ATCC 10261, 562, 1949, 1912, 5982, 2730), *C. tropicalis* (ATCC 750), *C. krusei* (ATCC 6258), *C. glabrata* (ATCC 90030, 6144, 2175, 863), *C. parapsilosis* (ATCC 4344), *C. dubliniensis* (ATCC 8500, 7987, 8501), *Cryptococcus neoformans* (ATCC 9011), *Aspergillus flavus* (ATCC 64025), *A. fumigatus* (ATCC 14110) and *A. clavatus* (CBS 514.65) were determined.

The antibacterial activities of the EO against standard species of *S. aureus* (ATCC 700698), *Escherichia coli* (ATCC 25912), *E. faecalis* (ATCC11700), *P. aeruginosa* (ATCC 27853), and *B. cereus* (11778) were also determined in this study. The susceptibility of all clinical isolates of bacteria and fungi against select antibiotics were examined by microdilution and disk diffusion methods. The susceptibility of all isolates of fungi against select antibiotics was examined by microdilution and disk diffusion methods.²¹ Oxacillin and ciprofloxacin were used as reference drugs for bacteria and fluconazole as a reference drug for fungi.

2.3.2. Determination of Minimum Inhibitory Concentration

MICs were determined using broth microdilution method recommended by the CLSI with some modifications.^{22–24} Briefly, for determination of antifungal activities against fungi, serial dilutions of the synthetic compounds (0.25 to $256.0 \mu\text{g mL}^{-1}$) were prepared in 96-well microtitre plates using RPMI-1640 media (Sigma, St. Louis, USA) buffered with MOPS (Sigma, St. Louis, USA). To determine the antibacterial activities, serial dilutions of the synthetic compounds (0.25 to $256.0 \mu\text{g mL}^{-1}$) were prepared in Muller-Hinton Broth media (Merck, Darmstadt, Germany). Test fungi or bacteria strains were suspended in the media and

the cell densities were adjusted to 0.5 McFarland standards at 530 nm wavelength using a spectrophotometric method (this yields stock suspension of $1-5 \times 10^6$ cells mL^{-1} for yeast and $1-1.5 \times 10^8$ cells mL^{-1} for bacteria). One hundred microlitre of the working inoculums was added to the microlitre plates which were incubated in a humid atmosphere at 30°C for 24–48 h (fungi) or at 37°C for 24 h (bacteria). Two hundred microlitre of the uninoculated medium was included as a sterility control (blank). In addition, growth controls (medium with inoculums but without drugs) were also included. The growth in each well was compared with that of the growth control well. MICs were visually determined and defined as the lowest concentration of the synthetic compounds or drugs produced no visible growth. Each experiment was performed in triplicate. Each experiment was performed in triplicate. In addition, media from wells with fungi showing no visible growth were further cultured on Sabouraud Dextrose Agar (Merck, Darmstadt, Germany) and from wells with bacteria showing no visible growth on Muller-Hinton agar (Merck, Darmstadt, Germany) to determine the minimum fungicidal concentration (MFC) and minimum bactericidal concentration (MBC). MBCs and MFCs were determined as the lowest concentration yielding no more than four colonies, which corresponds to a mortality of 99.9 % of the microbes in the initial inoculums.

3. Results and Discussion

3.1. Optimization of the Reaction Conditions

We investigated the synthesis of 5-substituted 1H-tetrazole in the presence of nano- $\text{TiCl}_4 \cdot \text{SiO}_2$. However our study is the first to characterize and report the IR spectra of nano- SiO_2 , nano- $\text{TiCl}_4 \cdot \text{SiO}_2$ and TiCl_4 .^{25–28}

In literature, the XRD pattern of nano- SiO_2 has a strong peak in the 2θ value of 21.8024° with full width at half maximum (FWHM) equal to 0.1771.

By contrast, our data showed that the particle size of nano- $\text{TiCl}_4 \cdot \text{SiO}_2$ in TEM and SEM patterns are calculated as 14–20 nm and 37–41 nm, respectively (Fig. 1). Thermal gravimetric analysis (TGA) pattern of nano- $\text{TiCl}_4 \cdot \text{SiO}_2$ was detected from 23.43 to 513.43°C . The catalyst is stable below 173.43°C and only 2.98 % of its weight was reduced at 173.43°C , which is related to removal of catalyst moisture.

Initially, in an effort to develop better reaction conditions, different solvents and conditions were screened for the preparation of 5-substituted 1H-tetrazole from the reaction of benzonitrile with sodium azide in the presence of nano- $\text{TiCl}_4 \cdot \text{SiO}_2$ and the results are summarized in Table 1. Among the different solvents screened, DMF gave the product in good yield at reflux tempera-

Figure 1 (a) TEM and (b) SEM micrographs of nano- $\text{TiCl}_4 \cdot \text{SiO}_2$.

Table 1 Screening of various solvent for the reaction of benzonitrile with sodium azide.^a

Entry	Catal. (mol%)/g	Solvent	Condition[Time/h]	Yield/% ^b	Ref.
1	TiCl ₄ .SiO ₂ (0.2)	DMF	Reflux[2]	90	
2	TiCl ₄ .SiO ₂ (0.2)	DMSO	Reflux[2]	87	
3	TiCl ₄ .SiO ₂ (0.2)	Toluene	Reflux[2]	40	
4	TiCl ₄ .SiO ₂ (0.2)	THF	Reflux[2]	10	
5	TiCl ₄ .SiO ₂ (0.2)	Solvent-free	120 °C [3]	60	
6	TiCl ₄ .SiO ₂ (0.2)	–	MM ^c [5]	10	
7	TiCl ₄ .SiO ₂ (0.2)	EtOAc	Ultrasonic ^d [1]	10	
8	TiCl ₄ .SiO ₂ (0.2)	–	Microwave ^e [1]	30	
9	TiCl ₄ .SiO ₂ (0.1)	DMF	Reflux[3]	80	
10	TiCl ₄ .SiO ₂ (0.075)	DMF	Reflux[3.5]	72	
11	TiCl ₄ .SiO ₂ (0.05)	DMF	Reflux[4]	60	
12	TiCl ₄ .SiO ₂ (0.025)	DMF	Reflux[6]	55	
13	Nano-TiCl ₄ .SiO ₂ (0.1)	DMF	Reflux[1]	95	
14	Nano-TiCl ₄ .SiO ₂ (0.05)	DMF	Reflux[2]	86	
15	Nano-TiCl ₄ .SiO ₂ (0.1), 2nd run	DMF	Reflux[3]	67	
16	Nano-TiCl ₄ .SiO ₂ (0.1), 3rd run	DMF	Reflux[4]	40	
17	Montmorillonite K-10 Clay	–	Microwave[15 min]	75	14
18	FeCl ₃ .SiO ₂	DMF	120 °C[12]	79	16
19	Cu ₂ O(2.5 mol %)	MeOH/DMF	80 °C[12]	92	15
20	BaWO ₄	DMF	120 °C[24]	80	18

^a Reaction conditions: benzonitrile (1 mmol), NaN₃ (2 mmol), nano-TiCl₄.SiO₂ (0.1 g) in DMF (5 mL) at reflux for 2 h.^b Yield of isolated products.^c Using mixer mill (MM 400) in 25 Hz frequency.^d Using BANDELIN Sonopulse HD 3200 Ultrasonic apparatus with power equal to 20 KHz.^e Using microwave oven Kenwood, 1300W.

ture (Table 1, entry 1). Other solvents such as toluene gave the desired products in low yield (Table 1, entry 3). On the other hand, the product was formed in 60 % when the reaction was performed under solvent-free conditions (Table 1, entry 5).

According to the obtained data, the best conditions were at reflux temperature in DMF using 0.2 g of TiCl₄.SiO₂ or 0.1 g of nano-TiCl₄.SiO₂ (Table 1, entries 1, 13).

Once the scope of the reaction condition was established, the reusability of the catalyst was examined. After performing the reaction, the catalyst was separated, washed with acetone, dried and re-used up to three times in reaction (Table 1, entries 15, 16). The catalyst was reusable although a gradual decline was observed in its activity.

Also, we tried to reaction of benzonitrile with sodium azide using mixer mill, ultrasonic and microwave, but these conditions didn't give the product in good yield (Table 1, entries 6, 7 and 8).

Under the optimized reaction conditions, we chose a variety of structurally divergent benzonitriles to explore the scope and generality of the nano-TiCl₄.SiO₂ promoted [2+3] cycloaddition reaction to form 5-substituted 1H-tetrazoles and the results are presented in Table 2. It seems that the nature of the substituents on the aromatic ring of benzonitriles exert different influences. It is of interest to note that electron-withdrawing groups that increase the polarity of the cyanide group inductively (Table 2, entry 4-nitro) give higher yields of products compared to the electron-donating groups (Table 2, entry 3-hydroxy). Different halogen substituted benzonitriles, such as 4-chlorobenzonitrile and 4-bromobenzonitrile reacted smoothly and gave the desired products in decent yields (Table 2, entries 5, 6).

Heteroaromatic nitriles such as 2-pyridinecarbonitrile gave the corresponding tetrazoles in shorter reaction times with

excellent yields (Table 2, entry 2). Unfortunately the present method is not amenable for aliphatic nitriles.

Although some of the previous studies reported promising antimicrobial activities for tetrazole derivatives,^{31,32} none of the synthetic compounds in this study exhibited antifungal activity against the examined fungi at the tested concentrations which is similar to the study of Bekhit *et al.*³³ Moreover, the examined compounds failed to inhibit the growth of the tested Gram-positive and Gram-negative bacteria at the concentration up to 256 µg mL⁻¹.

Besides to antimicrobial activities,^{31,32} some of the tetrazole derivatives exhibit biological properties including anti-inflammatory, inhibition of cyclo-oxygenase, antidiabetic, anticonvulsant, and anticancer activities.^{34–36} Hence, further studies are required to investigate the other potential biological activities of these synthetic compounds.

4. Conclusion

We have demonstrated a simple method for the preparation of 5-substituted 1H-tetrazole derivatives using nano-TiCl₄.SiO₂ as eco-friendly and efficient catalyst in a one-pot procedure. Short reaction times, high yields, a clean process, simple methodology, easy work-up and green conditions are advantages of this protocol. Since some of the tetrazole derivatives previously showed antibacterial³⁰ and antifungal^{32,33} activities, further studies are still required for the design and synthesis of more novel tetrazole derivatives with better antimicrobial activities by this simple green method.

Supplementary material

The IR, ¹³C and ¹H NMR spectra of the novel 5-substituted

Table 2 Synthesis of 5-substituted 1H-tetrazole derivatives at reflux/DMF in the presence of nano-TiCl₄.SiO₂.^a

		$\text{ArCN} + \text{NaN}_3 \xrightarrow[\text{Reflux/DMF}]{\text{TiCl}_4 \cdot \text{SiO}_2} \text{Ar}-\text{C}_5\text{H}_3\text{N}_4$			
Entry	R (Ar)	Products	Yield/% ^b Time/h	MP/°C Found Reported ^[Ref]	
1	Ph		95 1.5	214–216	213–214 ^[15]
2	4-CH ₃ Ph		84 2.5	251–152	250–251 ^[15]
3	4-OHPh		82 2.5	231–233	234–235 ^[29]
4	3-NO ₂ Ph		92 1.5	217–220	219 ^[5]
5	4-ClPh		91 2	261–262	262–263 ^[30]
6	4-BrPh		90 2	267–269	267–269 ^[15]
7	PhCH ₂		78 3	122–123	123–124 ^[30]
8	4-OMePhCH ₂		81 3.5	232–233	231–232 ^[30]
9	4-ClPhCH ₂		84 3	260–261	–
10	(Ph) ₂ CH		88 2	164–166	164–165 ^[30]
11	3,4-diClPhCH ₂		84 2	128–130	–
12	Py		92 1.2	218–219	–

^a Reaction conditions: nitrile (1 mmol), NaN₃ (2 mmol), (0.1 g), nano-TiCl₄.SiO₂, DMF (5 mL) at reflux.^b Isolated yields.

1H-tetrazole derivatives (Table 2, compounds 9, 11 and 12) are presented in the online supplement.

Acknowledgements

The Research Council of Yazd University is gratefully acknowl-

edged for the financial support of this project. The antifungal activities of the synthetic compounds were evaluated in Shiraz University of Medical Sciences (grant number 90-01-43-3451).

References

- 1 S. Wittenberger, Recent developments in tetrazole chemistry. A review, *J. Org. Prep. Proced.*, 1994, **26**, 449–531.
- 2 R.N. Bulter, Tetrazoles, reference module in chemistry, molecular sciences and chemical engineering, *Com. Heterocyclic Chem.*, 1996, **4**, 621–678.
- 3 R.E. Ford, P. Knowles, E. Lunt, S.M. Marshall, A.J. Penrose, C.A. Ramsden, A.J.H.S.J.L. Walker and D.E. Wright, Synthesis and quantitative structure-activity relationships of antiallergic 2-hydroxy-N-(1H-tetrazol-5-yl)benzamides and N-(2-hydroxyphenyl)-1H-tetrazole-5-carboxamides, *J. Med. Chem.*, 1986, **29**, 538–549.
- 4 N. P. Peet, L.E. Baugh, S. Sundler, J.E. Lewis, E.H. Matthews, E.L. Olberding and D.N. Shah, 3-(1H-Tetrazol-5-yl)-4(3H)-quinazolinone sodium salt (MDL 427): a new antiallergic agent, *J. Med. Chem.*, 1986, **29**, 2403–2409.
- 5 L.V. Myznikov, A. Hrabalek and G. I. Koldobskii, Drugs in the tetrazole series. (Review), *Chem. Heterocyclic Chem.*, 2007, **43**, 1–9.
- 6 M. J. Schocken, R.W. Creekmore, G. Theodoridis, G.J. Nystrom and R.A. Robinson, Microbial transformation of the tetrazolinone herbicide F5231, *Appl. Environmen. Microb.*, 1989, **55**, 1220–1222.
- 7 S.J. Lim, Y. Sunohara, and H. Matsumoto, Action of fentrazamide on protein metabolism and cell division in plants, *J. Pest. Science*, 2007, **32**, 249–254.
- 8 P. Rhonstad and D. Wensbo, On the relative strength of the 1H-tetrazol-5-yl- and the 2-(triphenylmethyl)-2H-tetrazol-5-yl-group in directed ortho-lithiation, *Tetrahedron Lett.*, 2002, **43**, 3137–3139.
- 9 R.J. Herr, 5-Substituted-1H-tetrazoles as carboxylic acid isosteres: medicinal chemistry and synthetic methods, *Bioorg. Med. Chem.*, 2002, **10**, 3379–3393.
- 10 T. Ichikawa, T. Kizakazi, Y. Matsushita, H. Hosono, M. Yamada, M. Mizuno and K. Itoh, Optically active antifungal azoles. XI. An alternative synthetic route for 1-[(1R, 2R)-2-(2,4-Difluorophenyl)-2-hydroxy-1-methyl-3-(1H-1,2,4-triazol-1-yl)propyl]-3-[4-(1H-1-tetrazolyl)phenyl]-2-imidazolidinone (TAK-456) and its analog, *Chem. Pharma. Bull.*, 2000, **48**, 1947–1953.
- 11 A.R. Modarresi-Alam and M. Nasrollahzadeh, Synthesis of 5-aryl-amino-1H (2H)-tetrazoles and 5-amino-1-aryl-1H-tetrazoles from secondary arylcyanamides in glacial acetic acid: a simple and efficient method, *Turkish J. Chem.*, 2009, **33**, 267–270.
- 12 F. Himo, Z.P. Demko, L. Noodleman and K.B. Sharpless, Mechanisms of tetrazole formation by addition of azide to nitriles, *JACS*, 2002, **124**, 12210–12216.
- 13 D. Amantini, R. Beleggia, F. Fringuelli, F. Pizzo and L. Vaccaro, TBAF-catalyzed synthesis of 5-substituted 1H-tetrazoles under solventless conditions, *J. Org. Chem.*, 2004, **69**, 2896–2898.
- 14 O. Marvi, A.H. Alizadeh and S. Zarrabi, Montmorillonite K-10 clay as an efficient reusable heterogeneous catalyst for the solvent-free microwave mediated synthesis of 5-substituted 1H-tetrazoles, *Bull. Korean Chem. Soc.*, 2011, **32**, 4001–4004.
- 15 T. Jin, F. Kitahara, S. Kamijo and Y. Yamamoto, Copper-catalyzed synthesis of 5-substituted 1H-tetrazoles via the [3+2] cycloaddition of nitriles and trimethylsilyl azide, *Tetrahedron Lett.*, 2008, **49**, 2824–2827.
- 16 M. Nasrollahzadeh, Y. Bayat, D. Habibi and S. Mosharef, FeCl₃.SiO₂ as a reusable heterogeneous catalyst for the synthesis of 5-substituted 1H-tetrazoles via [2+3] cycloaddition of nitriles and sodium azide, *Tetrahedron Lett.*, 2009, **50**, 4435–4438.
- 17 A. Teimouri and A. Najafi Chermahini, Zeolite and sulfated zirconia as catalysts for the synthesis of 5-substituted 1H-tetrazoles via [2+3] cycloaddition of nitriles and sodium azide, *Polyhedron*, 2011, **30**, 2606–2610.
- 18 J. He, B. Li, F. Chen, Z. Xu and G. Yin, Tungstates: Novel heterogeneous catalysts for the synthesis of 5-substituted 1H-tetrazoles, *J. mol. Catal. A.*, 2009, **304**, 135–138.
- 19 S.M. Sajadi, M. Naderi and S. Babadoust, Nano TiO₂ as an efficient and reusable heterogeneous catalyst for the synthesis of 5-substituted 1H-tetrazoles, *J. Nat. Sci. Res.*, 2011, **1**, 10–14.

- 20 S.M. Sajadi, $\text{MoO}_3\text{-SiO}_2$ as an efficient and reusable heterogeneous catalyst for the synthesis of 5-substituted 1H-tetrazoles, *Chem. Mater. Res.*, 2012, **2**, 12–16.
- 21 Clinical and Laboratory Standards Institute (CLSI), Performance standards for antimicrobial disk susceptibility tests; approved standard, 9th edn, Clinical and Laboratory Standards Institute, Wayne, PA, CLSI M2-A9, 2006.
- 22 Clinical and Laboratory Standards Institute (CLSI), Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically; Approved Standard, 7th edn, Wayne, PA: Clinical and Laboratory Standards Institute, CLSI M7-A7, 2006.
- 23 Clinical and Laboratory Standards Institute (CLSI), Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi; Approved Standard, Clinical and Laboratory Standards Institute, Wayne, PA, CLSI M38-A, 2006.
- 24 Clinical and Laboratory Standards Institute. (CLSI), Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Approved Standard, Clinical and Laboratory Standards Institute, Wayne, PA, CLSI document M27-A7, 2006.
- 25 B.F. Mirjalili and L. Zamani, Nano- $\text{TiCl}_4\text{-SiO}_2$: A versatile and efficient catalyst for synthesis of dihydropyrimidones via Biginelli condensation, *S. Afr. J. Chem.*, 2014, **67**, 21–26.
- 26 B.F. Mirjalili, A. Bamoniri and L. Zamani, One-pot synthesis of 1,2,4,5-tetrasubstituted imidazoles promoted by Nano- $\text{TiCl}_4\text{-SiO}_2$, *Scientia Iranica*, 2012, **19**, 565–569.
- 27 B.F. Mirjalili, A. Bamoniri and L. Zamani, Nano- $\text{TiCl}_4\text{-SiO}_2$: An Efficient and Reusable Catalyst for the Synthesis of Tetrahydrobenzo[a]xanthenes-11-ones, *Lett. Org. Chem.*, 2012, **9**, 338–343.
- 28 L. Zamani, B.F. Mirjalili, K. Zomorodian, M. Namazian, S. Khabnabideh and E.F. Mirzaei, Synthesis of benzimidazoles in the presence of nano- $\text{TiCl}_4\text{-SiO}_2$ as antifungal agents and tautomerism theoretical study of some products, *Farmacia*, 2014, **62**, 467–482.
- 29 B. Sreedhar, A.S. Kumar and D. Yada, CuFe_2O_4 nanoparticles: a magnetically recoverable and reusable catalyst for the synthesis of 5-substituted 1H-tetrazoles, *Tetrahedron Lett.*, 2011, **52**, 3365–3369.
- 30 H. M. Nanjundaswamy and H. Abrahamse, Green synthesis of 5-substituted-1H-1,2,3,4-tetrazoles and 1-substituted-1H-1,2,3,4-tetrazoles via [3+2] cycloaddition by reusable immobilized AlCl_3 on $\gamma\text{-Al}_2\text{O}_3$, *Heterocycles*, 2014, **89**, 2137–2150.
- 31 D. Varadaraji, S.S. Suban, V.R. Ramasamy, K. Kubendiran, J.S.K.G. Raguraman, S.K. Nalilu and H.N. Pati, Synthesis and evaluation of a series of 1-substituted tetrazole derivatives as antimicrobial agents, *Org. Commun.*, 2012, **3**, 45–46.
- 32 R.S. Upadhyaya, S. Jain, N. Sinha, N. Kishore, R. Chandra and S.K. Arora, Synthesis of novel substituted tetrazoles having antifungal activity, *Eur. J. Med. Chem.*, 2004, **39**, 579–592.
- 33 A.A. Bekhit, O.A. El-Sayed, E. Aboulmagd and J.Y. Park, Tetrazolo [1,5-a]quinoline as a potential promising new scaffold for the synthesis of novel anti-inflammatory and antibacterial agents, *Eur. J. Med. Chem.*, 2004, **39**, 249–255.
- 34 S.R. Pattan, P. Kekare, A. Patil, A. Nikalge and B.S. Kittur, Studies on the synthesis of novel 2,4-thiazolidinedione derivatives with anti-diabetic activity, *IJPS*, 2009, **5**, 225–230.
- 35 A. Rajasekaran and K.A. Rajagopal, Synthesis of some novel triazole derivatives as anti-nociceptive and anti-inflammatory agents, *Acta Pharm.*, 2009, **59**, 355–364.
- 36 S.A. Rostom, H.M. Ashour, H.A.E. Razik, F.E.F. Ael and N.N. El-Din, Azole antimicrobial pharmacophore-based tetrazoles: synthesis and biological evaluation as potential antimicrobial and anticonvulsant agents, *Bioorg. Med. Chem.*, 2009, **17**, 2410–2422.

Supplementary Information, *S. Afr. J. Chem.* 68: 133–137

Synthesis and Characterization of 5-Substituted 1*H*-tetrazoles in the Presence of Nano-TiCl₄.SiO₂

L. Zamani^{a,*}, B. F. Mirjalili^a, K. Zomorodian^b and S. Zomorodian^b

^aDepartment of Chemistry, College of Science, Yazd University, Yazd, P.O.Box 8915813149, Iran.

^bCenter of Basic Researches in Infectious Diseases and Department of Medical Mycology and Parasitology School of Medicine, Shiraz University of Medical Sciences, Post code 71348-45794, Shiraz, Iran.

*E-mail: L.zamani2008@gmail.com, L.zamani@stu.ayzd.ac.ir

Here, we describe the IR, ¹³C and ¹H NMR spectra of the novel 5-substituted 1*H*-tetrazole derivatives (Table 2, compounds 9, 11 and 12).

TABLE OF CONTENTS

Figure S1. FT-IR of 5-Phenyl-1*H*-tetrazole

Figure S2. ¹H NMR of 5-Phenyl-1*H*-tetrazole

Figure S3. FT-IR of 5-(4-Methylphenyl)-1*H*-tetrazole

Figure S4. ¹H NMR of 5-(4-Methylphenyl)-1*H*-tetrazole

Figure S5. ¹H NMR of 5-(4-Methylphenyl)-1*H*-tetrazole (expand)

Figure S6. FT-IR of 5-(4-Hydroxyphenyl)-1*H*-tetrazole

Figure S7. ¹H NMR of 5-(4-Hydroxyphenyl)-1*H*-tetrazole

Figure S8. ¹H NMR of 5-(4-Hydroxyphenyl)-1*H*-tetrazole (expand)

Figure S9. FT-IR of 5-(3-Nitrophenyl)-1*H*-tetrazole

Figure S10. ¹H NMR of 5-(3-Nitrophenyl)-1*H*-tetrazole

Figure S11. ¹H NMR of 5-(3-Nitrophenyl)-1*H*-tetrazole (expand)

Figure S12. FT-IR of 5-(4-Chlorophenyl)-1*H*-tetrazole

Figure S13. ¹H NMR of 5-(4-Chlorophenyl)-1*H*-tetrazole

Figure S14. ¹H NMR of 5-(4-Chlorophenyl)-1*H*-tetrazole (expand)

Figure S15. FT-IR of 5-(4-Bromophenyl)-1*H*-tetrazole

Figure S16. ¹H NMR of 5-(4-Bromophenyl)-1*H*-tetrazole

Figure S17. ¹H NMR of 5-(4-Bromophenyl)-1*H*-tetrazole (expand)

Figure S18. FT-IR of 5-Benzyltetrazole

Figure S19. ^1H NMR of 5-Benzyltetrazole

Figure S20. ^1H NMR of 5-Benzyltetrazole (expand)

Figure S21. FT-IR of 5-((4-Methoxyphenyl)methyl)tetrazole

Figure S22. ^1H NMR of 5-((4-Methoxyphenyl)methyl)tetrazole

Figure S23. ^1H NMR of 5-((4-Methoxyphenyl)methyl)tetrazole (expand)

Figure S24. FT-IR of 5-((4-Chlorophenyl)methyl)tetrazole

Figure S25. ^1H NMR of 5-((4-Chlorophenyl)methyl)tetrazole

Figure S26. ^1H NMR of 5-((4-Chlorophenyl)methyl)tetrazole (expand)

Figure S27. ^{13}C NMR of 5-((4-Chlorophenyl)methyl)tetrazole

Figure S28. FT-IR of 5-Benzydryltetrazole

Figure S29. ^1H NMR of 5-Benzydryltetrazole

Figure S30. ^1H NMR of 5-Benzydryltetrazole (expand)

Figure S31. FT-IR of 5-((3,4-dichlorophenyl)methyl)tetrazole

Figure S32. ^1H NMR of 5-((3,4-dichlorophenyl)methyl)tetrazole

Figure S33. ^1H NMR of 5-((3,4-dichlorophenyl)methyl)tetrazole (expand)

Figure S34. ^{13}C NMR of 5-((3,4-dichlorophenyl)methyl)tetrazole

Figure S35. FT-IR of 4-(1*H*-tetrazole-5-yl)pyridine

Figure S36. ^1H NMR of 4-(1*H*-tetrazole-5-yl)pyridine

Figure S37. ^1H NMR of 4-(1*H*-tetrazole-5-yl)pyridine (expand)

Figure S38. ^{13}C NMR of 4-(1*H*-tetrazole-5-yl)pyridine

5-Phenyl-1*H*-tetrazole

Yield: 95%, White crystal.

Figure S1. FT-IR: $\bar{\nu}$ (KBr) = 2600-300, 1608, 1563, 1485, 1465, 1409, 1480, 726, 687 cm^{-1} .

Figure S2. ^1H NMR (400 MHz, DMSO- d_6): 8.01 (brs, 2H), 7.51 (brs, 3H) ppm.

Figure S1. FT-IR (KBr) 5-Phenyl-1*H*-tetrazole

Figure S2. ^1H NMR (500 MHz, DMSO- d_6) 5-Phenyl-1*H*-tetrazole

5-(4-Methylphenyl)-1*H*-tetrazole

Yield: 84%, White crystal.

Figure S3. FT-IR: $\bar{\nu}$ (KBr) = 2600-300, 1608, 1563, 1485, 1465, 1409, 1480, 726, 687 cm^{-1} .

Figure S4, S5. ^1H NMR (500 MHz, CDCl_3): 8.81 (d, $J = 5.8$, 2H), 8.09 (d, $J = 5.8$, 2H), 2.5 (s, 3H) ppm.

Figure S3. FT-IR (KBr) 5-(4-Methylphenyl)-1*H*-tetrazole

Figure S4: ¹H NMR (500 MHz, CDCl₃) 5-(4-Methylphenyl)-1*H*-tetrazole

Figure S5: ¹H NMR (500 MHz, CDCl₃) 5-(4-Methylphenyl)-1*H*-tetrazole (expand)

5-(4-Hydroxyphenyl)-1*H*-tetrazole

Yield: 82%, White crystal.

Figure S6. FT-IR: $\bar{\nu}$ (KBr) = 2500-3400, 1648, 1600, 1515, 1470, 1435, 1080, 842 cm^{-1} .

Figure S7, S8. ^1H NMR (400 MHz, DMSO- d_6): 16.5 (brs, NH), 10.17 (s, 1H), 7.84 (d, $J=8$, 2H), 6.93 (d, $J=7.6$, 2H) ppm.

Figure S6: FT-IR (KBr) 5-(4-Hydroxyphenyl)-1*H*-tetrazole

Figure S7. ^1H NMR (500 MHz, DMSO-d_6) 5-(4-Hydroxyphenyl)-1H-tetrazole

Figure S8. ^1H NMR (500 MHz, DMSO-d_6) 5-(4-Hydroxyphenyl)-1H-tetrazole (expand)

5-(3-Nitrophenyl)-1*H*-tetrazole

Yield: 92%, White crystal.

Figure S9. FT-IR: $\bar{\nu}$ (KBr) = 2600-3300, 1626, 1569, 1528, 1349, 872, 743, 973 cm^{-1} .

Figure S10, S11. ^1H NMR (400 MHz, DMSO- d_6): 8.82 (s, 1H), 8.43 (dd, J = 7.6 and 8 Hz, 2H), 7.89 (t, J = 8.4, 1H) ppm.

Figure S9: FT-IR (KBr) 5-(3-Nitrophenyl)-1*H*-tetrazole

Figure S10. ¹H NMR (500 MHz, DMSO-d₆) 5-(3-Nitrophenyl)-1*H*-tetrazole

Figure S11. ¹H NMR (500 MHz, DMSO-d₆) 5-(3-Nitrophenyl)-1*H*-tetrazole (expand)

5-(4-Chlorophenyl)-1*H*-tetrazole

Yield: 91%, White crystal.

Figure S12. FT-IR: $\bar{\nu}$ (KBr) = 2500-3000, 1654, 1610, 1561, 1487, 1434, 831 cm^{-1} .

Figure S13, S14. ^1H NMR (500 MHz, DMSO- d_6): 8.10 (d, J = 10.53, 2H), 7.69 (d, J = 8.41, 2H) ppm.

Figure S12: FT-IR (KBr) 5-(4-Chlorophenyl)-1*H*-tetrazole

Figure S13. ^1H NMR (500 MHz, DMSO-d_6) 5-(4-Chlorophenyl)-1*H*-tetrazole

Figure S14. ^1H NMR (500 MHz, DMSO-d_6) 5-(4-Chlorophenyl)-1*H*-tetrazole (expand)

5-(4-Bromophenyl)-1*H*-tetrazole

Yield: 90%, White crystal

Figure S15. FT-IR: $\bar{\nu}$ (KBr) = 2600-300, 1649, 1604, 1560, 1482, 1431, 1053, 829 cm^{-1} .

Figure S16, S17. ^1H NMR (500 MHz, CDCl_3): 8.06 (d, $J=7.2$, 2H), 7.69 (d, $J=7.2$, 2H) ppm.

Figure S15: FT-IR (KBr) 5-(4-Bromophenyl)-1*H*-tetrazole

Figure S16. ^1H NMR ((500 MHz, CDCl_3): 5-(4-Bromophenyl)-1H-tetrazole

Figure S17. ^1H NMR ((500 MHz, CDCl_3): 5-(4-Bromophenyl)-1H-tetrazole (expand)

5-Benzyltetrazole

Yield: 78%, White crystal.

Figure S18. FT-IR: $\bar{\nu}$ (KBr) = 2400-300, 1592, 1549, 1496, 1248, 775 cm^{-1} .

Figure S19, S20. ^1H NMR (400 MHz, DMSO-d_6): 4.26 (s, 2H, CH_2), 7.25-1.31 (m, 5H) ppm.

Figure S18: FT-IR (KBr) 5-Benzyltetrazole

Figure S19. ¹H NMR (500 MHz, DMSO-d₆) 5-Benzyltetrazole

Figure S20. ¹H NMR (500 MHz, DMSO-d₆) 5-Benzyltetrazole (expand)

5-((4-Methoxyphenyl)methyl)tetrazole

Yield: 81%, White crystal.

Figure S21. FT-IR: $\bar{\nu}$ (KBr) = 2600-3400, 1636, 1514, 1124, 848 cm^{-1} .

Figure S22, S23. ^1H NMR (400 MHz, DMSO- d_6): 7.19 (brs, 2H), 6.86 (brs, 2H), 4.18 (s, 2H), 3.69 (s, 3H) ppm.

Figure S21: FT-IR (KBr) 5-((4-Methoxyphenyl)methyl)tetrazole

Figure S22. ^1H NMR (500 MHz, DMSO-d_6) 5-((4-Methoxyphenyl)methyl)tetrazole

Figure S23. ^1H NMR (500 MHz, DMSO-d_6) 5-((4-Methoxyphenyl)methyl)tetrazole (expand)

5-((4-Chlorophenyl)methyl)tetrazole

Yield: 84%, White crystal.

Figure S24. FT-IR: $\bar{\nu}$ (KBr) = 2600-300, 1538, 1492, 1407, 1263, 1207, 834 cm^{-1} .

Figure S25, S26. ^1H NMR (400 MHz, DMSO- d_6): 7.40 (m, 2H), 7.31 (d, $J = 8$, 2H), 4.30 (s, 2H) ppm.

Figure S27. ^{13}C -NMR (125 MHz, DMSO) δ = 155.0, 132.5, 128.9, 124.7, 123.6, 28.8 ppm.

Figure S24: FT-IR (KBr) 5-((4-Chlorophenyl)methyl)tetrazole

Figure S25. ^1H NMR (500 MHz, DMSO- d_6) 5-((4-Chlorophenyl)methyl)tetrazole

Figure S26. ^1H NMR (500 MHz, DMSO- d_6) 5-((4-Chlorophenyl)methyl)tetrazole (expand)

Figure S27. ¹³C NMR (500 MHz, DMSO) of 5-((4-chlorophenyl)methyl)tetrazole

5-Benzyhydriyltetrazole

Yield: 88%, White crystal.

Figure S28. FT-IR: $\bar{\nu}$ (KBr) = 2600-300, 1567, 1496, 1245, 745 cm^{-1} .

Figure S29, S30. ^1H NMR (500 MHz, CDCl_3): 5.82 (s, 1H), 8.128 (brs, 1H), 7.25-7.41 (m, 10H) ppm.

Figure S28. FT-IR (KBr) of 5-Benzyhydriyltetrazole

Figure S29. ^1H NMR (500 MHz, CDCl_3) of 5-Benzyhydriyltetrazole

Figure S30. ^1H NMR (500 MHz, CDCl_3) of 5-Benzyhydriyltetrazole (expand)

5-((3,4-dichlorophenyl)methyl)tetrazole

Yield: 84%, White crystal.

Figure S31. FT-IR: $\bar{\nu}$ (KBr) = 2500-3300, 1560, 1472, 1440, 1260, 1210, 827, 766, 706, 674 cm^{-1} .

Figure S32, S33. ^1H NMR (500 MHz, CDCl_3): 7.61 (d, J = 8.45, 2H, 2H), 7.28 (d, J = 8.2, 1H), 4.32 (s, 2H) ppm.

Figure S34. ^{13}C -NMR (125 MHz, DMSO) δ = 28.8, 130.2, 131.6, 131.8, 174.8 ppm

Figure S31: FT-IR (KBr) 5-((3,4-dichlorophenyl)methyl)tetrazole

Figure S32. ^1H NMR (500 MHz, CDCl_3) 5-((3,4-dichlorophenyl)methyl)tetrazole

Figure S33. ^1H NMR (500 MHz, CDCl_3) 5-((3,4-dichlorophenyl)methyl)tetrazole (expand)

Figure S34. ^{13}C NMR (500 MHz, DMSO) 5-((3,4-dichlorophenyl)methyl)tetrazole

4-(1*H*-tetrazole-5-yl)pyridine (table 2, entry 12)

Yield: 92%, White crystal.

Figure S35. FT-IR: $\bar{\nu}$ (KBr) = 2500-3000, 1631, 1529, 1440, 1338, 1292, 1042, 990, 846, 751 cm^{-1} .

Figure S36, S37. ^1H NMR (500 MHz, CDCl_3): 8.00 (d, $J = 7.89$, 2H), 7.40 (d, $J = 7.86$, 2H) ppm.

Figure S38. ^{13}C -NMR (125 MHz, DMSO) $\delta = 127.9$, 130.7 ppm.

Figure S35. FT-IR (KBr) 4-(1*H*-tetrazole-5-yl)pyridine

Figure S36. ¹H NMR (500 MHz, CDCl₃) 4-(1*H*-tetrazole-5-yl)pyridine

Figure S37. ¹H NMR (500 MHz, CDCl₃) 4-(1*H*-tetrazole-5-yl)pyridine (expand)

Figure S38. ¹³C NMR (500 MHz, DMSO) 4-(1*H*-tetrazole-5-yl)pyridine

FT-IR, ^1H NMR and ^{13}C NMR Elucidation of 5-((3,4-dichlorophenyl)methyl)tetrazole

Characterization of 5-((3,4-dichlorophenyl)methyl)tetrazole was completed using FT-IR, ^1H NMR and ^{13}C NMR. The marked structure of 5-((3, 4-dichlorophenyl) methyl) tetrazole is showed in Figure 1.

Figure 1

As can be seen in the IR spectrum, the stretching frequencies of C-H and N-H groups are indicated at 2500-3300 cm^{-1} , The stretching frequency of C=C group is demonstrated at 1560 cm^{-1} , The frequency absorption of tetrazole ring is specified at 1472 cm^{-1} , The stretching frequency of C-H benzyl group at 1260 cm^{-1} and the bending frequency of C-H phenyl ring is appeared at 827, 766, 706 cm^{-1} (Figure 1).

In the ^1H NMR spectrum, the appearance of the methylene protons (Hc, figure 1) as singlet at 4.32 ppm, integrating to two. The signal related to the one aromatic proton (Hh, figure 1) appear as a doublet at 7.28 ppm, integrating to one. In the appearance of a doublet at 7.61 ppm due to the two aromatic protons (Hg and Hd, figure 1), integrating to two, This signal appears at up field due to the deshielding nature of the neighbouring chlorine atoms.

The ^{13}C NMR spectrum for 5-((3,4-dichlorophenyl)methyl) tetrazole displays signals characteristic various carbones (Cb, Cd, Cg, Ch, Cc, figure 1) at 28.8, 130.2, 131.6, 131.8, 174.8 ppm, respectively.

References

1. H. M. Nanjundaswamy, and H. Abrahamse, *Hetrocycles*, 2014, **89**, 2137-2150.
2. T. Jin, F. Kitahara, S. Kamijo, and Y. Yamamoto, *Tetrahedron Lett.*, 2008, **49**, 2824-2827.
3. B. Sreedhar, A. Suresh Kumar, and D. Yada, *Tetrahedron Lett.*, 2011, **52**, 3565-3569.
4. L. V. Myznikov, A. Hrabalek, and G. I. Koldobskii, *Chem. Hetrocyclic Chem.*, 2007, **43**, 1-9.