

## **Efficacy of *Tragopogon graminifolius* medicinal powder as an inulin source for laying hens**

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(Submitted 11 February2022; Accepted 7 April 2022; Published 13 May 2022)

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### **Abstract**

In an experiment, the efficacy of *Tragopogon graminifolius* (TG) plant powder as a medicinal supplement for laying hens was investigated. A total of 192 Hy-line (W36) hens that had been laying for 52 to 63 weeks, were used. They were allocated to four replicates of four treatments (12 birds per in each replicate) in a completely randomized design. Diets were formulated to contain 0%, 2%, 4% and 6% TG powder. Dietary augmentation with TG powder affected the production and blood indexes of laying hens significantly. Egg weight, egg production percentage, egg mass and feed intake were greatest and feed conversion ratio, high yolk colour index, and Haugh unit were best when the hens were supplemented with 6% TG powder. Eggshell thickness decreased when this high level of TG powder was used. The lowest blood white cell count was observed with 2% TG in the diet. The TG levels did not change the blood indexes meaningfully. Based on these results, the provision of TG to laying hens at up to 6% of their diet improved the production traits.

**Keywords:** egg production, egg yolk, feed intake, haematology

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### **Introduction**

Feeding organic products to animals is becoming more acceptable to consumers and the use of herbal supplements may make animal products more healthful (Torki *et al.*, 2021). Recently, the use of medicinal plants in poultry diets has increased (Darabighane *et al.*, 2017; Gadde *et al.*, 2017). they are added for a variety of reasons, including antimicrobial and antioxidant activities, growth promotion, reducing the use of pharmaceutical and chemical additives, improved bird health, increased beneficial properties of the poultry products, and appeal to customers (Oluwafemi *et al.*, 2020; Pliego *et al.*, 2020).

*Tragopogon graminifolius* is a valuable source of inulin (Falahi *et al.*, 2019), which acts as a prebiotic because of its beneficial nutritional effects and is widely used in the food industry worldwide (Birmani *et al.*, 2019; Stolzenbach *et al.*, 2020). Dietary supplementation with inulin improved egg production and eggshell thickness of laying hens (Shang *et al.*, 2020b; Bucław, 2017). Bell *et al.* (2017) recommended supplementation of the ration for laying hens with 1.0% of inulin powder as it increased egg production and shell thickness. Shang *et al.* (2020a) concluded that supplementation of laying hen with inulin also prolonged the shelf-life of eggs.

In layers and broilers, medicinal plants do not merely stimulate appetite and digestion, but can influence other physiological functions, help to sustain good health and welfare, and improve their performance (Torki *et al.*, 2021). Ghasemi Shansabadi *et al.* (2015) indicated that low-density thyme powder in the rations of Japanese quails increased their growth, prevented oxidation of fats, and was a highly effective antioxidant. The use of *Silybum marianum* L. powder at up to 2% in layer diets upgraded their performance, egg traits, blood biochemicals, and immunity significantly (Nobakht, 2015b). As an inulin source, the use of *Cichorium intybus* L powder as a medicinal additive for layers at rates up to 3% was

shown to improve their performance, egg yolk colour index, Haugh unit, and blood cell immunity (Nobakht, 2015a).

The current experiment aimed to examine the effects of various levels of *Tragopogon graminifolius* medicinal plant powder as a source of inulin on egg traits, and blood indexes of laying hens.

## Materials and Methods

The Animal Welfare Committee of Islamic Azad University (Maragheh Branch) approved the animal care protocol used in this experiment (Protocol no. 1396-IAU. 21.05.2017). A total of 192 laying hens between the ages of 52 and 63 weeks were allotted to four replications of four treatments (12 hens in each replicate) in a completely randomized design. The treatments consisted of 0% (control), 2%, 4% and 6% of TG powder provided as a supplement to the usual ration of the hens. Feed and water were provided ad libitum. UFFDA software (User-friendly Feed Formulation Done Again) (University of Georgia, USA) was used for diet formulation. Diets were formulated as recommended nutrient requirements of the Hy-Line (W36) strain (Table 1).

**Table 1** Feed ingredients and chemical composition of diets for laying hens that incorporated *Tragopogon graminifolius* medicinal powder

Feed ingredients. %	Level of <i>Tragopogon graminifolius</i> medicinal plant, %			
	0	2	4	6
Corn	50.	50.	50.	50.
Wheat	17.92	15.11	12.55	10.00
Soybean meal	19.74	19.78	19.83	19.85
Soybean oil	2.65	3.20	3.75	4.31
<i>Tragopogon graminifolius</i>	0	2	4	6
Oyster shell	7.37	7.39	7.41	7.44
Bone powder	1.75	1.69	1.63	1.56
Sodium chloride	0.31	0.31	0.30	0.30
Vitamin premix <sup>1</sup>	0.25	0.25	0.25	0.25
Mineral premix <sup>2</sup>	0.25	0.25	0.25	0.25
DL-Methionine	0.01	0.02	0.03	0.04
Calculated composition				
Metabolizable energy, kcal/kg	2900	2900	2900	2900
Crude protein, %	14.50	14.50	13.78	13.78
Calcium, %	3.40	3.40	3.40	3.40
Available phosphorus, %	0.33	0.33	0.33	0.33
Crude fibre, %	2.79	3.30	3.82	4.31
Sodium, %	0.16	0.16	0.16	0.16
Lysine, %	0.70	0.70	0.70	0.70
Methionine + cysteine, %	0.56	0.56	0.56	0.56
Tryptophan, %	0.18	0.18	0.18	0.18

<sup>1</sup> Vitamin A: 2.7 mg, vitamin D<sub>3</sub>: 0.05 mg, vitamin E: 18 mg, vitamin K<sub>3</sub>: 2 mg, thiamine: 1.8 mg, riboflavin: 6.6 mg, pantothenic acid: 10 mg, pyridoxine: 3 mg, cyanocobalamin: 0.015 mg, niacin: 30 mg, biotin: 0.1 mg, folic acid: 1 mg, choline chloride: 250 mg, antioxidant: 100 mg per kg of diet

<sup>2</sup> Ferrous sulfate: 50 mg, manganese sulfate: 100 mg; zinc oxide: 100 mg; copper sulfate: 10 mg, potassium iodide: 1 mg, sodium selenite: 0.2 mg per kg of diet

During the experiment the laying hens were kept on a regime of 16 hours of light and 8 hours of darkness. Temperature in the poultry house was set automatically between 18 °C and 24 °C. Eggs were collected daily, and data for feed intake, egg production percentage, egg mass, egg weight and feed conversion ratio were determined weekly. Mortality was recorded when it occurred, and hen days were determined at the end of each week based on the number of surviving hens. The eggs were classified as normal or damaged. Damaged eggs included fully cracked eggs, defined as an egg with a broken shell and destroyed membrane, eggs with a hairline crack, defined as an egg with broken shell, but intact membrane, and eggs without a shell, but with an intact membrane.

At the end of the experiment, three eggs were collected from each replicate and the traits of egg quality were determined. Specific gravity was determined by floating the egg in various concentrations of salt water (Farkhoy et al., 1994). After the internal contents of the eggs had been emptied, eggshells were cleaned and kept at room temperature for 48 hours, then dried and weighed, and the average thickness was recorded. The colour of the yoke was determined with the Roche yolk colour fan (ORKA Food Technology, LLC., West Bountiful, Utah, USA), and the yolk index, egg albumin index, and Haugh units were determined, as described by Farkhoy et al. (1994). At the end of the experiment, two birds from each replicate were randomly chosen for blood collection. After the birds had been fasted for approximately six hours, 5 mL blood samples were collected from the brachial vein. One mL of the blood was transferred to tubes with EDTA for haematological analysis, including the numbers of red and white blood cells, haemoglobin concentration, packed cell volume, and the number of lymphocytes (Gross & Sigel, 1983). The remaining 4 mL of blood was centrifuged to obtain serum for biochemical analysis, including the concentrations of cholesterol, triglyceride, albumin, total protein, and uric acid. Kits for the Anision-300 auto-analyser system (Pars Azmoon Company, Tehran, Iran) were used in the analysis of blood biochemistry (Nazifi, 1997).

The results were calculated with the general linear model procedure of SAS (SAS Institute, Inc., Cary, North Carolina, USA). The linear model was:

$$Y_{ij} = \mu + T_i + \varepsilon_{ij}$$

where:  $Y_{ij}$  = a response variable for the  $i$ th treatment in the  $j$ th replicate,  $\mu$  = overall mean,  $T_i$  = the effect of the level of *Tragopogon graminifolius* powder in the diet, and  $\varepsilon_{ij}$  = the residual deviation of the observation from the effect in the model.

## Results and Discussion

The effects of *TG* on egg productivity of hens are shown in Table 2. Increasing levels of *TG* improved all of these measures of productivity ( $P < 0.01$ ). The highest values of egg weight, egg production, egg mass, and feed intake, and the best feed conversion ratio were observed when 6% *TG* was used in the diet.

**Table 2** Effects of dietary augmentation with *Tragopogon graminifolius* plant powder on the performance of laying hens

Dietary level, %	Egg weight, g	Productivity, %	Daily production, g	Feed intake, g	Feed conversion ratio
0	60.71 <sup>d</sup>	56.51 <sup>c</sup>	34.30 <sup>d</sup>	110.11 <sup>b</sup>	3.21 <sup>a</sup>
2	60.93 <sup>c</sup>	57.99 <sup>c</sup>	35.34 <sup>c</sup>	110.43 <sup>ab</sup>	3.12 <sup>b</sup>
4	61.39 <sup>b</sup>	60.61 <sup>b</sup>	37.21 <sup>b</sup>	110.71 <sup>a</sup>	2.98 <sup>c</sup>
6	61.66 <sup>a</sup>	62.49 <sup>a</sup>	38.53 <sup>a</sup>	110.72 <sup>a</sup>	2.87 <sup>d</sup>
SE	0.04	0.47	0.28	0.10	0.02
P-value	0.0001	0.0001	0.0001	0.0088	0.0001

<sup>a,b,c,d</sup> Within a column, means followed by a common superscript were not different with probability  $P = 0.05$

Table 3 shows the results of the use of *TG* powder on egg quality. Supplementation with up to 6% *TG* powder improved the yolk colour and Haugh unit significantly and reduced shell thickness ( $P < 0.01$ ), whereas *TG* powder had no significant effect on shell percentage, egg white percentage, and egg yolk percentage ( $P > 0.05$ ).

**Table 3** Effects of dietary augmentation with *Tragopogon graminifolius* plant powder on the quality of eggs produced by laying hens

Medicinal plant	Yolk colour	Shell weight, %	White weight, %	Yolk weight, %	Haugh unit	Shell thickness, mm
0	2.00 <sup>c</sup>	10.62	63.19	26.20	73.20 <sup>c</sup>	0.34 <sup>a</sup>
2	3.11 <sup>b</sup>	10.34	63.13	26.51	73.92 <sup>b</sup>	0.33 <sup>b</sup>
4	4.66 <sup>a</sup>	10.80	62.37	26.84	74.55 <sup>a</sup>	0.33 <sup>b</sup>
6	4.67 <sup>a</sup>	11.46	60.85	27.71	74.77 <sup>a</sup>	0.33 <sup>b</sup>
SE	0.02	0.36	0.84	0.74	0.12	0.01
P-value	0.0001	0.2415	0.2457	0.5395	0.0001	0.0086

<sup>a,b,c</sup> Within a column, means followed by a common superscript were not different with probability  $P = 0.05$

Tables 4 and 5 present the effects of augmenting the diets of laying hens with *TG* powder on their blood biochemical profile. Among these variables, only the use of *TG* had a significant effect on the number of white blood cells ( $P < 0.05$ ). The highest number of white blood cells characterized those hens that received 4% of the powder, whereas those that received 2% had the lowest number of white blood cells.

**Table 4** Effects of using *Tragopogon graminifolius* plant powder on the blood lipid profile, total protein and uric acid concentrations of laying hens

Dietary level, %	Triglyceride, mg/dL	Cholesterol, mg/dL	Album, g/dL	Total protein, g/dL	Uric acid, g/dL	High density lipoprotein, mmol/L
0	1939.21	145.25	2.84	5.31	3.28	1.61
2	1524.92	124.24	3.05	4.85	2.09	6.08
4	1780.60	142.72	2.85	5.19	2.25	2.67
6	1292.40	78.63	3.18	4.90	2.25	2.72
SE	285.27	19.19	0.10	0.30	0.43	1.18
P-value	0.4425	0.1435	0.0931	0.6664	0.2574	0.1158

**Table 5** Effect of using *Tragopogon graminifolius* plant powder on blood cell numbers

Dietary level, %	Haematocrit, %	Haemoglobin, %	Red blood cells, $\times 10^6/\text{mm}^3$	White blood cells, $\times 10^6/\text{mm}^3$	Heterophil (H), %	Lymphocyte (L), %	H/L
0	33.00	11.50	2.80	25167 <sup>ab</sup>	6.67	92.67	0.07
2	32.67	10.64	2.73	22666 <sup>b</sup>	9.00	90.00	0.10
4	30.67	12.40	3.04	27000 <sup>a</sup>	14.00	86.00	0.17
6	33.00	12.10	2.75	25000 <sup>ab</sup>	10.67	89.00	0.12
SE	0.90	0.62	0.10	799	2.29	2.24	0.03
P-value	0.2723	0.2680	0.1753	0.0315	0.2214	0.2849	0.2491

<sup>a,b</sup> Within a column, means followed by a common superscript were not different with probability  $P = 0.05$

The use of *TG* powder increased feed intake, so the hens received more nutrients and had improved laying performance, probably because of the general positive effects of medicinal plants, such as promoting antimicrobial properties, stimulating the secretion of digestive enzymes, increasing dietary fibre, and enhancing the speed of gastrointestinal passage (Shirani *et al.*, 2019; Jin *et al.*, 2020; Youssef *et al.*, 2021). *Tragopogon graminifolius* makes feed more appetizing and this might increase feed intake (Leila *et al.*, 2011). However, in a study with mice, a methanolic extract of *TG* reduced feed intake (Zeeni *et al.*, 2014).

Inulin is an important active ingredient of *TG* merit and has prebiotic properties that can improve performance through functionalities. These include reducing the harmful microbial population of the

gastrointestinal tract (Liu et al., 2018; Xia et al., 2019), increasing the height of intestinal villi (Li et al., 2018; Ding et al., 2021), and increasing immunity (Song et al., 2020; Gong et al., 2020). Thus, it improves the condition of the gastrointestinal tract and increases digestion, absorption, and utilization of dietary nutrients, with a downstream effect of improving performance. The positive results of augmentation with *TG* in the present study were consistent with other studies that used inulin in poultry diets (Elrayeh & Yildiz, 2012; Park & Park, 2012; Abdelqader et al., 2013).

The positive effects of *TG* on the laying capacity of hens may arise from its antioxidant properties (Sasmakov et al., 2012). By protecting nutrients from oxidation, antioxidants facilitate their greater absorption from the gastrointestinal tract. As a result, fewer nutrients are excreted and the conditions for their utilization for improving egg production are provided.

Improved intestinal health might be another reason for the observed improvement of performance by the laying hens in this study. Farzaei et al. (2014) used *TG* as an effective treatment for intestinal inflammation in Wistar mice. Reducing intestinal inflammation could provide for greater nutrient absorption.

Medicinal plants such as *TG* are also rich in vitamins A and C and minerals such as iron, potassium, zinc, and copper, which can improve performance (Milani et al., 2011). Green plants such as *TG* are rich in pigments such as carotenoids (Marounek & Pebriansyah, 2018; Upadhyay, 2018). The use of green plants in the diet of laying hens could result in the transfer of their pigments to the egg yolk, making it more colourful (Nobakht, 2015a; Nobakht, 2015b).

Haugh units are an important indicator of quality of egg whites (Kraus et al., 2021; Malfatti et al., 2021). Egg white is rich in high-quality protein. Because an increase in the level of *TG* improved performance, the amount of absorbed protein possibly also increased, resulting in improved condition of egg whites and an increase in the Haugh unit.

The use of reduced shell thickness *TG* in comparison with the control, because of the increase in egg size. However, the weight of the eggshell did not increase as a percentage of egg weight, indicating that there was no disturbance in calcium uptake by chickens. The use of *TG* had no significant effects on blood biochemical parameters. The ratio of lymphocytes to heterophils remained low with the increasing levels of supplemental *TG*, indicating that this dietary augmentation did not materially increase the level of stress on the birds (Gross & Siegel, 1983). Another report indicated the use of inulin reduced blood cholesterol and triglyceride levels (Elrayeh & Yildiz, 2012).

The use of an aqueous extract of the aerial parts of the *TG* over four weeks improved the lipid profile significantly (triglyceride, total cholesterol, low density lipoprotein, and high density lipoprotein) of mice (Zeeni et al., 2014) unlike the results in the present study. However, numerous factors might have contributed to these differences, including the type of animal, test conditions, and production status. In the present study *TG* powder increased the white blood cell count. However, the differences were numerically small, and possibly the body was prepared to deal with harmful external factors (Signore et al., 2018; Manav et al., 2019). Flavonoids, and vitamins A and C are among the active ingredients in *TG* that can effectively increase immune indicators such as the number of white blood cells.

## Conclusion

The use of *TG* powder at up to 6% of the diets of laying hens improved performance and egg quality without meaningful adverse effects on haematology or serum lipids. Therefore, *TG* powder can be recommended as a feed additive for laying hens.

## Conflict of Interest Declaration

The authors declare that they have no conflict of interest.

## Authors' Contributions

AN supervised the experiment, VP and TA advised the project. AN conducted the laboratory analyses. TA and VP supervised the experiment and finalized the manuscript. IC edited the article.

## References

- Abdelqader, A., Al-Fataftah, A.R. & Daş, G., 2013. Effects of dietary *Bacillus subtilis* and inulin supplementation on performance, eggshell quality, intestinal morphology and microflora composition of laying hens in the late phase of production. *Animal Feed Science and Technology* 179(1-4), 103-111. <https://doi.org/10.1016/j.anifeedsci.2012.11.003>
- Bell, I.S., Aguilar, Y.M., Bertot, R.R., González, C.O., Ginarte, R. A., Torres, G.P. & Toro, D.M., 2017. Effects of supplementation with powder from various medicinal plants on productivity and quality of eggs from laying hens. *Revista de Producción Animal* 29(3), 1-5. <https://revistas.reduc.edu.cu/index.php/rpa/article/view/2224>
- Birmani, M.W., Nawab, A., Ghani, M., Li, G., Xiao, M. & An, L., 2019. A review: Role of inulin in animal nutrition. *Journal of Food Technology Research* 6, 18-27. <https://doi.org/10.18488/journal.58.2019.61.18.27>

Bucław, M., 2017. Inulin in poultry production. *World's Poultry Science Journal* 73(2), 301-308. <https://doi.org/10.1017/S0043933917000010>

Darabighane, B., Mirzaei Aghjeh Gheshlagh, F., Navidshad, B., Mahdavi, A., Zarei, A. & Nahashon, S., 2017. Effects of peppermint (*Mentha piperita*) and aloe vera (*Aloe barbadensis*) on ileum microflora population and growth performance of broiler chickens in comparison with antibiotic growth promoter. *Iranian Journal of Applied Animal Science* 7(1), 101-108. [http://ijas.iaurasht.ac.ir/article\\_528873.html](http://ijas.iaurasht.ac.ir/article_528873.html)

Ding, B., Chen, L., Lin, H., Wang, X., Zhang, L., Ni, X. & Fronte, B., 2021. Effects of inulin diet supplementation on production performance, gut traits, and incidence of ascites in Haidong chicks under hypoxic conditions. *Animal Bioscience* 34(3), 417. <https://doi.org/10.5713/ajas.20.0508>

Elrayeh, A.S. & Yildiz, G., 2012. Effects of inulin and beta-glucan supplementation in broiler diets on growth performance, serum cholesterol, intestinal length, and immune system. *Turkish Journal of Veterinary and Animal Sciences* 36(4), 388-394. <https://journals.tubitak.gov.tr/veterinary/abstract.htm?id=12827>

Falahi, E., Delshadian, Z., Ahmadvand, H. & Shokri Jokar, S., 2019. Head space volatile constituents and antioxidant properties of five traditional Iranian wild edible plants grown in west of Iran. *AIMS Agriculture and Food*. <http://eprints.lums.ac.ir/id/eprint/1881>

Farkhoy, M., Sigharody, F. & Niknafas, F. 1994. *Poultry breeding*. Coasar Publication. DOI

Farzaei, M.H., Ghasemi-Niri, S.F., Abdolghafari, A.H., Baeeri, M., Khanavi, M., Navaei-Nijeh M., Abdollahi M. & Rahimi R., 2014. Biochemical and histopathological evidence on the beneficial effects of *Tragopogon graminifolius* in TNBS-induced colitis. *Pharm. Biol.* 53(3), 429-436.

Gadde, U., Kim, W.H., Oh, S.T. & Lillehoj, H.S., 2017. Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry: A review. *Animal Health Research Reviews* 18(1), 26-45. <https://doi.org/10.1017/S1466252316000207>

Ghasem Shamsabadi, M., Zamani, A., Habibian Dehkordi, S. & Hassanpour, H., 2015. The effect of *Cinnamomum zeylanicum* Blume and *Thymus vulgaris* on lipid oxidation and nitric oxide in Japanese quail (*Coturnix japonica*). *Iranian Journal of Veterinary Clinical Sciences* 8(2), 41-50.

Gong, Q., Peng, Y.G., Niu, M.F. & Qin, C.L., 2020. Research Note: The immune enhancement ability of inulin on *ptfA* gene DNA vaccine of avian *Pasteurella multocida*. *Poultry Science* 99(6), 3015-3019. <https://doi.org/10.1016/j.psj.2020.03.006>

Gross, W.B. & Siegel, H.S., 1983. Evaluation of heterophile to lymphocyte ratio as measure of stress in chickens. *Avian Disease* 27, 972-979.

Jin, L.Z., Dersjant-Li, Y. & Giannenas, I., 2020. Chapter 10 - Application of aromatic plants and their extracts in diets of broiler chickens. *Feed Additives* 159-185. <https://doi.org/10.1016/B978-0-12-814700-9.00010-8>

Kraus, A., Zita, L., Krunt, O., Härtlová, H. & Chmelíková, E., 2021. Determination of selected biochemical parameters in blood serum and egg quality of Czech and Slovak native hens depending on the housing system and hen age. *Poultry Science* 100(2), 1142-1153. <https://doi.org/10.1016/j.psj.2020.10.039>

Leila, J., Ghassem, H & Hamide, S., 2011. Introduction the medicinal species of Asteraceae family in Ilkhji and Sharafaldin regions of East Azerbaijan in Iran. *Journal of American Science* 7(5), 45.

Li, B., Schroyen, M., Leblois, J., Wavreille, J., Soyeurt, H., Bindelle, J. & Everaert, N., 2018. Effects of inulin supplementation to piglets in the suckling period on growth performance, postileal microbial and immunological traits in the suckling period and three weeks after weaning. *Archives of Animal Nutrition*. 72(6), 425-442. DOI: 10.1080/1745039X.2018.1508975

Liu, H.Y., Hou, R., Yang, G.Q., Zhao, F. & Dong, W.G., 2018. In vitro effects of inulin and soya bean oligosaccharide on skatole production and the intestinal microbiota in broilers. *Journal of Animal Physiology and Animal Nutrition*. 102(3), 706-716. DOI: 10.1111/jpn.12830 \*\*\*\*

Malfatti, L.H., Zampar, A., Galvão, A.C., da Silva Robazza, W. & Boiago, M.M., 2021. Evaluating and predicting egg quality indicators through principal component analysis and artificial neural networks. *Lebensmittel-Wissenschaft und-Technologie* 148(9), 111720. DOI: 10.1016/j.lwt.2021.111720

Manav, M., Ceylan, M. & Büyükkapar, H.M., 2019. Investigation of reproductive efficiency, growth performance and survival of the southern medicinal leech *Hirudo verbana Carena* 1820 fed with mammalian and poultry blood. *Animal Reproduction Science* 206, 27-37. DOI: 10.1016/j.anireprosci.2019.05.004

Marounek, M. & Pebriansyah, A., 2018. Use of carotenoids in feed mixtures for poultry: A review. *Agricultura Tropica et Subtropica* 51(3), 107-111. DOI: 10.1515/ats-2018-0011

Milani, E., Golimovahhed, Q. A. & Hosseini, F., 2011. Application of response surface methodology for optimization of inulin extraction from salsify plant. *Food Technology Research Journal* 21(1), 35-43.

Nazifi, S. 1997. *Hematology and clinical biochemistry of birds*. Shiraz University Publication. [https://www.researchgate.net/publication/215778143\\_Haematology\\_and\\_serum\\_biochemistry\\_of\\_golden\\_eagle\\_Aquila\\_chrysaetos\\_in\\_Iran](https://www.researchgate.net/publication/215778143_Haematology_and_serum_biochemistry_of_golden_eagle_Aquila_chrysaetos_in_Iran)

Nobakht, A., 2015a. Effects of *Cichorium intybus* L. on performance, egg traits, blood biochemical and cellular parameters of laying hens. *Animal Research Journal* 4(3), 47-55. [https://ar.guilan.ac.ir/article\\_1413.html?lang=en](https://ar.guilan.ac.ir/article_1413.html?lang=en)

Nobakht, A., 2015b. Effects of different levels of *Silybum marianum* L. medicinal plant powder on performance, blood biochemical parameters and blood cells of laying hens. *Iranian Journal of Medicinal and Aromatic Plants*. 31(4), 698-708.

Oluwafemi, R.A., Olawale, I. & Alagbe, J.O., 2020. Recent trends in the utilization of medicinal plants as growth promoters in poultry nutrition-A review. *Research in: Agricultural and Veterinary Sciences* 4(1), 5-11.

Park, S.O. & Park, B.S., 2012. Effect of feeding inulin oligosaccharides on cecum bacteria, egg quality and egg production in laying hens. African Journal of Biotechnology 11(39), 9516-9521. DOI: 10.5897/AJB12.5250

Pliego, A.B., Tavakoli, M., Khusro, A., Seidavi, A., Elghandour, M.M., Salem, A.Z. & Rene Rivas-Caceres, R., 2020. Beneficial and adverse effects of medicinal plants as feed supplements in poultry nutrition: A review. Animal Biotechnology 33(2), 369-391. DOI: 10.1080/10495398.2020.1798973

Sasmakov, S.A., Putieva, Z.M., Azimova, S.S. & Lindequist, U., 2012. In vitro screening of the cytotoxic, antibacterial and antioxidant activities of some Uzbek plants used in folk medicine. Asian Journal of Traditional Medicines 7(2), 73-80.  
<https://www.semanticscholar.org/paper/In-vitro-screening-of-the-cytotoxic%2C-antibacterial-Sasmakov-Putieva/ed63286ec2a58280ed24990c9a4818247dd80247>

Shang, H., Zhang, H., Guo, Y., Wu, H. & Zhang, N., 2020a. Effects of inulin supplementation in laying hens diet on the antioxidant capacity of refrigerated stored eggs. International Journal of Biological Macromolecules 153, 1047-1057. DOI: 10.1016/j.ijbiomac.2019.10.234

Shang, H., Zhao, J., Dong, X., Guo, Y., Zhang, H., Cheng, J. & Zhou, H., 2020b. Inulin improves the egg production performance and affects the cecum microbiota of laying hens. International Journal of Biological Macromolecules. 155, 1599-1609. DOI: 10.1016/j.ijbiomac.2019.11.137

Shirani, V., Jazi, V., Toghyani, M., Ashayerizadeh, A., Sharifi, F. & Barekatain, R., 2019. *Pulicaria gnaphalodes* powder in broiler diets: Consequences for performance, gut health, antioxidant enzyme activity, and fatty acid profile. Poultry Science 98(6), 2577-2587. DOI: 10.3382/ps/pez010

Signore, A., Jamar, F., Israel, O., Buscombe, J., Martín-Comín, J. & Lazzeri, E., 2018. Clinical indications, image acquisition and data interpretation for white blood cells and anti-granulocyte monoclonal antibody scintigraphy: an EANM procedural guideline. European Journal of Nuclear Medicine and Molecular Imaging. 45(10), 1816-1831. DOI: 10.1007/s00259-018-4052-x

Song, J., Li, Q., Everaert, N., Liu, R., Zheng, M., Zhao, G. & Wen, J., 2020. Effects of inulin supplementation on intestinal barrier function and immunity in specific pathogen-free chickens with *Salmonella* infection. Journal of Animal Science 98(1). DOI: 10.1093/jas/skz396

Stolzenbach, S., Myhill, L.J., Andersen, L.O.B., Krych, L., Mejer, H., Williams, A.R., Nejsum, P., Stensvold, C.R., Nielsen, D.S. & Thamsborg, S.M., 2020. Dietary inulin and *Trichuris suis* infection promote beneficial bacteria throughout the porcine gut. Frontiers in Microbiology. 11, 312. DOI: 10.3389/fmicb.2020.00312

Torki, M., Mohebbifar, A. & Mohammadi, H., 2021. Effects of supplementing hen diet with *Lavandula angustifolia* and/or *Mentha spicata* essential oils on production performance, egg quality and blood variables of laying hens. Veterinary Medicine and Science 7(1), 184-193. DOI: 10.1002/vms3.343

Upadhyay, R.K., 2018. Plant pigments as dietary anticancer agents. International Journal of Green Pharmacy 12(01).

Xia, Y., Kong, J., Zhang, G., Zhang, X., Seviour, R. & Kong, Y., 2019. Effects of dietary inulin supplementation on the composition and dynamics of cecal microbiota and growth-related parameters in broiler chickens. Poultry Science 98(12), 6942-6953. DOI: 10.3382/ps/pez483

Youssef, I. M., Männer, K. & Zentek, J., 2021. Effect of essential oils or saponins alone or in combination on productive performance, intestinal morphology and digestive enzymes' activity of broiler chickens. Journal of Animal Physiology and Animal Nutrition 105(1), 99-107. DOI: 10.1111/jpn.13431

Zeeni, N., Daher, C.F., Saab, L. & Mroueh, M., 2014. *Tragopogon porrifolius* improves serum lipid profile and increases short-term satiety in rats. Appetite. 72: 1-7. DOI: 10.1016/j.appet.2013.09.023