

Concentrations of faecal glucocorticoid metabolites in South African Mutton Merino sheep fed various sources of roughage

H.A. O'Neill^{1#}, O.B. Einkamerer¹, B.T. Elago¹ & A. Ganswindt²

¹Department of Animal, Wildlife and Grassland Sciences, University of the Free State, Bloemfontein, South Africa

²Mammal Research Institute, Department of Zoology and Entomology, Faculty of Natural and Agricultural Sciences, University of Pretoria, Hatfield, South Africa

(Submitted 4 June 2019, Accepted 6 May 2020, Published online 20 August 2020)

Copyright resides with the authors in terms of the Creative Commons Attribution 4.0 South African Licence.

See: <http://creativecommons.org/licenses/by/4.0/za>

Condition of use: The user may copy, distribute, transmit and adapt the work, but must recognise the authors and the South African Journal of Animal Science.

Abstract

The objective of this study was to determine whether various sources of roughage in nutritionally balanced feedlot diets would evoke a stress response in sheep. A nutritional stress response test was performed on 20 ewes, randomly divided into four treatment groups with 5 ewes per group. Ewes were kept individually in metabolic crates for thirty days. Rations were balanced nutritionally, and various sources of roughage were included; T1: alfalfa hay, T2: maize stover, T3: soya hulls; and T4: *Eragrostis tef*. Faeces was removed manually from the caudal rectum of each ewe at 05h00 and 19h00. Samples were frozen at -20°C until analyses. A total of 520 faecal samples were analysed to determine the concentrations of faecal glucocorticoid metabolites (fGCM) by enzyme immunoassay (EIA). These concentrations were expressed as mass/g dry weight (ng/g DW). Baseline values were determined, and those greater than the mean plus 2 standard deviations were removed. Baseline values were compared between the groups with one-way ANOVA analysis. The average concentrations of fGCM were 178.77 ± 21.7 in the morning and 183.2 ± 14.4 in the evening. Sheep fed T4 had significantly higher fGCM concentrations in both morning (302.0 ± 86.1 ng/g DW) and evening (237.0 ± 48.1 ng/g DW) compared with the other treatment groups. Thus, fGCM concentrations in sheep were related to the source of roughage, and *Eragrostis tef* caused a stress response.

Keywords: animal welfare, cortisol, dietary fibre, selective grazer, stress response

#Corresponding author: oneillha@ufs.ac.za

Introduction

Systems of mutton production in South Africa include extensive and intensive practices. Pasture conditions in extensive conditions often expose sheep to harsh habitats, hazardous environmental and climatic conditions and hot temperatures (Abecia *et al.*, 2006; Corson *et al.*, 2007). As a result of these challenging environmental conditions, producers are turning towards intensive production, which is characterized by a closed system of feeding pens or feedlots. This practice requires large amounts of labour and capital input relative to land area to keep the animals healthy while production performance is enhanced (Morand-Fehr *et al.*, 2007). The stocking density is high, and farmers use cultivated pastures.

Roughages are included in a nutritionally balanced diet through proper feed formulation. The fibre quality of a roughage influences the digestibility and feed conversion ratio (FCR) of a diet. Alfalfa is a high-quality fibre source, which makes it an ideal nutritional feed source for sheep (Doran *et al.*, 2007). Maize stover is the by-product that remains after harvesting maize. It is often used as livestock feed during the dry season. Its quality may differ owing to variations among maize genotypes (Ertiro *et al.*, 2013). But its high fibre and low nitrogen content make maize stover less nutritious, unless animals are supplemented with nitrogen (Manyuchi, 1994). Soybean hulls are a by-product of soybean seeds after oil extraction. Soybean meal guarantees maximum protein of 44% to 48% and 0.5% fat (McEllhiney, 1995) after oil extraction. For this reason, soybean hulls are a good source of energy in diets that are more forage based. Teff (*Eragrostis tef* (Zuccagni trotter)) is known for its quality, high yield and ability to grow faster than other crops (Zeid *et al.*,

2011). It is used mostly as forage production in areas that are dry and have short rain seasons (Kaholongo, 2016). Today, teff is grown mainly for hay, particularly in South Africa, Australia and the USA (Miller, 2010).

Intensive production systems prevent sheep from grazing naturally. Sheep are naturally selective grazers and some breeds are more selective than others. For example, Dorper sheep are less selective than Merino-type breeds (Brand, 2000). Feedlot production restricts the normal selection behaviour of sheep, which are fed formulated diets. Restricting normal behaviour in sheep may evoke a stress response.

Stress may be defined as 'the inability of an animal to cope with its environment'. This phenomenon is revealed by failure to achieve genetic potential for growth rate, milk yield, disease resistance and fertility (Bayazit, 2009). Stress may be described in biological terms as a response that is elicited when an individual is under threat (Engelmann *et al.*, 2004). An individual will attempt to counter stress through behavioural responses or unconscious adaptations via physiological and biochemical mechanisms (Bashaw *et al.*, 2016). However, there are individual differences in behaviour and in the ways in which livestock species respond to stressful situations. It is important to understand not only herd differences, but also individual animal ones (O'Neill, 2019). Stress experienced by sheep cannot be avoided and forms part of normal management practices (Dwyer, 2009). The hypothalamic pituitary adrenal response is an evolutionary mechanism that enables an individual to return to homeostasis. Whereas short-term stressors and the physiological consequences may be desirable in some cases, chronic stress has a negative impact on production efficiency, that is, the suppression of normal oestrus behaviour and reduced ewe fertility (Marai *et al.*, 2007).

Researchers have attempted to measure stress and stress responses in animals for many years. Three main methods are used to measure stress in sheep: i) observing overall body condition, ii) analysing physiological responses such as cortisol concentration, and iii) observing general behavioural changes (Weary *et al.*, 2006; Gougoulis *et al.*, 2010; Kruger *et al.*, 2016). Sheep are known to be stoical creatures and do not always reveal discomfort or distress that is experienced physically or behaviourally. Close observation is important in identifying stress in sheep.

Glucocorticoids (GC) are a class of steroid hormones that bind to the glucocorticoid receptor, which is present in almost every vertebrate animal cell (Royo *et al.*, 2004). The GC are part of the feedback mechanism in the immune system that inhibits access of leukocytes to a site of inflammation (Möstl & Palme, 2002; Von der Ohe & Servheen, 2002). Glucocorticoids have diverse effects, including potentially harmful side effects (Pihl & Hau, 2003). Stress increases the GC concentrations in the blood of mammals, making blood an obvious matrix to evaluate stress responsiveness (Tuli *et al.*, 1995). Circadian cycles control adrenocortical responses after a stressful event (Przekop *et al.*, 1985; De *et al.*, 2017). The effect of the circadian rhythm on cortisol patterns should be considered carefully in stress assessments and use of a single sample of cortisol per day should be avoided (De Jong *et al.*, 2000). It is preferable to measure endogenous cortisol levels over 24 hours (Monfort *et al.*, 1993).

Unfortunately, capture and restraint of the animal for blood sampling causes it stress. Non-invasive methods to determine the concentration of GC or its metabolites are more desirable (Kruger *et al.*, 2016; Nemeth *et al.*, 2016; Palme, 2019). Samples that can be obtained using alternative non-invasive techniques include faecal, salivary, milk and hair (Siswanto *et al.*, 2008). However, there are many challenges in most sampling procedures. Saliva collection means handling an animal, and milk sampling is limited to lactating animals (Möstl & Palme, 2002). Faecal and salivary samples present an opportunity for non-invasive measurement of corticosteroids and immunoglobulin A (Pihl & Hau, 2003; Touma & Palme, 2005). Faecal glucocorticoid metabolite concentrations reflect glucocorticoid production in an animal's system after a certain time. The lag time for fGCM concentration in ruminants is 13 ± 4 hours. Faecal and salivary samples are less affected by short-term fluctuations of hormone secretion than blood samples (Touma & Palme, 2005).

Foraging involves interaction between the characteristics of the animals and those of the food in the environment (Prache *et al.*, 1998). The purpose of this study was to assess whether various sources of roughage that were fed to animals in an intensive controlled system would evoke stress responses in sheep.

Methods and Materials

This study was carried out in accordance with the guidelines of the Ethics Committee (Reference UFS-AED2017/0031) of the University of Free State, in autumn 2018 at Paradys Experimental Farm (29° 13' 19.78" S, 26° 12' 26.48" E), which is situated 20 km from Bloemfontein, Free State, South Africa. A total of 20 South African Mutton Merino (SAMM) ewes, seven months old, and weighing 28 kg on average were randomly allocated to one of four treatment groups. These groups consisted of T1 (n = 5): alfalfa hay, T2 (n = 5): maize stover, T3 (n = 5): soya hulls, and T4 (n = 5): *Eragrostis tef*. Rations were balanced nutritionally, and various fibre sources were used (Table 1).

Sheep were kept individually in metabolic crates for thirty days after an adaptation period of eight days. Diets were fed ad libitum for the experimental period. At the onset and end of the study, all animals

were fasted overnight (minimum of 12 hours) and individual empty stomach bodyweight was recorded the next morning. Animals were provided with feed twice daily at 07h00 and at 17h00 during the study. Feed troughs were designed/ to determine individual quantities of ingested feed and to minimize feed wastage.

Table 1 Physical and chemical composition of four diets including different sources of fibre

Raw materials (% inclusion)	Dietary treatment			
	T1	T2	T3	T4
Maize meal	55.00	55.00	55.00	55.00
Lucerne	34.97	0.00	0.00	0.00
Maize stover	0.00	19.53	0.00	0.00
Eragrostis	0.00	0.00	0.00	21.63
Citrus pulp	6.89	20.05	17.70	17.94
Soya oil cake	0.03	0.29	0.00	0.24
Soya hulls	0.00	0.00	22.85	0.00
Urea	0.66	2.07	1.43	1.91
Limestone	0.25	0.72	0.67	0.85
Ammonium chloride	0.75	0.75	0.75	0.75
Mono-calcium phosphate	0.70	0.85	0.85	0.92
Salt	0.50	0.50	0.50	0.50
Premix	0.25	0.25	0.25	0.25
<i>Nutrient composition (% DM)</i>				
Crude protein	14.50	14.50	14.50	14.50
Metabolizable energy (MJ ME/kg DM)	11.28	11.60	12.19	11.25
Neutral detergent fibre	22.83	22.83	22.83	22.83
Acid detergent fibre	15.48	13.85	15.35	12.06
Ether extract	3.35	3.35	3.35	3.35
Calcium	0.83	0.83	0.83	0.83
Phosphorus	0.40	0.40	0.40	0.40

T1: alfalfa hay, T2: maize stover, T3: soya hulls, T4: *Eragrostis tef*

Approximately 20 g faeces were removed manually twice a day. A total of 520 faecal samples were collected for 14 days and then again at day 30 of the trial. Samples were collected at the same times each day at 05h00 - 06:00 and 19h00 - 20h00. Each faecal sample was placed in a sterile glass container with a screw lid. Samples were put on ice in a cooler bag and frozen within 30 minutes of collection. Samples were kept in a freezer at -20 °C until analyses.

Laboratory analyses were conducted at the Endocrine Research Laboratory, University of Pretoria (Ganswindt *et al.*, 2002). Faecal glucocorticoid metabolite concentrations were measured with an enzyme immunoassay (EIA). 11-oxoetiocholanolone was used as a group-specific enzyme detecting immunoassay by measuring 11,17-dioxoandrostane (see Palme & Möstl, 1997).

Frozen samples were lyophilized, pulverized and then sieved through a wire-mesh strainer to remove undigested material. Enzyme immune assays were performed on microtiter plates according to Ganswindt *et al.* (2002). Fifty µl aliquots of standards (range 1.9 - 500 pg for EA), quality controls and diluted faecal extract were pipetted in duplicate into the microtiter plate wells. Then 50 µl of EA label was added and the plates were incubated overnight at 4 °C. After incubation, the plates were washed four times and 150 µl (20 ng) streptavidin-peroxidase was added to each well. After incubation in the dark for 30 minutes while shaking,

plates were washed again before 150 µl peroxidase substrate solution was added. Plates were then incubated for 30 - 60 min while shaking. The reaction was terminated by adding 50 µl of 4 normal sulphuric acid and optical density was measured at 450 nm. To adjust for variations in water content, faecal hormone concentrations were expressed as mass/g DW. The sensitivity of the EIA was 1.2 ng/g faecal DW. The intra-assay coefficient of variance, which was determined by repeated measurement of high- and low-value quality controls, ranged from 3.83% to 4.09% and the inter-assay coefficient of variance ranged from 7.72% to 9.59%.

Baseline values of fGCM concentrations were determined for each group using an iterative process in which all values greater than the mean plus 2 standard deviations (SD) were removed (Brown *et al.*, 1999). Averages were recalculated and the elimination process was repeated until there were no values greater than the mean plus 2 SD.

For all groups, fGCM data were pooled and differences in hormone concentration levels were examined with one-way ANOVA, followed by Tukey test post-hoc analysis. The baseline values from the fGCM concentration levels of individual sheep were then compared among all groups. The initial rise, peak and fall in fGCM concentration after the introduction of the treatment feed were determined and analysed using descriptive statistics. Means and variances of all groups were compared using ANOVA analysis of R statistical package and Kruskal-Wallis ANOVA on ranks. The package was also used to detect significant differences between the groups according to treatments. For all statistical analyses, significance was assumed when $P < 0.05$.

Results and Discussion

This study formed part of a production trial. Average daily weight gain (ADG) and average daily feed intake (ADI) were recorded and used to calculate FCR. There were no significant differences in FCR between groups. The experimental diets were comparable and ADI and ADG were not significantly different among groups in this study.

Table 2 Average (\pm SE), maximum and minimum faecal glucocorticoid metabolites (ng/g DW) in the morning and evening for South African Mutton Merino ewes fed various sources of roughage

Measurement	T1	T2	T3	T4	Average
AM Average	113.10 \pm 25.91 ^a	153.69 \pm 42.30 ^a	146.28 \pm 35.46 ^a	301.99 \pm 86.05 ^b	178.77 \pm 21.69
Minimum	21.43	10.85	16.04	18.98	16.83 \pm 1.17
Maximum	223.07	330.28	296.81	796.62	411.70 \pm 67.26
PM Average	165.73 \pm 41.86 ^a	169.45 \pm 45.86 ^a	162.30 \pm 34.96 ^a	236.98 \pm 48.10 ^b	183.2 \pm 14.38
Minimum	15.09	14.76	22.89	48.46	25.30 \pm 4.10
Maximum	351.47	385.89	311.95	442.94	373.07 \pm 14.34

T1: alfalfa hay, T2: maize stover, T3: soya hulls, T4: *Eragrostis tef*

^{a,b} Within a row values with a common superscript do not differ at $P = 0.05$

Average morning fGCM concentration ranged from 113.10 \pm 25.91 ng/g DW (T1) to 301.99 \pm 86.05 ng/g DW (T4). Average evening fGCM concentration ranged from 162.30 \pm 34.96 ng/g DW (T3) to 236.98 \pm 48.10 ng/g DW (T4). Pooled average morning fGCM concentration was 178.77 \pm 21.69 ng/g DW and evening fGCM concentration was 183.2 \pm 14.38 ng/g DW. Overall, there were no significant differences in fGCM concentrations of samples collected in the mornings compared with those collected in the evening (Table 2). This contrasts with the patterns observed in cattle (Touma & Palme, 2005) and buffaloes (Ganswindt *et al.*, 2012). Ganswindt *et al.* (2012) demonstrated that glucocorticoids have circadian rhythm and diurnal differences, which classifies ruminants as diurnal animals with activity peaking during the day. The lag time for fGCM was about 13 hours. Thus the time of sampling is a reflection of the situation about 13 hours earlier (Touma & Palme, 2005). T4 was an exception, with higher fGCM levels on average in the mornings (301.99 \pm 86.05 ng/g) compared with the evenings (236.98 \pm 48.10 ng/g), but the reason is not clear. High variations in data indicate a possible sampling error and may be a result of small sampling groups.

There were no significant differences for fGCM between T1, T2, and T3 for mornings and evenings. Faecal glucocorticoid metabolite concentration for T4 was significantly higher ($P < 0.05$) in the morning and the evening compared with T1, T2, and T3. Preventing animals from grazing selectively and feeding a

roughage source that consisted mainly of teff evoked a stress response in sheep. In comparison, the voluntary intake of a teff only diet was lower. It was suggested that an optimum level of condensed tannin sources must be fed to sheep to improve feed intake and growth performance (Kaitho *et al.*, 1997). Santhi *et al.* (2017) suggested that parasitized animals have the ability to self-medicate because ingested plant phenolics, mainly tannins, reduce strongyle nematode infections in mammalian herbivores.

Conclusion

This study confirmed that fGCM measurement offered a practical alternative for investigators to monitor adrenocortical activity in sheep non-invasively. *Eragrostis tef* induced physiological stress in sheep that were not able to graze naturally. Other roughage sources must be fed to feedlot animals in conjunction with teff to ensure voluntary food intake and production efficiency. This study opened an avenue for further research on fibre preferences and animal health. In future, more research must be done on the natural selection of roughage sources and its provision in a feedlot diet because teff evoked a stress response. The ability to measure fGCM in SAMM sheep reliably with a non-invasive method would give a firm basis for further examination of physiological responses to stressful circumstances in commercially reared sheep.

Acknowledgements

The support and financial aid from Dr O.B. Einkamerer is appreciated.

Authors' Contributions

Conception, design and drafting of paper AON; data collection and analyses BTE; and critical review OBE

Conflict of Interest Declaration

There are no conflicts of interest.

References

- Abecia, J.A., Sosa, C., Forcada, F. & Meikle, A., 2006. The effect of undernutrition on the establishment of pregnancy in the ewe. *Reprod. Nutr. Dev.* 46(4), 367-378. DOI: 10.1051/rnd:2006018
- Bashaw, M.J., Sicks, F., Palme, R., Schwarzenberger, F., Tordiffe, A.S. & Ganswindt, A., 2016. Non-invasive assessment of adrenocortical activity as a measure of stress in giraffe (*Giraffa camelopardalis*). *BMC Vet. Res.* 12(1), 235. DOI org/10.1186/s12917-016-0864-8
- Bayazit, V., 2009. Evaluation of cortisol and stress in captive animals. *Aust. J. Basic. Appl. Sci.* 3, 1022-31.
- Brand, T.S., 2000. Grazing behaviour and diet selection by Dorper sheep. *Small Ruminant Res.* 36(2), 147-158. DOI org/10.1016/S0921-4488(99)00158-3
- Corson, M.S., Rotz, C.A. & Skinner, R.H., 2007. Evaluating warm-season grass production in temperate-region pastures: A simulation approach. *Agric. Syst.* 93(1-3), 252-268. DOI org/10.1016/j.agsy.2006.06.001
- De Jong, I.C., Prelle, I.T., Van de Burgwal, J.A., Lambooi, E., Korte, S.M., Blokhuis, H.J. & Koolhaas, J.M., 2000. Effects of environmental enrichment on behavioral responses to novelty, learning, and memory, and the circadian rhythm in cortisol in growing pigs. *Physiol. Behav.* 68(4), 571-578. DOI org/10.1016/S0031-9384(99)00212-7
- De, K., Kumar, D., Saxena, V.K. & Naqvi, S.M.K., 2017. Study of circadian rhythmicity of physiological response and skin temperature of sheep during summer and winter in semi-arid tropical environment. *Physiol. Behav.* 169, 16-21. DOI org/10.1016/j.physbeh.2016.11.010
- Doran, M.P., Laca, E.A. & Sainz, R.D., 2007. Total tract and rumen digestibility of mulberry foliage (*Morus alba*), alfalfa hay and oat hay in sheep. *Anim. Feed. Sci. Tech.* 138(3-4), 239-253. <https://doi.org/10.1016/j.anifeedsci.2006.11.016>
- Dwyer, C.M., 2009. Welfare of sheep: Providing for welfare in an extensive environment. *Small, Rumin. Res.* 86(1-3), 14-21. DOI org/10.1016/j.smallrumres.2009.09.010
- Engelmann, M., Landgraf, R. & Wotjak, C.T., 2004. The hypothalamic-neurohypophysial system regulates the hypothalamic-pituitary-adrenal axis under stress: An old concept revisited. *Front. Neuroendocrinol.* 25(3-4), 132-149. DOI org/10.1016/j.yfrne.2004.09.001
- Ertiro, B.T., Twumasi-Afryie, S., Blümmel, M., Friesen, D., Negera, D., Worku, M., Abakemal, D. & Kitenge, K., 2013. Genetic variability of maize stover quality and the potential for genetic improvement of fodder value. *Field Crops Res.* 153, 79-85. DOI org/10.1016/j.fcr.2012.12.019
- Ganswindt, A., Heistermann, M., Borrigan, S. & Hodges, J.K., 2002. Assessment of testicular endocrine function in captive African elephants by measurement of urinary and faecal androgens. *Zoo Biology* 21(1), 27-36. DOI org/10.1002/zoo.10034
- Ganswindt, A., Brown, J.L., Freeman, E.W., Kouba, A.J., Penfold, L.M., Santymire, R.M., Vick, M.M., Wielebnowski, N., Willis, E.L. & Milnes, M.R., 2012. International Society for Wildlife Endocrinology. *Biol. Lett.* 8, 695-697. DOI org/10.1098/rsbl.2011.1181
- Gougoulis, D.A., Kyriazakis, I. & Fthenakis, G.C., 2010. Diagnostic significance of behaviour changes of sheep: A selected review. *Small Ruminant Res.* 92(1-3), 52-56. DOI org/10.1016/j.smallrumres.2010.04.018
- Kaholongo, L.T., 2016. Screening of indigenous forage legumes as potential fodder crops and protein source for livestock in central Namibia. Doctoral dissertation, University of Namibia.

- Kaitho, R.J., Umunna, N.N., Nsahlai, I.V., Tamminga, S. & Van Bruchem, J., 1997. Utilization of browse supplements with varying tannin levels by Ethiopian Menz sheep: 1. Intake, digestibility and live weight changes. *Agrofor. Syst.* 39(2), 45-159.
- Kruger, L.P., Nedambale, T.L., Scholtz, M.M. & Webb, E.C., 2016. The effect of environmental factors and husbandry practices on stress in goats. *Small Ruminant Res.* 141, 1-4. DOI org/10.1016/j.smallrumres.2016.06.004
- Manyuchi, B., 1994. High quality forages as supplements to low quality forages for ruminants. Effects on intake, digestibility and rumen digesta dynamics. Doctoral dissertation, University of Aberdeen.
- Marai, I.F.M., El-Darawany, A.A., Fadiel, A. & Abdel-Hafez, M.A.M., 2007. Physiological traits as affected by heat stress in sheep — a review. *Small Ruminant Res.* 71(1-3), 1-12. DOI org/10.1016/j.smallrumres.2006.10.003
- McElhiney, R., 1995. Quality control in feed manufacturing. American Soybean Association, Brussels. Pp.1-20.
- Miller, D.R., 2010. Teff grass crop overview and forage production guide (second edition). Cal/West Seed Company. Woodland, CA.
- Monfort, S.L., Brown, J.L. & Wildt, D.E., 1993. Episodic and seasonal rhythms of cortisol secretion in male Elds deer (*Cervus eldi thamin*). *J. Endocrinol.* 138(1), 41-49. DOI org/10.1677/joe.0.1380041
- Morand-Fehr, P., Fedele, V., Decandia, M. & Le Frileux, Y., 2007. Influence of farming and feeding systems on composition and quality of goat and sheep milk. *Small Rumin. Res.* 68(1-2), 20-34. DOI org/10.1016/j.smallrumres.2006.09.019
- Möstl, E. & Palme, R., 2002. Hormones as indicators of stress. *Domest. Anim. Endocrinol.* 23(1-2), 67-74. DOI org/10.1016/S0739-7240(02)00146-7
- Nemeth, M., Pschernig, E., Wallner, B. & Millesi, E., 2016. Non-invasive cortisol measurements as indicators of physiological stress responses in guinea pigs. *PeerJ.* 4, e1590. DOI org/10.7717/peerj.1590
- O'Neill, H.A., 2019. A review on the involvement of catecholamines in animal behaviour. *S. Afr. J. Anim. Sci.* 49(1), 1-8. <http://dx.doi.org/10.4314/sajas.v49i1.1>
- Palme, R. & Möstl, E. 1997. Measurement of cortisol metabolites in faeces of sheep as a parameter of cortisol concentration in blood. *Mamm. Biol.* 62 (Suppl. II), 192-197.
- Palme, R., 2019. Non-invasive measurement of glucocorticoids: Advances and problems. *Physiol. Behav.* 199 229-243. DOI org/10.1016/j.physbeh.2018.11.021
- Pihl, L. & Hau, J., 2003. Faecal corticosterone and immunoglobulin A in young adult rats. *Lab. Anim.* 37(2), 166-171. DOI org/10.1258/00236770360563822
- Prache, S., Gordon, I.J. & Rook, A.J., 1998. Foraging behaviour and diet selection in domestic herbivores. *Ann. Zootech.* 47, 335-345.
- Przekop, F., Stupnicka, E., Wolińska-Witort, E., Mateusiak, K., Sadowski, B. & Domański, E., 1985. Changes in circadian rhythm and suppression of the plasma cortisol level after prolonged stress in sheep. *Eur. J. Endocrinol.* 110(4), 540-545.
- Royo, F., Bjork, N., Carlsson, H.E., Mayo, S. & Hau, J., 2004. Impact of chronic catheterization and automated blood sampling (Accusampler) on serum corticosterone and faecal immunoreactive corticosterone metabolites and immunoglobulin A in male rats. *J. Endocrinol.* 180(1), 145-154.
- Santhi, V.S., Salame, L., Dvash, L., Muklada, H., Azaizeh, H., Mreny, R., Awwad, S., Markovics, A., Landau, S.Y. & Glazer, I., 2017. Ethanolic extracts of *Inula viscosa*, *Salix alba* and *Quercus calliprinos* negatively affect the development of the entomopathogenic nematode *Heterorhabditis bacteriophora* – A model to compare gastrointestinal nematodes developmental effect. *J. Invertebr. Pathol.* 145, 39-44. DOI org/10.1016/j.jip.2017.03.005
- Siswanto, H., Hau, J., Carlsson, H.E., Goldkuhl, R. & Abelson, K.S., 2008. Corticosterone concentrations in blood and excretion in faeces after ACTH administration in male Sprague-Dawley rats. *In Vivo* 22(4), 435-440.
- Touma, C. & Palme, R., 2005. Measuring faecal glucocorticoid metabolites in mammals and birds: The importance of validation. *Ann. N. Y. Acad. Sci.* 1046(1), 54-74. DOI 10.1196/annals.1343.006
- Tuli, J.S., Smith, J.A. & Morton, D.B., 1995. Stress measurements in mice after transportation. *Lab. Anim.* 29(2), 132-138. DOI org/10.1258/002367795780740249
- Von der Ohe, C.G. & Servheen, C., 2002. Measuring stress in mammals using faecal glucocorticoids: opportunities and challenges. *Wildl. Soc. Bull.* 1215-1225.
- Weary, D.M., Niel, L., Flower, F.C. & Fraser, D., 2006. Identifying and preventing pain in animals. *Appl. Anim. Behav. Sci.* 100(1-2), 64-76. DOI org/10.1016/j.applanim.2006.04.013
- Zeid, M., Echenique, V., Díaz, M., Pessino, S. & Sorrells, M.E., 2011. Eragrostis. In: *Wild crop relatives: Genomic and breeding resources*. Springer, Berlin, Germany.