

## Genetic diversity in Zimbabwean Sanga cattle breeds using microsatellite markers

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### Abstract

Zimbabwe's smallholder land-based livelihoods are dominated by the three local Sanga cattle breeds, namely Mashona, Tuli, and Nkone. A study was carried out to determine genetic diversity and differentiation among conservation populations of these breeds using 16 bovine-specific microsatellite markers. These markers included BM1818, BM1824, BM2113, CSRM60, CSSM66, ETH10, ETH225, ETH3, ILST006, INRA23, RM067, SPS115, TGLA122, TGLA126, TGLA227, and TGLA53. All marker loci contributed to breed differentiation based on the infinitesimal model ( $F_{ST}$ ), with the most powerful markers being CSSM66 (25%), ETH225 (20.6%), and TGLA122 (13.8%) and the least powerful being RM067 (0.7%), BM1824 (1.0%), and ETH10 (1.1%). Three marker loci (BM1824, ETH225, and ETH3) revealed significant deviations from Hardy-Weinberg equilibrium (HWE) proportions. A total of 119 alleles were observed, ranging from 4 to 11 and averaging 7.4 alleles per locus. Thirty-four of these alleles were unique to specific breeds. Mean ( $N_a$ ) and effective ( $N_e$ ) numbers of alleles were  $5.167 \pm 0.302$  and  $3.462 \pm 0.163$  alleles per locus, respectively, with no significant differences between breeds. Observed heterozygosity ( $H_o$ ) (0.73) was higher than expected heterozygosity ( $H_e$ ) (0.71), revealing that breeds were in HWE. Global F-statistics  $F_{IT}$ ,  $F_{ST}$ , and  $F_{IS}$  gave mean values of 0.059, 0.084 and -0.028, respectively. Overall breed genetic differentiation was moderate ( $F_{ST} = 8.4\%$ ) and significant ( $P < 0.001$ ). Multivariate analyses separated the three breeds completely, and revealed that most of the genetic variation was within breed (92%). The analyses revealed that a significant amount of variation is maintained in these breeds and that they are distinct genetic entities that may be considered for utilization or conservation.

**Keywords:** Breed relationships, cattle conservation, genetic variation, molecular markers, Zimbabwe

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### Introduction

Cattle are critical to the overall agricultural production and livelihood system of most smallholder farming households in southern African countries, including Zimbabwe (Assan, 2012; Nyamushamba *et al.*, 2017). Most farms are diversified, keeping cattle resources that are multi-functional in nature to provide a wide variety of goods and services that generate income, ensure food security and support rural livelihoods (Rege *et al.*, 2011). Most common livestock benefits include food supply in the form of meat and milk, income from sale of live animals and their products, manure for soil fertility amelioration, and transport and draft power services. Other benefits include a supply of hides and skins for leather tanning, dung as a source of fuel, provision of savings and insurance services (especially where these services are non-existent, inaccessible or unreliable), diversification of rural livelihood options, and meeting the socio-cultural roles and obligations of their owners (Rege *et al.*, 2011; Nyamushamba *et al.*, 2017). Cattle act as an insurance bank for use in lean cropping seasons, and make households less vulnerable to crop failure because of pest, disease and drought occurrences. Cattle ownership, therefore, forms a social safety net and an important component of the resilience of land-based livelihoods for millions of people living in marginal production areas (Murungweni *et al.*, 2014; Van Vliet *et al.*, 2015).

Humped cattle breeds of southern Africa are classified into zebu (thoracic hump) and Sanga (cervico-thoracic hump) types. Sanga-type cattle constitute the bulk of the cattle genetic resources in this region and

include breeds such as the Nguni, Drakensburger and Afrikaner of South Africa, Landim of Mozambique, Sanga of Namibia, Nguni of Swaziland and Zambia, and Mashona, Nkone and Tuli of Zimbabwe (Ramsay, 2010; Pienaar *et al.*, 2015; Nyamushamba *et al.*, 2017). It is presumed that the Sanga group of cattle resulted from the crossbreeding of African taurine and zebu cattle around 700 AD in Ethiopia, followed by migration to southern Africa with tribesmen via the tsetse-free Mozambique corridor (Felius *et al.*, 2014). Natural adaptation to agricultural habitats in diverse environments and systematic breed-oriented selection led to genetic and phenotypic differentiation into various ecotypes and breeds. Adaptation in Sanga cattle is perceptible in the form of heat tolerance, resistance to local disease epidemics, ticks and tick-borne diseases, and improved and diversified production ability (Ramsay, 2010; Nyamushamba *et al.*, 2017). This makes these genotypes not only acclimatised to their habitat, but also fit for the purpose for which they are kept (Wilson, 2009). In addition, there are anecdotal reports that these genotypes harbour great genetic diversity and adaptive traits that make them potentially valuable resources for breeding programmes in regions facing similar biological stresses, and in response to the present and future needs of livestock production (Van Arendonk, 2011; Acosta *et al.*, 2013; FAO, 2015). Local Sanga genotypes can be used in various pure and cross breeding systems, helping farms to cope with projected management, market and climate scenarios in the future (Wilson, 2009; Edea *et al.*, 2013).

Zimbabwean Sanga cattle breeds have previously been characterized with morphological descriptors (Rowlands *et al.*, 2003), and protein-based biochemical markers (Carr *et al.*, 1966), and based on productive and reproductive performance traits (Dlodlo & Ward, 1987; Holness, 1992; Moyo, 1997; d'Hotman & Hatendi, 1998). Morphometric characterization is useful for differentiating animals based on observable phenotype, but is subjective, has low polymorphism, and provides no basis for differentiating animals that look similar (Yang *et al.*, 2013). In addition, morphological markers are limited in the evaluation of quantitative traits. In the various productive performance experiments, Mashona and Tuli cattle were observed to be more resistant to ticks and heat stress, and had high calving rates, weaning rates, and cow productivity under marginal production environments compared with some exotic breeds (Moyo, 1997; Khombe, 2002). While morphological markers and productive traits indicate the functional genetic diversity that a population carries (Ajmone-Marsan, 2010), they do not account for systematic environmental effects on these phenotypes (Yang *et al.*, 2013). Biochemical markers have higher polymorphisms than morphological markers and are useful for elucidating the origin and classification of breeds. However, the degenerate nature of the genetic code makes them vulnerable to environmental impact and individual growth discrepancies. As result, a great amount of genetic variations at biochemical loci remain undetected (Singh *et al.*, 2014). As such, the genetic diversity and distinctiveness among these breeds remains largely unknown (Nyamushamba *et al.*, 2017). Lack of genetic characterization represents a glaring gap in cattle genetic research in Zimbabwe.

Phenotypic and biochemical characterization needs to be verified by complementary molecular characterization (Acosta *et al.*, 2013; FAO, 2015). In recent times, various types of DNA-based (molecular) markers have been utilized to evaluate genetic variability within and between African cattle breeds (Kugonza *et al.*, 2011; Ngono-Ema *et al.*, 2014; Ndiaye *et al.*, 2015). Among them, microsatellite and single nucleotide polymorphism (SNP) markers are the most efficient molecular markers to evaluate genetic diversity, population differentiation and breed relationships, and determine parentage in animal populations (Yang *et al.*, 2013). Advantages of microsatellites include wide distribution in the genome, low quantities of template DNA required (10–100 ng), multi-allelic attributes, a neutral co-dominant inheritance pattern that allow ease of distinction between homozygous and heterozygous alleles, the option of simultaneous sequencing using multiplex polymerase chain reaction (PCR), and a wide range of immediate applications (Yang *et al.*, 2013; Singh *et al.*, 2014). In southern Africa, microsatellite markers have been used in population genetic studies of some Sanga cattle genotypes such as the Mozambican Landim (Bessa *et al.*, 2009) and the South African Nguni (Sanarana *et al.*, 2016). The genetic diversity of cattle has also recently been studied using SNP markers in Ethiopia (Edea *et al.*, 2013) and South Africa (Makina *et al.*, 2016).

Molecular markers could be powerful tools in the identification of important alleles and their frequencies, rare alleles and unique genetic traits, and in revealing current status of genetic diversity within and differentiation between livestock populations. This is essential for future monitoring of gene flow, parental definitions, genetic traceability, and working evidence-based decisions on conservation farms in Zimbabwe. It could also strengthen the present and future utility of these breeds in varied cattle production systems (Nyamushamba *et al.*, 2017). The present study was therefore, aimed at determining both within and between breed genetic diversity and differentiation among the three most important Zimbabwean Sanga cattle conservation populations, namely Mashona, Tuli, and Nkone, using microsatellite markers.

## Materials and methods

Conservation herds of improved indigenous cattle (Mashona, Tuli, and Nkone) at Henderson, Matopos, Makoholi, and Grasslands Research Institutes in Zimbabwe were used as the study populations.

These four public research institutes are managed by the Department of Research & Specialist Services (DR&SS), Zimbabwe. Among the sampling herds, only breeding animals with traceable records and known breed were purposively selected for the study. The highly purposive sampling strategy had an effect on sample sizes, with fewer than the recommended 20–30 animals per breed (FAO, 2011; Lenstra *et al.*, 2012) sampled in the Nkone (Table 1). Two of the sampling sites (Henderson and Grasslands) are located in the high rainfall Highveld areas of Zimbabwe (Agro Ecological Region II) while Makoholi and Matopos Research Institutes are located in lower-rainfall sweetveld areas of the country (Agro Ecological Region IV). Samples were collected from healthy animals by a qualified veterinarian according to the 'Guidelines: Animal handling and sample collection' of the Animal Production Laboratory, Chinhoyi University of Technology. Blood was collected by jugular venipuncture using 4 ml vacutainer tubes containing 0.5 M EDTA as an anticoagulant, mixed homogeneously to avoid clotting, transported on ice to the laboratory within five hours and stored in a freezer at -20 °C until required for analyses.

**Table 1** Site description and number of animals sampled for the molecular diversity study of Zimbabwean Sanga cattle

Site	Site description			Number sampled			
	Eco-zone	Rainfall	Grassland type	Mashona	Nkone	Tuli	Total
Grasslands	IIa	High	Sour	8	-	6	14
Henderson	IIb	High	Sour	6	-	-	6
Makoholi	IV	Low	Sweet	7	-	-	7
Matopos	IV	Low	Sweet	-	7	16	23
Total				21	7	22	50

A total of 16 bovine-specific microsatellite marker pairs (Table 2), as recommended by the International Society for Animal Genetics (ISAG) and the Food and Agriculture Organization of the United Nations (FAO, 2011) for cattle genetic studies, were used to characterize all study animals. Primers were synthesized and supplied by Inqaba Biotechnical Industries Ltd (Pretoria, South Africa) and shipped as lyophilised oligonucleotide pellets. Upon arrival at the Animal Production Laboratory of Chinhoyi University of Technology, the pellet tubes were centrifuged at 3 000 x g for 30 seconds in a Universal Centrifuge PLC-036 (Gemmy Industrial Corporation, Taiwan), diluted to 100 µM stock solution in sterile water and stored at -20 °C. To avoid frequent freeze-thaw cycles during work, aliquots of the primers (oligonucleotides) were prepared from the stock solution. A working solution was prepared from each aliquot to a 10 µM concentration by diluting 10 µl of the 100 µM primer with 90 µl nuclease-free sterile water. Presence of the primer was confirmed by 1.5% agarose gel electrophoresis using 1X TAE buffer on an OmniPAGE system (Bio.com Direct, UK) and visualization on an Infinity ST5 gel documentation system (Vilber Lourmat, France).

The DNA was extracted from peripheral blood lymphocytes using the ZR Genomic DNA TM-Tissue MiniPrep (D3050) commercial kit (Zymo Research Europe GmbH, Freiburg im Breisgau, Germany). The eluted DNA was kept in a freezer at -20 °C until required for polymerase chain reaction (PCR) amplification. DNA from nine purebred animals of the three study breeds and one Brahman that was registered in the Zimbabwe Herd Book were used as standard samples for screening the initial 26 primers for polymorphism in the study population. This work was done at the Animal Production Laboratory of Chinhoyi University of Technology, Zimbabwe. All PCR reagents were supplied by Thermo Fisher Scientific. Each reaction was in a final volume of 25 µl, containing 12.5 µl DreamTaq PCR Master Mix 2X, 10 µM forward primer (1 µl), 10 µM reverse primer (1 µl), 20 ng template DNA (1 µl), and nuclease free water (9.5 µl). The proprietary DreamTaq Master Mix was composed of DreamTaq DNA polymerase, 2x DreamTaq buffer, dATP, dCTP, dGTP, and dTTP, 0.4 mM each, and 4 mM MgCl<sub>2</sub>. Control reactions with no-DNA template were also prepared to check for contamination and primer dimers after PCR. The PCR was performed on a 48-well plate dual block Arktik Thermal Cycler V01.07 (Thermo Fisher Scientific). After an initial denaturing step of 60 seconds at 98 °C, PCR was performed for 30 cycles, each consisting of a denaturation step of 94 °C for 20 seconds, an annealing step of 54–62 °C for 75 seconds and an extension step of 72 °C for 30 seconds. A final extension step at 72 °C for 10 minutes was used before the temperature was brought down to 4 °C. Amplification was

confirmed by visualization of bands in a gel documentation system after electrophoresis using 1.5% agarose gel and 0.5X TBE buffer on an OmniPAGE system (Biocom Direct, UK).

**Table 2** Characteristics of 16 dinucleotide microsatellite loci used for genetic characterisation of Zimbabwean Sanga cattle

Locus	Primer Sequences (5'-3') F, R	Chr	Ta	Size (bp)	Reference
BM1818	F:5'-AGC TGG GAA TAT AAC CAA AGG-3' R:5'-AGT GCT TTC AAG GTC CAT GC-3'	23	55 °C	248–276	Bishop <i>et al.</i> , 1994
BM1824	F:5'-GAG CAA GGT GTT TTT CCA ATC-3' R:5'-CAT TCT CCA ACT GCT TCC TTG-3'	1	58 °C	170–218	Barendse <i>et al.</i> , 1994
BM2113	F:5'-GCT GCC TTC TAC CAA ATA CCC-3' R:5'-CTT CCT GAG AGA AGC AAC ACC-3'	2	55 °C	116–146	Sunden <i>et al.</i> , 1993
CSRM60	F:5'-AAG ATG TGA TCC AAG AGA GAG GCA-3' R:5'-AGG ACC AGA TCG TGA AAG GCA TAG-3'	10	55 °C	79–115	BCM, 2006
CSSM66	F:5'-AAT TTA ATG CAC TGA GGA GCT TGG-3' R:5'-ACA CAA ATC CTT TCT GCC AGC TGA-3'	14	60 °C	171–209	Barendse <i>et al.</i> , 1994
ETH10	F:5'-GTT CAG GAC TGG CCC TGC TAA CA-3' R:5'-CCT CCA GCC CAC TTT CTC TTC C-3'	5	58 °C	198–234	Toldo <i>et al.</i> , 1993
ETH225	F:5'-GAT CAC CTT GCC ACT ATT TCC T-3' R:5'-GTG TCT TAC ATG ACA GCC AGC TGC TAC T-3'	9	64 °C	132–166	Steffen <i>et al.</i> , 1993
ETH3	F:5'-GAA CCT GCC TCT CCT GCA TTG G-3' R:5'-GTG TCT TAC TCT GCC TGT GGC CAA GTA GG-3'	19	57 °C	90–135	Toldo <i>et al.</i> , 1993
ILST006	F:5'-TGT CTG TAT TTC TGC TGT GG-3' R:5'-ACA CGG AAG CGA TCT AAA CG-3'	7	58 °C	277–309	Brezinsky <i>et al.</i> , 1993
INRA023	F:5'-GAG TAG AGC TAC AAG ATA AAC TTC-3' R:5'-TAA CTA CAG GGT GTT AGA TGA ACT C-3'	3	55 °C	194–236	Vaiman <i>et al.</i> , 1994
RM067	F:5'-TGA GTA ATG CAA TAG ATA CAG TAT T-3' R:5'-GCT TTG GCC ATA TGA AGA GCT TT-3'	4	60 °C	83–101	Kossarek <i>et al.</i> , 1993
SPS115	F:5'-AAA GTG ACA CAA CAG CTT CTC CAG-3' R:5'-G TGT CTT AAC GAG TGT CCT AGT TTG GCT GTG-3'	15	55 °C	240–270	BCM, 2006
TGLA122	F:5'-AAT CAC ATG GCA AAT AAG TAC ATA C-3' R:5'-CCC TCC TCC AGG TAA ATC AGC-3'	21	58 °C	133–193	Barendse <i>et al.</i> , 1994
TGLA126	F:5'-CTA ATT TAG AAT GAG AGA GGC TTC T-3' R:5'-TTG GTC TCT ATT CTC TGA ATA TTC C-3'	20	55 °C	104–133	Barendse <i>et al.</i> , 1994
TGLA227	F:5'-GGA ATT CCA AAT CTG TTA ATT TGC T-3' R:5'-ACA GAC AGA AAC TCA ATG AAA GCA-3'	18	55 °C	63–115	Barendse <i>et al.</i> , 1994
TGLA53	F:5'-GCT TTC AGA AAT AGT TTG CAT TCA-3' R:5'-TGT CTT ATCT TCA CAT GAT ATT ACA GCA GA-3'	16	58 °C	147–197	Barendse <i>et al.</i> , 1994

For each locus, the forward (F) and reverse (R) base sequences, chromosomal location (Chr), annealing temperature (Ta), size range (base pairs) and source references are given. BCM: Baylor College of Medicine

The amplified products were separated by denaturing poly-acrylamide gel electrophoresis in an 8% gel concentration on an OmniPAGE vertical gel system (Biocom Direct, UK). The gels were then fixed with 7.5% (v/v) glacial acetic acid (CH<sub>3</sub>COOH) in deionized water (H<sub>2</sub>O), washed with water and stained using ethidium bromide (0.5µg/ml ethidium bromide in 1X TBE buffer) for 45 minutes. Molecular microsatellite visualization was done on the Infinity ST5 imaging system, and banding patterns and molecular weight data were generated using Vision Capt software (VilberLoumat, France).

During the primer screening exercise, all 26 primer pairs amplified successfully. However, based on the degree of polymorphism revealed by the primers, a panel consisting of only 16 primers was retained for analysing the rest of the samples (Table 2). The primers that did not make it into the final panel included HAUT27, HAUT24, ETH152, ILST005, INRA5, INRA37, INRA63, INRA124, INRA126, INRA189, and BM861. Although marker RM067 was not part of the original 26 primers that were screened, it was included in the final panel as it was used routinely in the laboratory. DNA samples were shipped to the Veterinary Genetics Laboratory (VGL) of the University of Pretoria, South Africa, for further analyses.

A 10-µl PCR reaction mixture was prepared with primer mix (2.8µl, 0.08–1.00 M), molecular grade water (1.2µl), Kapa Master Mix (5µl), and 20ng template DNA sample (0.5–1µl). DNA amplification was performed by multiplex PCR in a Perkin Elmer GeneAmp PCR System® 9700 (Applied Biosystems, CA, USA). The thermal cycler was programmed to run under these conditions: 95 °C for 3 minutes, 30 cycles of denaturation at 95 °C for 15 seconds, specific marker annealing temperature for 30 seconds, and extension at 72 °C for 30 seconds, followed by a final extension at 72 °C for 10 minutes and held at 4 °C. After amplification, the PCR products were quantified using 3% agarose gel in TBE buffer stained with ethidium

bromide and visualized under a UV trans-illuminator. DNA fragments were separated by capillary electrophoresis in an ABI Prism 3730 genetic analyser (Applied Biosystems, USA). GeneMapper version 4.1 was used to determine the fragment sizes in base pairs. Marker allele peaks and scores (integer values in base-pairs) were converted to an MS Excel datasheet for statistical analyses.

Statistical analyses were carried out on three datasets, namely a global sample of all test animals regardless of breed; a sample of test animals grouped into the three breeds Mashona, Tuli and Nkone; and a third dataset, which included the Brahman reference sample for population structure analysis. Allele score data were entered and managed in Microsoft Excel. Microsatellite Toolkit was used to check the data for errors and prepare input files for other software. Basic frequency-based population genetic parameters of gene diversity, total number of alleles (No),  $N_a$ ,  $N_e$ , breed private alleles (PAs), allele sizes and allele ranges (in base pairs) were estimated using Arlequin 3.5 (Excoffier & Lischer, 2015). Observed ( $H_o$ ), and expected ( $H_e$ ) heterozygosity (Nei, 1987) in each population and overall and an exact test of departure from Hardy-Weinberg Equilibrium (HWE) were carried out using a 30 000 Markov chain length and 10 000 dememorization steps using the same software. Polymorphism information content (PIC) of each marker per breed and overall was calculated in Microsoft Excel using the formula:

$$PIC = 1 - \sum_{i=1}^k (P_i)^2,$$

Where:  $P_i$  is the frequency of the  $i^{th}$  allele

$k$  is the total number of alleles at the locus (Botstein *et al.*, 1980)

To determine the level of genetic differentiation among the populations, fixation indices ( $F_{IS}$ ,  $F_{IT}$ , and  $F_{ST}$ ) and population pair-wise  $F_{ST}$  values (Weir & Cockerham, 1984) were calculated using FSAT 2.9.3 (Goudet, 2001). Analysis of molecular variance (AMOVA) was done in Arlequin 3.5 (Excoffier & Lischer, 2015). Significance of fixation indices was determined by running 1 000 permutations of the data.  $F_{ST}$  values were used to calculate the effective migration rate ( $Nm$ ) in Genalex 6.501 (Peakall & Smouse, 2012) using the formula:

$$Nm = 0.25(1 - F_{ST})/F_{ST}$$

Genetic relationships were inferred from principal coordinate analysis (PCoA) using the covariance matrix of Nei's genetic distance ( $D_A$ ) and unbiased genetic distances ( $D_S$ ) measures. Visualization of phylogenetic relationships among breeds was done by constructing dendrograms from pair-wise matrices of  $D_A$  using the agglomerative hierarchical clustering un-weighted pair group with arithmetic mean (UPGMA) method. Trees were drawn using the DendroUPGMA online application (Garcia-Vallvé & Puigbo, 2015) and visualized in TreeView (Page, 1996). Bootstraps of 1000 replicates were performed to provide confidence statements about the groupings of the breeds and test the reliability of the clusters. Population differentiation was further evaluated by performing supervised model-based clustering with prior population information in Arlequin 3.5 (Excoffier & Lischer, 2015). The purpose of this exercise was to determine whether allelic composition was independent of breed assignment (Frankham *et al.*, 2002; Lenstra *et al.*, 2012).

## Results

The marker diversity and polymorphism parameters across the entire sample are summarized in Table 3. A total of 119 alleles were observed from the 16 loci studies, yielding a mean of 7.4 alleles per locus. The  $N_o$  ranged from 4 (RM067) to 11 (TGLA122), with median and modal number of 8 alleles per locus. Mean ( $N_a$ ) and effective ( $N_e$ ) number of alleles per locus were 5.17 and 3.46 respectively, with no significant differences ( $P > 0.05$ ) among the three study populations. Average gene diversity over all loci was 0.74. Based on PIC analyses, all markers were informative, with TGLA53 (PIC 0.768) and RM067 (PIC 0.470) being the most and least informative, respectively.  $H_o$  (0.73) was higher than HWE  $H_e$  (0.71) proportions. However, total heterozygosity across the entire sample averaged 0.75.  $H_e$  varied between 0.49 (RM067) and 0.81 (TGLA53), while  $H_o$  ranged from 0.51 (SPS115) to 0.87 (BM2113). Three marker loci (BM1824, ETH225, and ETH3) were found to deviate significantly ( $P < 0.05$ ) from HWE expected proportions. TGLA53 and TGLA227 showed significant ( $P < 0.05$ ) departure from HWE in Tuli and Nkone populations, respectively. In the Mashona population, two loci, namely BM1824 ( $P < 0.001$ ) and ILST006 ( $P < 0.05$ ), deviated from HWE proportions.

Lists of observed alleles and breed-specific genetic variability parameters are given in Tables 4 and 5, respectively. Thirty-four out of a total of 119 alleles (29% of total) were unique to specific breeds. At 17, Tuli had the highest number of breed PAs. Rare alleles (with a frequency  $< 5\%$ ) were observed in the Mashona and Tuli populations, and none were observed in Nkone. Shannon's information index ( $I$ ) was moderate as all cattle breeds presented a value distant from zero with an overall mean of 1.343, and no significant differences between breeds. Similarly, PIC values averaged 0.682, 0.666, and 0.687 in Mashona, Nkone and

**Table 3** Genetic diversity parameters for 16 microsatellite loci analysed in three Zimbabwean Sanga cattle breeds

Locus	Allele sizes	No	Na	Ne	PIC	Ho	He	Ht	HWE*
BM1818	260–268	5	4.333	3.466	0.693	0.807	0.74	0.771	ns
BM1824	170–188	7	4.333	2.801	0.634	0.671	0.658	0.659	***
BM2113	121–141	8	6.000	4.414	0.737	0.869	0.802	0.817	ns
CSRM60	92–114	8	6.000	3.825	0.729	0.836	0.764	0.825	ns
CSSM66	179–199	8	4.667	2.595	0.572	0.731	0.624	0.724	ns
ETH10	201–225	7	5.667	4.266	0.719	0.804	0.794	0.801	ns
ETH225	140–156	8	5.667	3.038	0.661	0.639	0.697	0.818	*
ETH3	103–127	6	4.667	3.113	0.617	0.675	0.698	0.744	*
ILST006	284–302	8	6.000	4.332	0.748	0.847	0.801	0.819	ns
INRA23	196–214	7	4.667	3.378	0.694	0.638	0.711	0.731	ns
RM067	90–98	4	3.333	1.907	0.470	0.513	0.491	0.494	ns
SPS115	246–260	6	3.667	2.050	0.480	0.511	0.536	0.571	ns
TGLA122	137–169	11	5.333	3.490	0.700	0.774	0.747	0.831	ns
TGLA126	113–125	7	5.667	4.058	0.732	0.868	0.782	0.845	ns
TGLA227	77–103	9	6.000	3.984	0.665	0.786	0.748	0.805	ns
TGLA53	158–184	10	6.667	4.678	0.768	0.800	0.806	0.841	ns
Mean	-	7.438	5.167	3.462	0.664	0.736	0.712	0.756	-
SD	-	1.750	0.182	0.163	0.059	0.102	0.103	0.103	-

Parameters include allele size range (in bp), allelic diversity (No, observed number of alleles; Na, mean number of alleles; Ne, effective number of alleles), marker informativeness (PIC, polymorphism information content), heterozygosity (Ho, observed; He, expected; Ht, total),  $\chi^2$  test for HWE. \*HWE: ns: not significant, \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$

**Table 4** List of alleles genotyped from 16 microsatellite loci in Mashona, Nkone and Tuli cattle breeds of Zimbabwe

Locus	Mashona	Nkone	Tuli
BM1818	262, 264, 266, 268	260, 262, 264, 266	260, 262, 264, 266, 268
BM1824	172, 174, 186, <u>188</u>	174, 176, 186	172, 174, 176, <u>182</u> , <u>184</u> , 186
BM2113	121, 129, 133, 135, 141	121, 129, 133, 135, 139, 141	121, <u>125</u> , 133, 135, <u>137</u> , 139, 141
CSRM60	92, 96, 100, 102, <u>104</u> , <u>108</u> , 112, 114	92, 96, 100, 102, 112, 114	92, 96, 100, 102, 112, 114
CSSM66	185, 187, 189, <u>193</u>	185, 187, 189, <u>195</u>	<u>179</u> , <u>183</u> , 185, 187, 189, <u>199</u>
ETH10	209, 211, 217, 219, 221	209, 211, 217, 219, 221, <u>225</u>	209, 211, 217, 219, 221, <u>223</u>
ETH225	140, 144, <u>146</u> , 148, 150, 152	140, 148, 152, 158	140, 144, 148, 150, <u>154</u> , 158
ETH3	<u>103</u> , 117, 121, 125, 127	115, 117, 121, 125, 127	115, 117, 121, 125
ILST006	284, 288, 292, 294, 296, 300, 302	284, 288, 292, 294, 300, 302	284, 292, 294, 296, <u>298</u> , 302
INRA23	196, 198, <u>202</u> , 208, 214	196, 198, 206, 214	198, 206, 208, <u>210</u> , 214
RM067	90, 92, 94, 98	90, 92, 94	90, 92, 98
SPS115	246, 248, <u>256</u> , 260	246, 248, <u>250</u>	246, 248, <u>254</u> , 260
TGLA122	137, <u>139</u> , 143, 151, <u>157</u> , <u>159</u> , <u>161</u>	137, 143, <u>149</u> , 163	137, <u>141</u> , 143, 151, <u>169</u>
TGLA126	115, 117, 119, 121, 123, 125	<u>113</u> , 115, 117, 119, 121	115, 117, 119, 121, 123, 125
TGLA227	77, 81, 87, 89, 93, 97, 103	81, 83, 87, 89, 93, <u>95</u> , 97	77, 83, 87, 93, 103
TGLA53	160, 166, 172, 174, 176, 178, 184	160, 166, 172, 176, <u>180</u>	<u>158</u> , 160, 166, 174, 176, 178, <u>182</u> , 184

Underlined = alleles unique to only one breed

Tuli samples, respectively. Fixation index was significantly negative for Nkone and Tuli, indicating a significant excess of heterozygotes ( $P < 0.001$ ) in these populations. Estimates of  $H_o$ , including all loci and breeds (0.73), confirmed that the studied populations contain a high level of genetic diversity. Similar to the entire sample, breed estimates of  $H_o$  were significantly higher than HWE  $H_e$  proportions, except in the Mashona breed.

**Table 5** Summary of genetic diversity parameters in three Zimbabwean Sanga cattle breeds using 16 microsatellite markers

Parameter	Breed			Overall
	Mashona	Nkone	Tuli	
Observed number of alleles (No)	87.000	73.000	88.000	119.000
Number of private alleles (PA)	12.000	6.000	17.000	34.000
Mean number of alleles (Na)	5.438	4.562	5.500	5.167 ± 1.209
Effective number of alleles (Ne)	3.360	3.382	3.645	3.462 ± 0.163
Gene diversity over all loci	0.705	0.726	0.711	0.739 ± 0.375
Shanon's Information Index (I)	1.355	1.289	1.383	1.343 ± 0.044
Marker PIC	0.682	0.666	0.687	0.664 ± 0.009
Observed heterozygosity ( $H_o$ )	0.704	0.760	0.742	0.736 ± 0.102
Expected heterozygosity ( $H_e$ )	0.705	0.726	0.711	0.712 ± 0.115
$F_{IS}$	0.003	-0.052	-0.047	-0.028

Breed differentiation was estimated by the infinitesimal model ( $F_{ST}$ ) and  $D_A$  measures using number of alleles. Breed pairwise  $F_{ST}$  and  $D_A$  values (Table 6) showed great genetic divergence for Brahman from the three Sanga cattle genotypes and less divergence among the Sanga cattle genotypes. The Sanga breeds were significantly differentiated ( $P < 0.001$ ), indicating that the breeds could be considered genetically independent. Mashona and Nkone ( $F_{ST} = 0.093$ ) were the most genetically divergent pair compared with both Mashona and Tuli ( $F_{ST} = 0.082$ ) and Tuli and Nkone ( $F_{ST} = 0.069$ ). Wright's F-statistics  $F_{IT}$ ,  $F_{ST}$ , and  $F_{IS}$ , computed for the entire dataset were 0.059, 0.084 and -0.028, respectively (Table 7). The overall estimate of genetic differentiation ( $F_{ST}$ ) was moderate and significant ( $P < 0.001$ ), indicating that 8.4% of the total genetic variation corresponds to differences among breeds. All microsatellite marker loci contributed to breed differentiation ( $F_{ST}$ ) with the most powerful markers being CSSM66 (25%), ETH225 (20.6%), and TGLA122 (13.8%). Three markers, namely RM067, BM1824 and ETH10, were the least powerful, with lower  $F_{ST}$  values of 0.7%, 1.0%, and 1.1%, respectively. Overall estimate of inbreeding ( $F_{IT}$ ) was significantly positive (0.059) with three loci (BM1824, BM2113 and RM067) contributing to heterozygote excess and the rest to heterozygote deficiency. Within breeds, all loci except ETH225, ETH10, and INRA23 contributed to heterozygote excess ( $F_{IS}$ ). The mean number of migrants per generation ( $Nm$  estimate) of 3.420 was significantly different from zero and indicates moderate rates of gene flow from one breed to another. Analysis of molecular variance (AMOVA) revealed that most of the population variance could be explained by within-individual variability (91.8%) and less so (8.2%) by variation among breeds.

Population differentiation was further evaluated by performing supervised model-based clustering using allele frequencies and allele sharing matrices as implemented in Arlequin. All individuals were correctly assigned to their self-population. Four population clusters were observed on a two-dimensional structure diagram, namely two Mashona clusters, a distinct Tuli cluster, and a mixed Nkone-Tuli cluster (Figure 1). Phylogenetic relationships using the UPGMA algorithm separated the Brahman first, followed by the Nkone while the Tuli and Mashona breeds more or less clustered together (Figure 2). PCoA was carried out in GenAiEx 6.5 and corroborated these findings. The first two dimensions of the PCoA explained 100% of the variation (Figure 3). The first principal coordinate, which accounted for 62.73% of the total genetic variability, distinguished clearly the Mashona breed from the other two Sanga cattle populations. The second principal coordinate, which summarized 37.27% of the variation, separated the Tuli breed from Brahman and Nkone populations.

**Table 6** Pairwise estimates of genetic differentiation and Nei's genetic distance measures between Zimbabwean Sanga cattle breeds

Breed	Brahman	Mashona	Nkone	Tuli
Brahman	-	0.995	0.661	0.864
Mashona	0.203	-	0.432	0.333
Nkone	0.164	0.093	-	0.342
Tuli	0.190	0.082	0.069	-

Pairwise  $F_{ST}$  values are shown below the diagonal and  $D_A$  values above the diagonal. All  $F_{ST}$  values are significant ( $P < 0.001$ ).

**Table 7** Global F-statistics and estimates of gene flow across three Sanga cattle populations of Zimbabwe

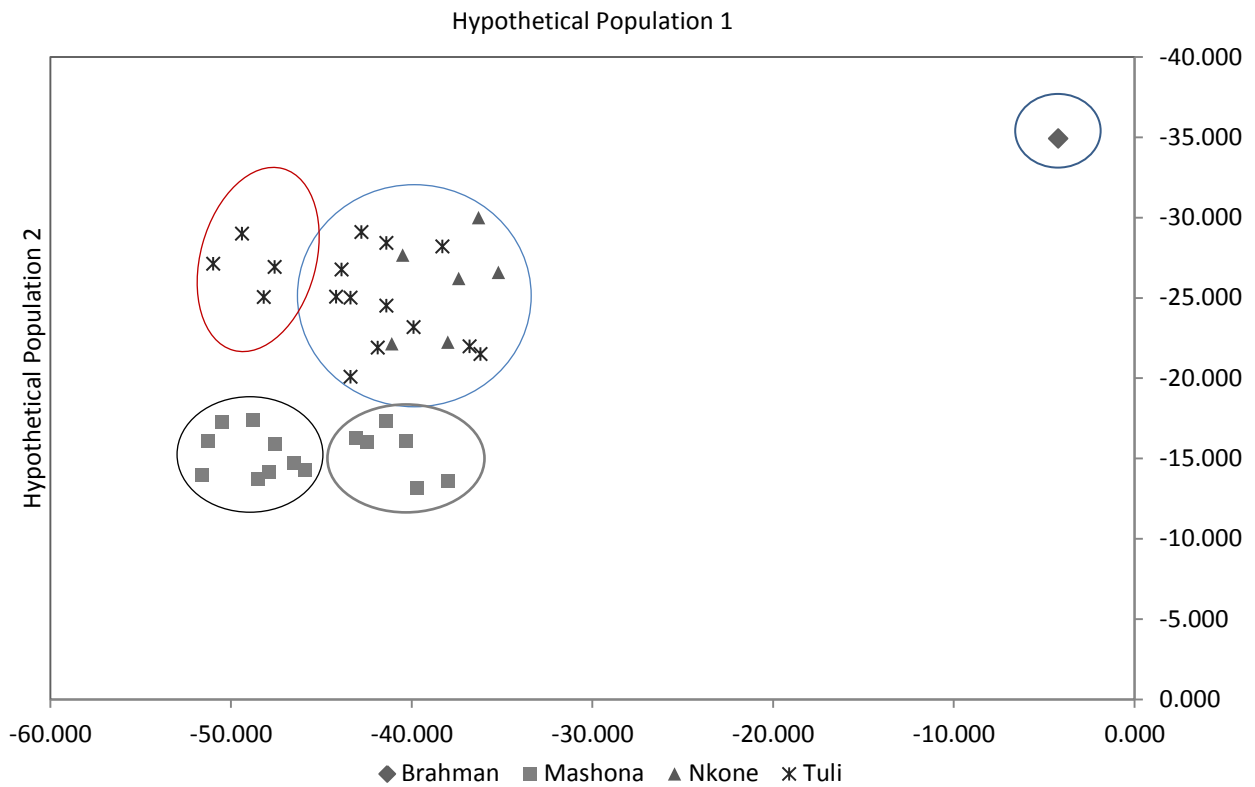
Locus	$F_{IT}$ (F)	$F_{ST}$ ( $\theta$ )	$F_{IS}$ (f)	Nm
BM1818	0.050	0.070	-0.021	3.367
BM1824	-0.114	0.010	-0.125	8.055
BM2113	-0.024	0.039	-0.066	4.820
CSRM60	0.043	0.097	-0.059	2.036
CSSM66	0.117	0.250	-0.177	1.355
ETH10	0.029	0.011	0.018	5.718
ETH225	0.244	0.206	0.047	1.138
ETH3	0.077	0.094	-0.019	2.252
ILST006	0.033	0.046	-0.013	4.488
INRA23	0.127	0.037	0.094	3.702
RM067	-0.014	0.007	-0.021	6.230
SPS115	0.075	0.075	0.000	2.384
TGLA122	0.129	0.138	-0.011	1.653
TGLA126	0.012	0.079	-0.072	2.192
TGLA227	0.061	0.084	-0.026	2.227
TGLA53	0.036	0.051	-0.016	3.106
Overall	0.059	0.084	-0.028	3.420

Nm was calculated using the formula:  $Nm = 0.25(1 - F_{ST})/F_{ST}$

### Discussion

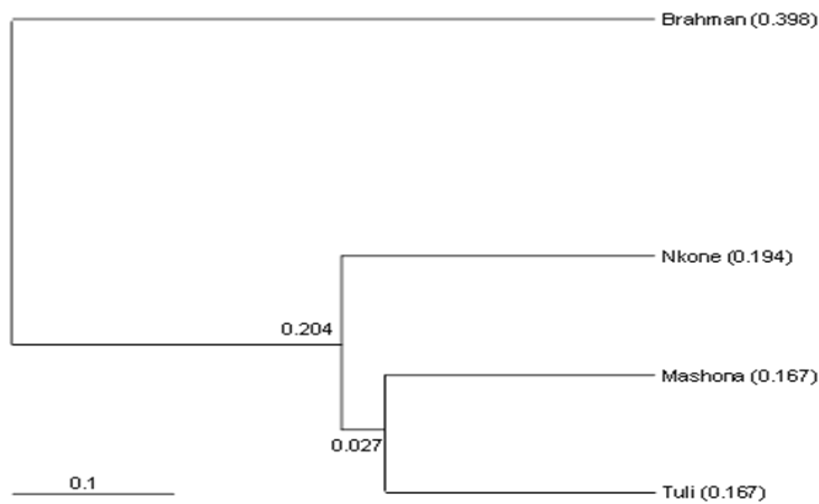
This is the first report on the genetic structure and diversity of improved Sanga cattle breeds in Zimbabwe. The Mashona, Tuli and Nkone, together with the Brahman, dominate cattle production systems and agricultural livelihoods in the country. Genetic characterization is free from environmental influences, and when inferred from highly polymorphic molecular markers such as microsatellites, it is fundamental for decision making in conservation and utilization programs (Groeneveld *et al.*, 2010). All 16 microsatellite loci exhibited a high degree of polymorphism and a large number of alleles, indicating the suitability of these markers for genetic variability studies in the Zimbabwean cattle. For breed differentiation, the three most powerful markers of the 16 were CSSM66, ETH225 and TGLA122, and the three least powerful were RM067, BM1824 and ETH10. Few loci deviated from HWE expected proportions in the entire sample and in individual breeds. Since these deviations were not consistent across breeds, they were considered to be caused by a Type I error sampling effect (FAO, 2011). The level of variation depicted by number and size of alleles at each microsatellite locus is a useful measure of genetic variability and differentiation among breeds within a species (Sharma *et al.*, 2013). All markers except RM067 exceeded FAO's recommended minimum threshold of five alleles per locus, which is required for estimation of genetic differences among animal breeds (FAO, 2011; Sanarana *et al.*, 2016). These microsatellite loci were thus informative enough for exploring within-breed genetic variability and breed differentiation in Zimbabwean Sanga cattle.





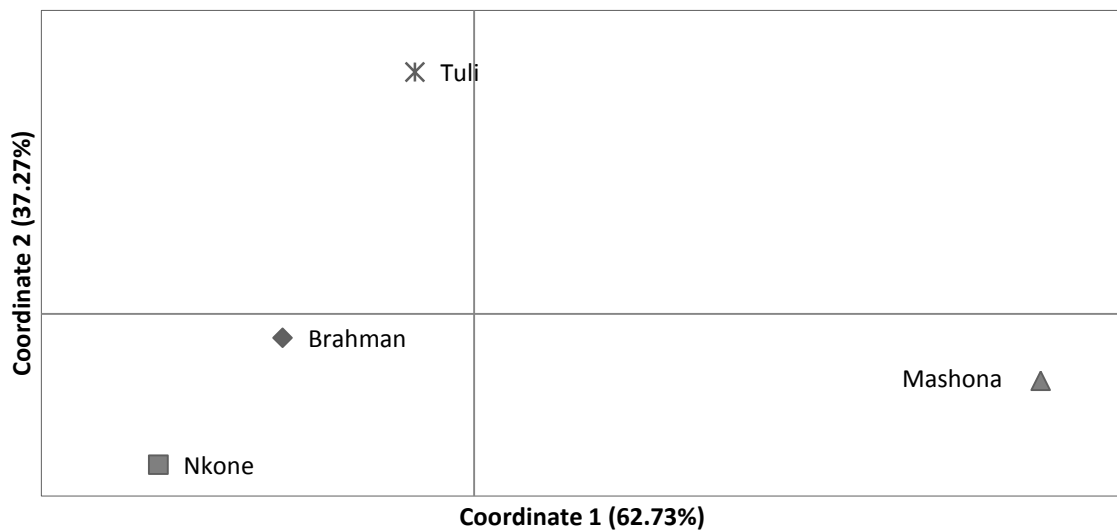
Note: The highest (least negative) value indicates the most likely population

**Figure 1.** Log likelihoods for cluster assignment of different breeds of animals on a two-dimensional scale diagram



Note: Numbers in this graph represent percentage occurrence of a node in 10 000 bootstrap replicates. The linear scale relates the branch lengths to units of  $D_A$ . The root of the tree was placed at the midpoint of the longest branch separating the Brahman from the other breed groups

**Figure 2** Hierarchical clustering of Zimbabwean Sanga cattle breeds and Brahman based on Nei's genetic and Euclidian distance measures



**Figure 3** Principal coordinates analysis based on covariance matrix of Nei's unbiased genetic distance for three Zimbabwean Sanga cattle breeds

Private alleles are useful for the identification and measurement of the genetic distinctiveness of a population or breed (Ngono-Ema *et al.*, 2014; Sanarana *et al.*, 2016). A total of 119 alleles were observed, with 34 being unique to specific breeds. Tuli had the highest number (17) of PAs, showing that the breed has been developed as a distinctive genotype without significant admixture from the other two breeds. Breed PAs have been observed in diversity studies on indigenous cattle in Ethiopia (Edea *et al.*, 2013; Ngono-Ema *et al.*, 2014), South Africa (Pienaar *et al.*, 2015; Sanarana *et al.*, 2016), and Mozambique (Bessa *et al.*, 2009). The Na and Ne alleles revealed no appreciable differences in the level of genetic variability among these breeds. The No of 7.4 alleles per locus was comparable with those for several other indigenous cattle breeds of Africa. For instance, Sanarana *et al.* (2016) reported 6.5 alleles per locus in South African Nguni cattle ecotypes. In Mozambique, Bessa *et al.* (2009) found allelic diversity of 5.9–6.4 alleles per locus. However, Ndiaye *et al.* (2015) reported 10.5 alleles per locus among Senegalese cattle, and Sharma *et al.* (2013) reported 9.0 alleles per locus in Indian cattle breeds. Although comparable with other studies, genetic variability in this study was moderate and similar between breeds. This could be explained by the relatively small sample size for each population, the highly purposive sampling nature (only improved populations under in-situ conservation) and a common ancestral history among sampled individuals. Tuli had greater diversity compared with the other two breeds, probably as a reflection of its larger population size. Tuli is the second most widely used breed in the country after Brahman.

The current study revealed higher heterozygosity values compared with 50–63%, 70%, 71%, 68%, and 70% reported in indigenous cattle from Mozambique (Bessa *et al.*, 2009), South Africa (Sanarana *et al.*, 2016), Senegal (Ndiaye *et al.*, 2015), Cameroon (Ngono-Ema *et al.*, 2014), and India (Sharma *et al.*, 2013), respectively. Higher  $H_o$  compared with  $H_e$  and fixation indices suggested that breeds were in HWE. All these researchers (above) observed heterozygote deficiency. Generally, populations subjected to long-term natural selection, adaptation and historic admixture have been observed to exhibit high heterozygosity levels (Frankham *et al.*, 2002; Sanarana *et al.*, 2016).

Slightly negative  $F_{IS}$  estimated for all the study populations may indicate random sampling error or a relatively recent migration of breeding individuals from one station to another, followed by successful mating, negating the possibility of inbreeding (Frankham *et al.*, 2002). This argument seems most likely since individuals within breed were less related than would be expected under a model of random mating. Although the population sizes are small, mild selection pressure and inter-population gene flow may have contributed to observed heterozygote excess compared with expected proportions. The estimated population gene flow ( $Nm = 3.4$ ) value indicated that there was just adequate gene flow among populations to negate the effects of genetic drift and inbreeding ( $Nm > 1$ ), but the populations were not mating at random ( $Nm < 4$ ) (Frankham *et al.*, 2002; Groeneveld *et al.*, 2010). This signifies low gene flow between breeds, resulting in moderately high measures of genetic differentiation. Ngono-Ema *et al.* (2014) obtained a similar result of  $Nm$

= 3.816 among Cameroonian cattle populations, while Kugonza *et al.* (2011) and Sharma *et al.* (2013) found considerably higher  $N_m$  values among Ankole and Indian cattle populations.

Overall, genetic divergence ( $F_{ST}$  of 8.4%) revealed moderate but significant genetic differentiation among the three Sanga cattle genotypes studied using the scale proposed by Nei (1987). This finding was comparable with those reported by Bessa *et al.* (2009) for indigenous breeds of Mozambique. Sanarana *et al.* (2016), Ngono-Ema *et al.* (2014), and Kugonza *et al.* (2011) reported lower genetic differentiation values of 4.8 % (South African Nguni ecotypes), 6.1% (Cameroonian breeds), and 4.6 % (Ankole cattle), respectively. Edea *et al.* (2013) and Zerabruk *et al.* (2012) reported very low levels of differentiation among Ethiopian cattle populations ( $F_{ST} = 1\%$  and  $1.1\%$ ) using single nucleotide polymorphisms (SNPs) and microsatellites, respectively. However, higher  $F_{ST}$  values of 13.3% were reported for Indian cattle breeds (Sharma *et al.*, 2013). The moderate value of genetic differentiation may be attributed to recent divergence of the breeds from a common ancestry, reproductive isolation, or moderate selection pressure in the populations from which samples were collected (Feliuss *et al.*, 2014).

The partitioning of the genetic variation from an AMOVA revealed that 8.4% of variation was among breeds. A similar pattern of variance partitioning was observed in similar studies (Edea *et al.*, 2013; Ngono-Ema *et al.*, 2014; Sanarana *et al.*, 2016), in which 90% or more of the variation is contained within breeds. In Ankole cattle breeds, Kugonza *et al.* (2011) reported within-population diversity to be 95.5 %. PCoA and UPGMA clustering revealed four major clusters for the three study populations. PCoA extracts from the dataset successive coordinates that correspond mathematically to combinations of variables and independent vectors of the total variation (Lenstra *et al.*, 2012). Phylogenetic reconstructions complement the other analysis methods by reducing highly multidimensional data to a two-dimensional diagram and indicating common ancestry at various taxonomic levels (individuals, strains, breeds, and species). However, they do not account for the effects of admixture between breeds and strains (Lenstra *et al.*, 2012; Sharma *et al.*, 2013). The first principal coordinate separated the Mashona and the second separated the Tuli from the Brahman and Nkone breeds. Therefore, in the multivariate space defined by the two first principal coordinates, the three Sanga cattle breeds of Zimbabwe are genetically distinct and completely separated. However, the Mashona is genetically more distant from the Nkone compared with the Tuli. These breeds have similar evolutionary patterns, having descended from the same Ethiopian ancestral cattle (Ramsay, 2010) and then separated geographically through ethnic or tribal ownership (Rowlands *et al.*, 2003; Makina *et al.*, 2016).

Carr *et al.* (1966) used biochemical markers (serum transferrin type and haemoglobin loci) on indigenous cattle genotypes of Zimbabwe and Zambia (Nkone, Mashona, Tuli, Angoni and Afrikaner) and found them to be distinct breeds. Holness (1992) speculated that the indigenous breeds in Zimbabwe (Mashona, Nkone, and Tuli) developed from Bantu stock in which the original genotypes must have been Mashona, and that there are close genetic similarities, especially between the Mashona and Nkone, although the animals fall into distinct breed groups. The results of this study, namely that Sanga cattle in Zimbabwe fall into distinct breed groups with moderate differentiation, are consistent with findings by Carr *et al.* (1966) and speculation by Holness (1992). However, contrary to speculation by Holness (1992), the Mashona have closer genetic similarities with Tuli compared with the Nkone. Molecular microsatellite markers used in this study were effective in revealing these genetic differences.

It may be postulated that the observed moderate allelic diversity levels and moderate genetic differentiation are due mainly to progenitor population diversity; bottleneck effects such as droughts and disease outbreaks; founder events of breed formation; intensity of within-breed selection for type, adaptability, reproductive fitness, and production traits; post-breed formation; adaptation to local conditions and farming systems; and recent divergence of the breeds from each other (d'Hotman & Hatendi, 1998; Feliuss *et al.*, 2014). The founder population of the local Sanga breeds in Zimbabwe consisted of small groups (fewer than 100 per breed) of cows that faced significant selective pressure for type, adaptability and reproductive fitness. Earlier, their progenitor populations had gone through two major genetic bottlenecks as a result of the Rinderpest (1896–1898) and East Coast Fever (1900–1906) disease epidemics, which decimated 90% of the original cattle in the country. These were followed by introgression as the government imported breeding animals from Zambia to restock (Holness, 1992). Later, successive droughts had a significant effect on national herd size. Other forces that may have shaped the current status of diversity include migration, mutation and genetic drift, since conservation populations under study are small fragmented populations, numbering a few hundred per site (Assan, 2012). It is important therefore that some form of gene flow from the unimproved native cattle population to the improved (conservation) population should be promoted so that allelic diversity can continue to be widened.

Genetic evaluation is relevant since genetic variability is beneficial for population survival and improvement, and genetic uniqueness is important for prioritizing populations for conservation (Rege *et al.*, 2011). This study also helped to identify the most informative microsatellite markers for genetic diversity

studies, mapping quantitative trait loci, marker assisted selection and parentage verification in these breeds. Sanga cattle breeds are widely used in commercial and communal livestock production systems of the country owing to their unique adaptive traits. Therefore, it is of utmost interest to the country to maintain them as distinct populations and maintain the genetic variability that each contributes to total cattle diversity.

## Conclusions

This study was the first attempt to genetically characterize the improved Zimbabwean Sanga cattle breeds of Mashona, Nkone, and Tuli, using microsatellite markers. These markers displayed sufficient polymorphism, and are therefore useful tools in the study of genetic diversity and uniqueness in Sanga cattle breeds of Zimbabwe. The analyses revealed that a significant amount of variation is maintained in these local breeds and that all breeds are distinct genetic entities. These three breeds therefore constitute a valuable reservoir of genetic diversity for future breeding and are viable targets for conservation. Although these conservation populations are small, the breeding strategy used is good enough to negate the effects of inbreeding and genetic drift. However, it may be necessary to promote some gene flow from the unimproved population to widen genetic diversity in these conservation populations. The study could therefore serve as a launch pad for future genetic diversity studies on conservation and utilization of Sanga cattle genotypes in Zimbabwe, including the lesser known ones. For instance, further investigation of the association between genetic markers and phenotypic parameters could better enable exploitation of valuable genetic material.

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## Author's Contributions

EG conceived the research, collected data, carried out molecular and statistical analyses, and wrote the manuscript. FPC conceived the research with EG, carried out laboratory analyses, and reviewed the final manuscript. SMM and FC reviewed the research concept and manuscript. TWS reviewed the statistical analyses and manuscript.

## Conflict of Interest Declaration

The authors declare that they have no affiliations with or involvement in any organization or entity with any interest (financial or non-financial) in the subject matter or materials discussed in this manuscript.

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