

Growth differentiation factor 9 gene variants in Sudanese desert sheep ecotypes

A.S. Ali¹, M.T. Ibrahim¹, M.M. Mohammed¹, A.A. Elobied¹ & G. Lühken^{2#}

¹College of Animal Production Science and Technology, Sudan University of Science and Technology, Khartoum, North-Sudan

²Department of Animal Breeding and Genetics, Justus Liebig University, Giessen, Germany

(Received 8 March 2016; Accepted 7 September 2016; First published online 12 November 2016)

Copyright resides with the authors in terms of the Creative Commons Attribution 2.5 South African Licence. See: <http://creativecommons.org/licenses/by/2.5/za> Condition of use: The user may copy, distribute, transmit and adapt the work, but must recognise the authors and the South African Journal of Animal Science.

Abstract

Certain variants in the growth differentiation factor 9 (*GDF9*) gene have major effects on the ovulation rate in sheep. The aim of this study was to analyse *GDF9* variability in the Sudanese desert sheep ecotypes Ashgar, Dubasi and Watish, and to test identified variants for association with litter size. For this purpose, ewes of these ecotypes with litter size records for at least two litters were sampled. The complete *GDF9* exon 2 was sequenced in a total of 28 ewes. An additional variant in exon 1 (c.260G>A) was genotyped by restriction-length polymorphism analysis in 97 ewes. Differences in genotype and allele frequencies of polymorphic positions between two groups differing in litter size (only a single lamb versus more than a single lamb) were tested for significance using Fisher's exact test. *GDF9* exon 2 variants c.477G>A and c.721G>A and exon 1 variant c.260G>A were found to be polymorphic in all three sheep ecotypes. Exon 2 variants c.471C>T and c.978 A>G were polymorphic in at least one ecotype. No significant associations were observed between allele and genotype frequencies of identified variants and litter size. This suggests that *GDF9* variants influencing ovulation are absent in these Sudanese sheep ecotypes, and therefore cannot be used to increase litter size within this population of sheep.

Keywords: Ashgar, Dubasi, fecundity, litter size, ovine fertility, Watish

Corresponding author: gesine.luehken@agrار.uni-giessen.de

Introduction

The growth differentiation factor 9 (*GDF9*) gene is located on chromosome 5 and contains two exons, divided by an intron of 1126 base pairs. The exons code for a propeptide with 453 amino acids (Bodensteiner *et al.*, 1999).

GDF9 belongs to the transforming growth factor β superfamily, and has a vital role in ovarian follicular development and ovulation rate. It has been widely studied in humans, sheep, and goats (Elvin *et al.*, 1999; McNatty *et al.*, 2005). In sheep, numerous mutations in the *GDF9* coding sequence have been reported, with one single exception all of them being located in the second exon (Table 1). Eight of these 11 single nucleotide polymorphisms (SNPs) cause amino acid substitutions, and some of them have an effect on ovulation rate and hence litter size. The three non-synonymous SNPs c.943C>T, c.1184C>T and c.1279A>C result in a phenotype of increased ovulation rate/litter size in heterozygous ewes, and infertility linked to hypoplasia of ovary and uterus in homozygous females (Hanrahan *et al.*, 2004; Juengel *et al.*, 2013; Nicol *et al.*, 2009; Souza *et al.*, 2014). Infertility due to the homozygous mutant genotype was not observed for two other non-synonymous SNPs, c.1111G>A (Vage *et al.*, 2013) and c.1034C>T (Silva *et al.*, 2011), which instead show an additive effect on ovulation rate and litter size.

As only the mature *GDF9* peptide is deemed biologically active (Paulini & Melo, 2011), mutations that are located proximal to the RRHR furin protease cleavage site (proximal to amino acid position 318) are regarded as not affecting the protein function (Hanrahan *et al.*, 2004). A single non-synonymous SNP in exon 1 (c.260G>A) is located before the furin cleavage site, and causes only a conservative substitution of amino acids (Arg87His) (Hanrahan *et al.*, 2004). However, this *GDF9* mutation was claimed by Barzegari *et al.* (2010) to be associated with infertility (genotype AA), and at least in combination with another mutation in the gene coding for the bone morphogenic protein 15 (*BMP15*) with higher ovulation rate (genotype AG) in Iranian sheep.

Associations of *GDF9* sequence variants at positions with no obvious impact on the gene function might be due to a linkage with undetected or until now not tested causal variants. Such a linkage was recently speculated by Albarella *et al.* (2015) for a silent G>A substitution they detected for the first time at

position 750 of *GDF9* in Bagnolese sheep. They observed a higher litter size in sheep with the genotype GG compared with AG ($P < 0.05$). The effect of the genotype AA was not tested because of its low frequency.

Table 1 Published sequence variants in the coding region of ovine *GDF9*

Position in coding sequence	Variant name(s)	Amino acid change	Breed	Variant first published by
Exon 1				
c.260G>A	G1	p.Arg87His	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
Exon 2				
c.471C>T	G2	p.Val157Val	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.477G>A	G3	p.Leu159Leu	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.531C>T		p.Asn177Asn	Bagnolese	Albarella <i>et al.</i> , (2015)
c.617G>A		p.Arg206Lys	Laticauda	Albarella <i>et al.</i> , (2015)
c.721G>A	G4	p.Glu241Lys	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.729G>T		p.Gln243His	Small Tail Han	Chu <i>et al.</i> , (2011)
c.750G>A		p.Arg250Arg	Thoka	Nicol <i>et al.</i> , (2009)
c.943C>T	FecG ^V	p.Arg315Cys	Ile de France	Souza <i>et al.</i> , (2014)
c.953G>T		p.Arg318Ile	Bagnolese	Albarella <i>et al.</i> , (2015)
c.978A>G	G5	p.Glu326Glu	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.994G>A	G6	p.Val332Ile	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.1034C>T	FecG ^{SI} /FecG ^E	p.Phe345Cys	Brazilian Santa Inês	Silva <i>et al.</i> , (2011)
c.1111G>A	G7	p.Val371Met	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.1184C>T	G8/FecG ^H	p.Ser395Phe	Cambridge & Belclare	Hanrahan <i>et al.</i> , (2004)
c.1203G>A		p.Val401Val	Bagnolese	Albarella <i>et al.</i> , (2015)
c.1279A>C	FecT ^T	p.Ser427Arg	Thoka	Nicol <i>et al.</i> , (2009)
c.1358G>A		p.Arg453His	Laticauda	Albarella <i>et al.</i> , (2015)

Sheep population in Sudan consists of about 39.6 million, representing 37.79% of the total Sudanese livestock population, which is approximately 104 million head. In recent years there has been growing interest in exporting desert-type Sudanese sheep to Arab countries (Ministry of Animal Resources, Fisheries and Ranges (MARFR), 2013). Desert sheep is one of the most widely distributed types in Sudan, representing about 65% of the total sheep population, and comprising seven regional ecotypes, namely Gezira (Ashgar and Dubasi), Watish, Butana, Bija, Meidob, and North Riverine Woolled Sheep (Mukhtar, 1985; El Hag *et al.*, 2001). Sulieman *et al.* (1990) found that Ashgar produced higher numbers of lambs per ewe (1.30) than Dubasi (1.18) and Watish (1.17).

The aim of this work was to analyse *GDF9* gene variability in the Sudanese desert sheep ecotypes Ashgar, Dubasi and Watish, and to test identified variants for association with litter size.

Ewes from the three sheep ecotypes (Figure 1) from various regions of Sudan were selected for sampling according to their history of litter size (River Nile and Khartoum states for Ashgar, Gazira state for Dubasi and Sinar state for Watish). Any selected ewe must have had at least two lambing records. The number of lambing records ranged from two to seven (on average 3.9 records).



Figure 1 Sudanese desert sheep ecotypes analysed in this study: A) Ashgar; B) Dubasi; C) Watish

Ewes were divided into two groups according to their average litter size. One group comprised ewes of all three ecotypes that gave birth to single lambs in all recorded lambings (hence the average of litter size was 1.0). The other group included ewes of all three ecotypes, which on average had more than a single lamb (average litter size per ewe in this group ranged from 1.5 to 3.0; the average litter size of the whole group was 2.1).

Blood samples (5 mL) were drawn from the jugular vein in EDTA vacutainer tubes. The genomic DNA was extracted from white blood cells according to Montgomery & Sise (1990).

Sampling of sheep was approved by the Federal Ministry of Animal Resources, Fishery and Ranges of North Sudan and in compliance with standard ethical norms.

To identify sequence variants in the second exon of *GDF9*, 28 samples were sequenced: 10 DNA samples each from Ashgar and Dubasi and eight from Watish. For each ecotype, 50% of the samples were selected from the single lamb group and the other 50% from the more than a single lamb group. To amplify two overlapping fragments covering the complete exon 2 of *GDF9*, these pairs of primers were designed using GenBank sequence AF078545.2 and the software Primer3 (Untergrasser *et al.*, 2012): forward primer 5'-GGCTTGAGAATGTGGGGAGAA-3' and reverse primer 5'-GGGACGATCTTACACCCTCA-3' to amplify a proximal fragment of 656 bp, and forward primer 5'-CACAAGTGCTCAGGCTTTTC-3' and reverse primer 5'-CATGAGGAAGGCAGCTGTTA-3' to amplify an overlapping distal fragment of 749 bp. Polymerase chain reaction (PCR) amplifications were carried out in a final volume of 50 μ L, including 0.2 mM dNTPs, 1 mM $MgCl_2$, 20 pmol of each primer, 1.5 U Go Taq Flexi Polymerase in 1-fold Colourless GoTaq Flexi buffer (Promega, Madison, Wis, USA) and 50 ng of DNA template. PCR reaction was run in a thermal cycler under these conditions: initial denaturation at 95 $^{\circ}C$ for 90 sec, followed by 35–40 cycles consisting of denaturation at 95 $^{\circ}C$ for 15 sec, annealing at 65 $^{\circ}C$ for 30 s, extension at 72 $^{\circ}C$ for 60 s, and a final extension at 72 $^{\circ}C$ for 5 min. Agarose gel electrophoresis and Nanodrop 2000 spectrophotometer (VWR International GmbH, Erlangen, Germany) were used to check PCR products for size, quality and quantity. PCR products were purified and then sequenced using Big Dye Terminator chemistry and the ABI 3130 Genetic Analyzer as recommended by the manufacturer (Applied Biosystems, Foster City, Calif, USA) with PCR forward primer (656-bp fragment) and reverse primer (749-bp fragment). Alignment and analysis of sequences from the samples was done with the software ChromasPro version 1.33 (Technelysium Pty Ltd, Tewantin, Australia).

The SNP in exon 1 of *GDF9* (c.260G>A) was genotyped in a total of 97 ewes with litter size records (35 Ashgar, 29 Dubasi and 33 Watish) by PCR-restriction fragment length polymorphism (RFLP) analysis using the *HhaI* restriction enzyme. Its cleavage site (GCGC) occurs only in the presence of the G allele. For amplification of a 357-bp fragment containing the polymorphic position c.260G>A of *GDF9* exon 1 and no additional *HhaI* cleavage site, these primers that were designed with Primer3 software were used: forward primer 5'-TGAGGCTGAGACTTGGTCCT-3' and reverse primer 5'-ATAAAGGAGTTGCCCTGCT-3'. PCR amplification was carried out in a final volume of 25 μ L, including 0.2 mM dNTPs, 2 mM $MgCl_2$, 20 pmol/ μ L of each primer, 1.0 U Go Taq Flexi Polymerase with 1 x Colourless Go Taq Flexi buffer (PROMEGA, Madison, Wis, USA) and 50 ng DNA template in a thermal cycler under these conditions: initial denaturation at 95 $^{\circ}C$ for 90 s, followed by 35 cycles consisting of denaturation at 96 $^{\circ}C$ for 15 s, annealing at 62 $^{\circ}C$ for 30 s, extension at 72 $^{\circ}C$ for 60 s, and a final extension at 72 $^{\circ}C$ for 5 min. The resulting PCR product was incubated with *HhaI* in 10 μ L final volume as recommended by the manufacturer of the enzyme (New England Biolabs GmbH, Frankfurt am Main, Germany). The resulting DNA fragments were separated on agarose gel (2.5%) and visualized by staining with Midori green (Nippon Genetics Europe GmbH, Dürren, Germany). Restriction fragment length polymorphism (RFLP) patterns were verified by sequencing PCR products with PCR forward primer as described before.

Genotype and allele frequencies were calculated for identified SNPs for all sheep and separately for each ecotype, and for the two groups of ewes with single and with more than a single lamb, respectively. Differences in genotype and allele frequencies between these two groups of divergent litter size were tested

for significance with Fisher's exact test using the program IBM SPSS Statistics for Windows, Version 20.0. Armonk, NY: IBM Corp.

Results and Discussion

Sequencing the complete exon 2 of the *GDF9* gene in a total of 28 sheep of the Ashgar, Dubasi and Watish ecotypes revealed four polymorphic positions: c.471C>T, c.477G>A, c.721G>A and c.978A>G. Minor allele frequencies for T at position 471 and G at position 978 were very low over all sheep (0.05 and 0.02, respectively). These two SNPs were monomorphic in Dubasi (c.471C>T) and Dubasi and Watish (c.978A>G) sheep, respectively. The two other SNPs in exon 2 were polymorphic in all three ecotypes. For the SNP c.477A>G, the A allele was predominant in Ashgar (0.44), whereas the G allele was the predominant allele in Dubasi and Watish (0.60 and 0.69, respectively). Genotype frequencies for all polymorphic exon 2 SNPs for all sheep, for the different ecotypes and for ewes with a single lamb and for ewes with an average of more than a single lamb are given in Table 2. No significant differences in allele or genotype frequencies between the two groups differing in litter size were observed for any of these SNPs.

Table 2 Genotype frequencies of *GDF9* exon 2 SNPs in Sudanese desert sheep, ecotypes, and in ewes with single and more than single lambs

Sheep group	Sheep (n)	Genotypes of SNPs at positions											
		c.471C>T			c.477G>A			c.721G>A			c.978A>G		
		CC	CT	TT	GG	AG	AA	GG	AG	AA	AA	AG	GG
Ashgar	10	0.80	0.20	0.00	0.10	0.60	0.30	0.70	0.20	0.10	0.90	0.10	0.00
Dubasi	10	1.00	0.00	0.00	0.40	0.40	0.20	0.80	0.10	0.10	1.00	0.00	0.00
Watish	8	0.88	0.12	0.00	0.50	0.38	0.12	0.88	0.12	0.00	1.00	0.00	0.00
All sheep	28	0.89	0.11	0.00	0.33	0.46	0.21	0.79	0.14	0.07	0.96	0.04	0.00
Lambing type													
Single	15	0.93	0.07	0.00	0.33	0.40	0.27	0.80	0.13	0.07	1.00	0.00	0.00
More than single	13	0.85	0.15	0.00	0.31	0.54	0.15	0.77	0.15	0.08	0.92	0.08	0.00

Incubation of the 357-bp fragment containing the polymorphic position c.260G>A in *GDF9* exon 1 with *HhaI* restriction enzyme resulted in fragments of 222 and 135 bp for genotype GG and in fragments of 357, 222 and 135 bp for genotype AG (Figure 2).

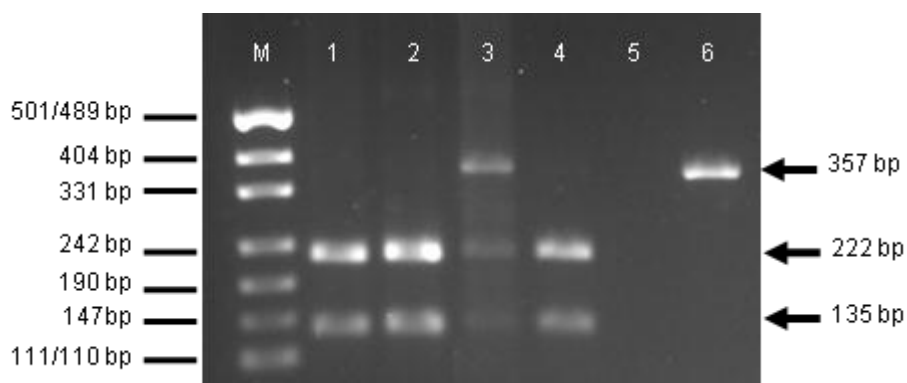


Figure 2 Determination of *GDF9* genotypes at position c.260G>A by RFLP analysis

M = DNA size marker pUC19 DNA/MspI (Thermo Fisher Scientific, Waltham, USA); 1–4 = PCR products digested with *HhaI* (1, 2, 4 = genotype GG, 3 = genotype AG); 5 = negative control; 6 = undigested PCR product

For any sample, only a single 357-bp fragment (as expected for genotype AA) was observed after digestion with *HhaI*.

Allele and genotype frequencies calculated for the c.260G>A variant are given in Table 3. The frequency of the A allele was 0.10 among all genotyped sheep. In Ashgar, it was higher than in Dubasi and Watish (0.19 compared with 0.03 and 0.06, respectively), but similar to these breeds, no sheep with the AA genotype was identified among the Ashgar sheep. Comparison of allele and genotype frequencies between ewes with a single lamb and with more lambs (see Table 3) revealed no significant differences.

Table 3 Allele and genotype frequencies of *GDF9* exon 1 SNP c.260G>A in Sudanese desert sheep ecotypes, and in ewes with single/more than single lambs

Sheep group	Sheep (n)	SNP c.260G>A				
		Allele frequency		Genotype frequency		
		A	G	GG	AG	AA
Ashgar	35	0.19	0.81	0.63	0.37	0.00
Dubasi	29	0.03	0.97	0.93	0.07	0.00
Watish	33	0.06	0.94	0.88	0.12	0.00
All sheep	97	0.10	0.90	0.80	0.20	0.00
Lambing type						
Single	54	0.11	0.89	0.78	0.22	0.00
More than single	43	0.08	0.92	0.84	0.16	0.00

In this study, five known *GDF9* variants (c.471C>T, c.477G>A, c.721G>A and c.978A>G in exon 2, and c.260G>A in exon 1) were found to be polymorphic in at least one of the three Sudanese desert sheep ecotypes Ashgar, Dubasi and Watish. Only the SNP c.721G>A causes an amino acid substitution (p.Glu241Lys), which owing to the change of an acidic group with a basic group is a non-conservative one. However, as for all of the identified SNPs, this variant is located proximal to the furin protease cleavage site. Therefore, it was not unexpected that no significant association could be observed between the identified variants in exon 2 of *GDF9* and litter size in Sudanese desert sheep ecotypes. Although the number of 28 sheep is very low for association testing, the authors refrained from genotyping the four polymorphic SNPs in exon 2 of *GDF9* in a higher number of sheep because literature relating these variants to ovulation rate could not be found.

Also for the SNP c.260G>A in exon 1 of *GDF9*, no significant association was found with litter size in the sampled sheep. However, a higher frequency of the minor A allele was observed in Ashgar compared with Dubasi and Watish. From the 36 Ashgar sheep that were genotyped, 21 had single lambs and 15 had more than a single lamb on average. Because Ashgar sheep were observed to have a higher litter size than Dubasi and Watish (Suliaman *et al.*, 1990), it may be interesting to genotype more Ashgar sheep for this SNP and to test for association with litter size within this breed. Results from Barzegari *et al.* (2010) indicate a possible effect of this SNP on ovulation rate/litter size. However, these results are based on very few sheep and should be taken with great care.

In addition to *GDF9*, the presence of other known major genes influencing ovulation rate could be tested for the desert sheep ecotypes analysed in this study. On the other hand, because these sheep do not show extraordinarily high litter sizes (compared with certain other breeds), the chance to identify such major gene variants seems to be low. By genotyping *BMP1B*, *BMP15* and *GDF9* variants in five Tunisian sheep breeds (Barbarine, Queue Fine de L'Ouest, Noire de Thibar, Sicilo-Sarde and D'man) with litter sizes ranging

from 1.14 (Queue Fine de L'Ouest) to 2.72 (D'man), Vacca *et al.* (2010) found an absence of all known ovulation-influencing alleles in these breeds. However, other breeds and genes are still open for research. A major gene variant that increases litter size in such a native African breed could be introduced in desert sheep ecotypes by classical intercrossing and backcrossing, and carriers of such a variant could then be identified and selected easily by genetic testing. Increased numbers of lambs have the potential to improve the livelihood and social status of the nomads. Of course, natural feed resources are limited in this region. However, it is more favourable to raise a larger number of lambs only once a year in the rain period than a lower number twice a year.

Conclusion

In summary, the authors concluded that five positions in *GDF9* were found to be polymorphic in at least one of the Sudanese desert sheep ecotypes Ashgar, Dubasi and Watish. Except for the exon 1 variant c.260G>A, which had a higher frequency of the A allele in the more prolific Ashgar sheep compared with the less prolific Dubasi and Watish sheep, significant associations of these *GDF9* variants with litter size were not observed. This suggests that the *GDF9* variants that influence ovulation are absent in these Sudanese sheep ecotypes, and therefore cannot be used to increase litter size within this population of sheep.

Acknowledgments

The visit of A.S. Ali to the Department of Animal Breeding and Genetics at the Justus Liebig University of Giessen in Germany was financed by the German Academic Exchange Service (DAAD).

Authors' Contributions

A.S. Ali collected samples, executed sequencing and genotyping methods, analysed genotyping data and wrote significant parts of the manuscript. M.T. Ibrahim contributed to overall experiment design and collection of samples. M.M. Mohammed supervised and contributed to preparation of DNA samples. A.A. Elobied contributed to collection of samples and DNA extraction. G. Lühken supervised the establishment of sequencing and genotyping methods, contributed to data analysis and wrote significant parts of the manuscript. The manuscript has been read and approved by all named authors.

Conflict of Interest Declaration

The authors declare no conflicts of interest.

References

- Albarella, S., Ciotola, F., Selvaggi, M., Dario, C., Licciardi, S., Scopino, G., Frate, D. & Peretti, V., 2015. Analysis of major fecundity genes in autochthonous Laticauda and Bagnolese sheep breeds. *Small Ruminant Res.* 133, 118–122.
- Barzegari, A., Atashpaz S., Ghabili K., Nemati Z., Rustaei M. & Azarbaijani, R., 2010. Polymorphisms in *GDF9* and *BMP15* associated with fertility and ovulation rate in Moghani and Ghezel sheep in Iran. *Reprod. Domest. Anim.* 45, 666–669.
- Bodensteiner, K.J., Clay, C.M., Moeller C.L. & Sawyer, H.R., 1999. Molecular cloning of the ovine growth/differentiation factor-9 gene and expression of growth/differentiation factor-9 in ovine and bovine ovaries. *Biol. Reprod.* 60, 381–386.
- Chu, M.X., Cheng, R.H., Fang, L. & Ye, S.C., 2011. *GDF9* as a candidate gene for prolificacy of Small Tail Han sheep. *Mol. Biol Rep.*, 38, 5199–5204.
- El-Hag, F.M., Fadlalla, B. & Mukhtar, H.K., 2001. Some production characteristics of Sudan desert sheep under range conditions in North Kordofan, Sudan. *Trop. Anim. Health Pro.* 33, 229–239.
- Elvin, J.A., Clark, A.T., Wang, P., Wolfman, N.M. & Matzuk, M.M., 1999. Paracrine actions of growth differentiation factor-9 in the mammalian ovary. *Mol. Endocrinol.* 13, 1035–1048.
- Hanrahan, P.J., Gregan, S.M., Mulsant, P., Mullen, M., Davis, G.H., Powell, R. & Galloway, S.M., 2004. Mutations in the genes for oocyte-derived growth factors *GDF9* and *BMP15* are associated with both increased ovulation rate and sterility in Cambridge and Belclare sheep (*Ovis aries*). *Biol. Reprod.* 70, 900–909.
- Juengel, J.L., Davis, G.H. & McNatty K.P., 2013. Using sheep lines with mutations in single genes to better understand ovarian function. *Reproduction* 146, R111–R123.
- MARFR, 2013. Ministry of Animal Resources, Fisheries and Ranges. Department of Statistic and Information. Khartoum, Sudan. Statistical Bulletin for Animal Resources, Issue No. 20, 13–16.
- McNatty, K.P., Juengel J.L., Reader, K.L., Lun, S., Myllymaa, S., Lawrence, S.B., Western, A., Meerasahib, M.F., Mottershead, D.G., Groome, N.P., Ritvos, O. & Laitinen, M.P., 2005. Bone morphogenetic

- protein 15 and growth differentiation factor 9 co-operate to regulate granulosa cell function in ruminants. *Reproduction* 129, 481–487.
- Montgomery, G.W. & Sise, J.A., 1990. Extraction of DNA from sheep white blood cells, *New Zeal. J. Agr. Res.* 33, 3, 437–441.
- Mukhtar, H.K., 1985. Constraints to desert sheep production in the sedentary and nomadic systems of North Kordofan. In: *Annual Research Report (1984–85)*, (El-Obeid Research Station, Agricultural Research Corporation (ARC), Wad Medani, Sudan). Ed: Lazim M.E., pp. 40–55.
- Nicol, L., Bishop, S.C., Pong-Wong R., Bendixen, C., Holm L.E., Rhind, S.M. & McNeilly, A.S., 2009. Homozygosity for a single base-pair mutation in the oocyte-specific GDF9 gene results in sterility in Thoka sheep. *Reproduction* 138, 921–933.
- Paulini, F., & Melo, E.O., 2011. The role of oocyte–secreted factors GDF9 and BMP15 in follicular development and oogenesis. *Reprod. Domest. Anim.* 46, 354–361.
- Silva, B.D., Castro, E.A., Souza, C.J., Paiva, S.R., Sartori, R., Franco, M.M., Azevedo, H.C., Silva, T.A., Vieira, A.M., Neves, J.P. & Melo, E.O., 2011. A new polymorphism in the growth and differentiation factor 9 (GDF9) gene is associated with increased ovulation rate and prolificacy in homozygous sheep. *Anim. Genet.* 42, 89–92.
- Souza, C.J.H., McNeilly, A.S., Benavides, M.V., Melo, E.O. & Moraes J.C.F., 2014. Mutation in the protease cleavage site of GDF9 increases ovulation rate and litter size in heterozygous ewes and causes infertility in homozygous ewes. *Anim. Genet.* 45, 732–739.
- Suliman, A.H., Sayers, A.R. & Wilson, R.T., 1990. Evaluation of Shugar, Dubasi and Watish subecotypes of Sudan desert sheep at El Huda National Sheep Research station, Gezira Province, Sudan. ILCA Research report, No. 18, Addis Ababa Ethiopia, pp. 30.
- Untergrasser, A., Cutcutache, I., Koressaar, T., Ye, J., Faircloth, B.C., Remm, M. & Rozen, S.G., 2012. Primer3 – new capabilities and interfaces. *Nucl. Acids Res.* 40, e115.
- Vacca, G.M., Dhaouadia, A., Rekikb, M., Carcangiua, V., Pazzolaa, M. & Dettoria, M.L., 2010. Prolificacy genotypes at BMP1B, BMP15 and GDF9 genes in North African sheep breeds. *Small Ruminant Res.* 88, 67–71.
- Vage, D.I., Husdal, M., Kent, M.P., Klemetsdal, G. & Boman, I.A., 2013. A missense mutation in growth differentiation factor 9 (GDF9) is strongly associated with litter size in sheep. *BMC Genetics* 14, 1.