

Effects of *post mortem* temperature on rigor tension, shortening and pH in ostrich muscle

S.StC. Botha^{1,2}, L.C. Hoffman^{1#} and T.J. Britz²

¹ Department of Animal Sciences,

² Department of Food Science, University of Stellenbosch, P. Bag X1, Matieland 7602, South Africa

Abstract

Fully developed *rigor mortis* in muscle is characterised by maximum loss of extensibility. The course of *post mortem* changes in ostrich muscle was studied by following isometric tension, shortening and change in pH during the first 24 h *post mortem* within muscle strips from the *muscularis gastrocnemius, pars interna* at constant temperatures of 7 °C and 37 °C. Maximum shortening was significantly higher at 37 °C (33.4 ± 3.57%) than at 7 °C (10.7 ± 2.63%). The rate of rigor development was temperature dependent, reaching a maximum tension at 4.08 ± 3.89 h *post mortem* in muscle strips at 37 °C; while at 7 °C maximum tension was reached at 10.5 ± 6.47 h *post mortem*. It was concluded that the completion of rigor occurred at the point of minimum pH.

Keywords: *Rigor mortis*, rigor tension, muscle shortening, ostrich, rigor shortening, cold-shortening, hot-deboning, rate of pH decline

Corresponding author. E-mail: lch@sun.ac.za

Introduction

During the development of *rigor mortis* muscles become inextensible due to the sum of each muscle fibre going into full rigor, with irreversible cross-bridge formation (actomyosin) of the contractile components, actin and myosin (Hwang *et al.*, 2003). Bendall (1973) defined the onset of *rigor mortis* as the beginning of the decrease in extensibility of the muscle, while completion of rigor is seen as the maximum loss of extensibility (Honikel *et al.*, 1983). The rigor process involves first a delay period, when the level of ATP (adenosine triphosphate) is constant, CP (creatine phosphate) is decreasing rapidly, and there is a slow production of lactate and no onset of rigor development. This is followed by a rapid phase when CP is low enough to initiate a rapid decline in ATP concentration, which is accompanied by a decreasing extensibility of the muscle due to irreversible cross-bridge formation of actin and myosin (Bendall, 1973). Shortening of muscles occurs before the onset of *rigor mortis* since contraction requires a minimum ATP concentration and an increased level of Ca²⁺ ions around the myofibrils (Honikel *et al.*, 1983).

Cold-shortening occurs when muscles are exposed to low temperatures (below 10 °C to 15 °C) early *post mortem*, when ATP and pH levels are still high (pH above 6.2) (Nuss & Wolfe, 1980-81; Lawrie, 1998). On the other hand, rigor tension occurs much later and at any temperature between 0 °C and 37 °C, reaching maximum values when ATP levels have been depleted and pH is at a minimum value (Nuss & Wolfe, 1980-81). The stiffness characteristic of *rigor mortis* is then maintained by continuous tension exerted by the cross-bridges between myosin and actin filaments (Pearson & Young, 1989).

In both pork and beef muscles shortening is a major determinant of tenderness when the tissue has not yet entered *rigor mortis* and when the rate of muscle pH decline is rapid (Møller & Vestergaard, 1987; Smulders *et al.*, 1990). Tornberg (1996) has reported that a more shortened beef muscle has a higher cooking loss and a higher number of fibres per unit cross-area, leading to higher Warner-Bratzler peak shear force values and greater toughness. Marsh & Leet (1966) and Locker & Daines (1975) also reported shortened beef muscles to be tougher, while Powell (1978) and Honikel *et al.* (1980) reported higher drip losses in contracted muscles than in non-shortened muscles.

Knowledge of the course of *rigor mortis* and *post mortem* pH changes, as well as the influence of temperature, on the onset of *rigor mortis*, might help to reduce purge and initial toughness in hot-deboned ostrich muscles. The aim of this study was therefore to investigate the development of isometric tension (developing tension while the muscle is prevented from contracting) and muscle shortening in ostrich *m. gastrocnemius, pars interna* during *rigor mortis* at 7 °C and at 37 °C in an attempt to determine the time

course of rigor, pH decline, and the degree and extent of shortening (occurrence of cold-shortening and/or rigor shortening).

Materials and Methods

Ten rested (12 h of lairage) ostriches (*Struthio camelus var. domesticus*) from different farms were slaughtered at ages of between 10 and 14 months, as described by Wotton & Sparrey (2002), during the summer months of 2004 (February to April) at an EU approved ostrich abattoir in Malmesbury, South Africa. The right leg *m. gastrocnemius, pars interna* was removed and mounted within 20 min to 1 h after stunning and exsanguination. Two muscle strips (dimensions: 10 x 10 x 30 mm; mass: 1.5 to 3 g) were carefully cut, parallel to the fibre axis, from the 10 individual muscles. The cross-sectional area of each muscle strip was calculated with the use of Rigotech[®] software (Rigotech version 3.0, ©ReoLogica, 1999, Sweden) by using the length and the weight of the strip, together with the density of the muscle (1.06 g/cm³). Muscle strips were glued onto the aluminium discs of two separate rigometers (Rigotech[®], ©ReoLogica, 1999, Sweden) at constant temperatures of 7 °C and 37 °C, respectively, with the use of cyanoacrylate glue (Lynlim Superglue, Norway). To provide an anaerobic environment and to minimise dehydration, the strips were covered with a mixture of paraffin oil and petroleum jelly. Rigor tension and muscle shortening were recorded during *rigor mortis* every 15 min for the first 23 to 24 h *post mortem*. Rigor tension was expressed as force per unit area (mN/mm²) and shortening was expressed as the percentage decrease in length relative to the initial length of the muscle strip.

Muscle pH and temperature were measured by using SenTix 41 probes (Germany), connected to portable pH meters 340i (WTW, GmbH & Co. KG, Weilheim, Germany). The probes were placed directly into larger portions of the same muscles used for the measurement of rigor tension and muscle shortening. These portions were also covered with paraffin oil and petroleum jelly and placed in the rigometers at 7 and 37 °C, respectively. Muscle pH and temperature were continuously measured every 10 min for the first 23 to 24 h *post mortem*.

The changes in pH and temperature (°C) were also measured continuously, every 10 min for the first 23 to 24 h, in the intact left leg *m. gastrocnemius, pars interna* from the same ostrich carcasses by inserting the probes to a depth of 4 to 5 cm into the intact muscles at 1 h *post mortem*. At approximately 2 h *post mortem* these carcasses were moved to the refrigerator (<4 °C) at the abattoir for 24 h.

The experimental design was a complete randomised block design, replicated in 10 blocks (ostrich carcasses), with two treatments for the rigometer results (muscle strips at 7 and 37 °C, respectively) and with three treatments for the pH and temperature results (intact muscle, and muscle strips at 7 and 37 °C, respectively). The data was subjected to factorial analysis of variance (ANOVA) using SAS version 8.2 Statistical Software (SAS, 1999). Shapiro-Wilk tests were performed for testing non-normality (Shapiro & Wilk, 1965).

Results and Discussion

The analysis of variance (ANOVA) of the dependant variables: maximum tension (mN.mm⁻²), pH at maximum tension, time (h) *post mortem* to reach maximum tension, maximum shortening (%), pH at maximum shortening, and the time (h) *post mortem* it took for the muscle strips to reach maximum shortening is presented in Table 1. Only the data for the time (h) to reach maximum shortening was not normally distributed (P <0.01) due to the occurrence of kurtosis. As the latter does not have a significant effect on the normality of the data (Glass *et al.*, 1972), the data were analysed without transformation thereof. Temperature had no effect (P >0.05) on maximum tension pH or time to reach maximum tension (Table 2). Nuss & Wolfe (1980-81) also indicated that the time to reach maximum tension in beef muscles decreased with an increase in temperature from 5 °C to 37 °C. Devine *et al.* (1999) demonstrated that the time required for bovine muscle to reach maximum tension was negatively correlated with temperature.

Similar to results found by Hertzman *et al.* (1993) for beef, maximum shortening for the ostrich muscle strips were significantly higher at 37 °C than at 7 °C (Table 2), while the pH values at the time of maximum shortening did not differ significantly (P >0.05) between 7 °C and 37 °C. This indicated that the muscle strips from the same individual ostrich carcass were, as expected, at the same initial energy levels (lactate and ATP concentrations) early *post mortem*, when the muscle strips were placed in the rigometers, leading therefore to similar pH values at the point of maximum tension development. The difference (P <0.01) in the time taken to reach maximum shortening for the main effect, namely 'temperature',

indicated that the maximum shortening was reached sooner in the muscle strips at 37 °C (5.6 ± 3.65 h) than in the muscle strips at 7 °C (23.0 ± 0.45 h). This also indicated that the rate of muscle shortening was therefore faster at 37 °C than at 7 °C.

Table 1. Mean values (\pm standard deviation) for maximum tension ($\text{mN}\cdot\text{mm}^{-2}$), pH at maximum tension, time (h) *post mortem* to reach maximum tension, maximum shortening (%), pH at maximum shortening and the time (h) *post mortem* to reach maximum shortening at 7 °C and 37 °C, respectively

Parameter	Temperature	
	7 °C	37 °C
Maximum tension (mN/mm^2)	$347.5^a \pm 41.53$	$284.8^a \pm 82.44$
pH at maximum tension	$6.1^a \pm 0.30$	$5.9^a \pm 0.13$
Time (h) <i>post mortem</i> to reach maximum tension	$10.5^a \pm 6.47$	$4.1^a \pm 3.89$
Maximum shortening (%)	$10.7^a \pm 2.63$	$33.4^b \pm 3.57$
pH at maximum shortening	$6.1^a \pm 0.25$	$6.1^a \pm 0.37$
Time (h) <i>post mortem</i> to reach maximum shortening	$23.0^a \pm 0.45$ h	$5.6^b \pm 3.65$

^{ab} Different superscripts within rows differ at $P < 0.05$.

It has been hypothesised that the occurrence of higher maximum tension and less shortening at 7 °C, compared to at 37 °C, could be due to the release of calcium ions (Ca^{2+}) from the sarcotubular system at low temperatures (Cornforth *et al.*, 1980; Whiting, 1980). Cornforth *et al.* (1980) and Whiting (1980) reported that the amount of Ca^{2+} released from bovine sarcoplasmic reticulum was least at pH 6.6, when the temperature declined from 38 °C to 0 °C. However, as the pH declined, the sarcoplasmic reticulum released increasingly more Ca^{2+} (Kanda *et al.*, 1977), with essentially no activity near pH 5.0 (Whiting 1980). Both cold temperatures and low pH values lead to a decrease in the ability to accumulate Ca^{2+} and to an increase in the release of Ca^{2+} from the sarcoplasmic reticulum. Whiting (1980) found that the maximum Ca^{2+} uptake for bovine mitochondria was at pH 7.2. This activity decreased rapidly at a pH of 6.5 and was very low at a pH value of 5.5. Whiting (1980) thus concluded that the mitochondria would be the first to lose their ability to sequester Ca^{2+} as the *post mortem* pH declines from 6.5 to 6.0. Temperatures below 15 °C also stimulate the release of Ca^{2+} ions from the sarcotubular system, enhancing the contractile actomyosin ATP-ase (Lawrie, 1998). With the increase in Ca^{2+} concentrations actin and myosin interconnect and, under normal circumstances, when there is enough ATP, the contractile ATP-ase provides the energy for the actin filament to be pulled inwards towards the centre of the sarcomere, causing the sarcomere to shorten.

For this study it was hypothesised that muscle strips from the same ostrich had similar energy levels when placed into the two separate rigometers at 7 °C and 37 °C, respectively. However, as illustrated by data in Figure 1, the influence of the low temperature of 7 °C slowed the rate of muscle contraction and therefore maximum tension, and maximum shortening was reached sooner in muscle strips at 37 °C than in muscle strips at 7 °C. It may also be hypothesised that the low temperature of 7 °C stimulated the release of Ca^{2+} ions from the sarcotubular system (Whiting, 1980; Hertzman *et al.*, 1993). Simultaneously, the rate of anaerobic glycolysis and therefore the production of ATP and lactic acid was slower, leading to lower levels of ATP than in the muscle strips at 37 °C. This resulted in tension due to the binding between actin and myosin, but very little shortening, since there was not enough energy to cause the sarcomeres to shorten. In the absence of ATP the continuous tension is exerted by the cross-bridges formed between actin and myosin filaments (Pearson & Young, 1989). This result differs from that of Hertzman *et al.* (1993) who noted that shortening occurs before tension. However, their lowest temperature was 15 °C whereas the temperature in this investigation was 7 °C and the possibility of cold shortening and decreased anaerobic glycolysis from the present investigation may explain these differences.

Table 2 Analysis of variance (ANOVA) of the dependant variables: maximum tension (mN/mm²), pH at maximum tension, time (h) *post mortem* to reach maximum tension, maximum shortening (%), pH at maximum shortening and the time (h) *post mortem* to reach maximum shortening with ostrich carcass and temperature (°C) as main effects, as well as the Shapiro-Wilk test for non-normality

Source	df	Max tension (mN/mm ²)		pH at max tension		Time to reach max tension		Max shortening (5%)		pH at max shortening		Time to reach max shortening		
		MS	P	MS	P	MS	P	MS	P	MS	P	df	MS	P
Ostrich carcass	5	4877.631	0.3783	0.061	0.3501	15.471	0.8490	9.384	0.5398	0.153	0.1059	5	18.458	0.2096
Temperature	1	11775.068	0.1321	0.072	0.2475	123.521	0.1454	1546.780	<0.0001	0.004	0.7539	1	757.422	0.0006
Error	5	3642.686		0.039		41.571		10.309		0.039		4	7.726	
Shapiro-Wilk			0.9941		0.9163		0.9032		0.8620		0.6608			0.9110

df – Degree of freedom
 MS – Mean sum of squares
 P – Probability value of F-ratio test
 Max - maximum

In this study the higher maximum shortening (%) at the higher temperature of 37 °C could, in turn, be explained by the ATP levels (Hertzman *et al.*, 1993) in conjunction with the influence of the high temperature on the sarcotubular system (Whiting, 1980). Hertzman *et al.* (1993) found a strong correlation between maximum shortening and ATP levels at the onset of the rapid shortening phase. These researchers explained the higher shortening at 37 °C compared to 15 °C to be due to the higher ATP level at the higher temperature. Whiting (1980) showed that the Ca²⁺ sequestering ability of the mitochondria (from beef muscles) at pH 7.2 decreased rapidly at temperatures higher than 20 °C and was practically non-existent after 30 min at 37 °C. On the other hand, the Ca²⁺ sequestering activity of the sarcoplasmic reticulum remained relatively constant at temperatures up to 37 °C at pH 7.2, whereafter its ability to sequester Ca²⁺ decreased rapidly at temperatures higher than 37 °C. Therefore, for this study, it was hypothesised that the reduced stability of the Ca²⁺ uptake ability of especially mitochondria, together with a faster decline in pH at 37 °C compared to 7 °C (see Figure 2), could initiate rigor shortening at higher ATP levels, resulting in the higher maximum shortening at 37 °C.

Isometric tension results for beef muscles, obtained by Hertzman *et al.* (1993), were characterised by a delay phase preceding the development of tension, and this delay was shorter at 37 °C than at 15 °C. In the case of shortening, these authors reported a delay period at 15 °C, while at 37 °C shortening started immediately. From the data in Figure 1 it can be seen that, similar to the findings of Hertzman *et al.* (1993), there was a short delay period at both 7 °C and 37 °C, before the onset of tension development, with the delay period being shorter at 37 °C than at 7 °C. The development of shortening started almost immediately in muscle strips at 37 °C; while at 7 °C there was a delay of approximately 100 min before the onset of shortening.

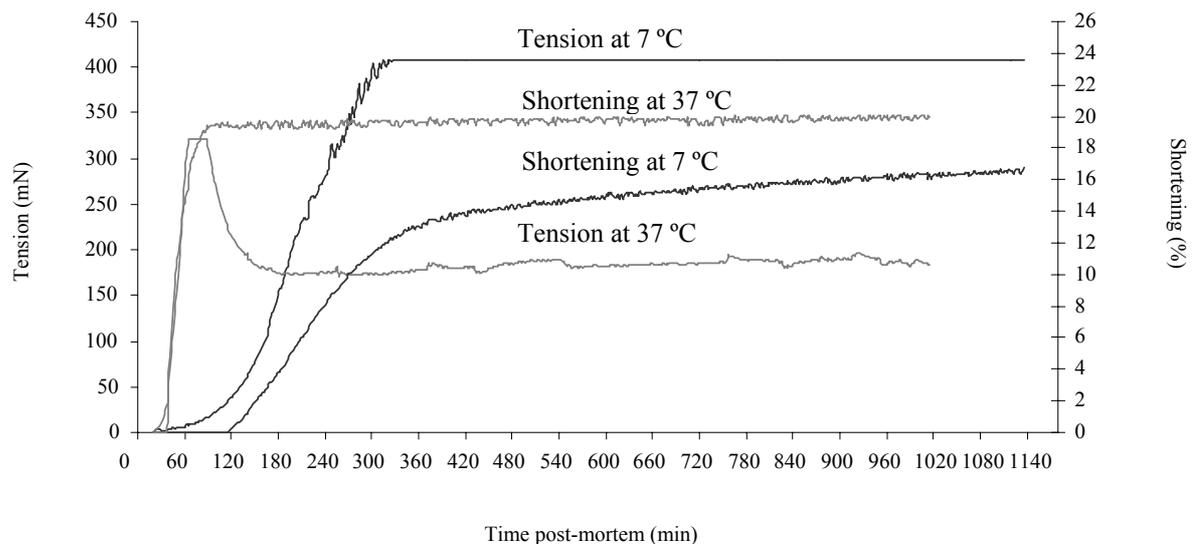


Figure 1 Development of muscle rigor tension and shortening from the time (20 min *post mortem*) the muscle strips from an individual ostrich were placed in the rigometers, maintained at 7 °C and 37 °C, respectively.

What is also noteworthy is that the tension peak for the muscle strips at 37 °C was followed by a decrease in tension, while at 7 °C the maximum tension remained constant. The constant tension at 7 °C, as depicted in Figure 1, was initially attributed to a possible, unexplained technical failure of the rigometer. However, the tension data obtained from the muscle strips from the other ostriches at 7 °C did in fact follow the same pattern - a constant maximum tension (measured up to 22 h *post mortem* in the rigometer). The decrease in tension at 37 °C could be ascribed to proteolysis or the initial ageing process of the muscle after fully developed rigor at high temperatures. The rate of tenderisation early *post mortem* would be enhanced in muscles that are at higher temperatures since the extent of proteolysis is temperature dependent; it is greater at 37 °C than at 5 °C. This phenomenon was also shown in beef by Hertzman (1993), who ascribed

the fall in tension to proteolysis. This was studied further by Tornberg *et al.* (2000) who studied the effect of pre-rigor conditions in beef under various temperature and pH regimens and also noted and postulated that this decrease is linked closely to proteolysis. What is noteworthy and warrants further research, is the very rapid occurrence of this phenomenon in ostrich muscle.

It was hypothesised that since the low temperature of 7 °C slowed the rate of proteolysis, the muscle strips at 7 °C would eventually also have shown this decrease in tension, but some hours later than the muscle strips at 37 °C.

The analysis of variance (ANOVA) of the dependable variables minimum pH and the time (h) *post mortem* at which the minimum pH was reached are presented in Table 3. Temperature had an effect on the rate of pH decline; the rate of decline was faster in muscle strips at 37 °C than in muscle strips at 7 °C (Figure. 2). The intact muscles reached a minimum pH value (5.6 ± 0.22) within 2 h after slaughter (Table 4), while muscle strips at 7 °C reached a minimum pH of 5.9 ± 0.21 at 6.4 ± 4.51 h (between 300 and 600 min) *post mortem* ($P < 0.05$). From these results it was hypothesised that *post mortem* glycolysis and anaerobic production of ATP were faster in muscles at higher muscle temperatures during the rigor process, which in turn could explain the higher percentage of maximum shortening obtained at 37 °C ($33.4 \pm 3.57\%$) compared to 7 °C ($10.7 \pm 2.63\%$), as described earlier.

Table 3 Analysis of variance (ANOVA) of the dependable variables: minimum pH and the time (h) *post mortem* at which the minimum pH was reached with ostrich and deboning [intact *m. gastrocnemius, pars interna* refrigerated for 24 h at 4 °C, excised (20 min to 1 h *post mortem*) *m. gastrocnemius, pars interna* from which muscle strips were kept at respectively 7 °C and 37 °C in two separate rigometers] as main effects, as well as the Shapiro-Wilk test for non-normality

Source	df	Minimum pH		Time (h) of minimum pH	
		MS	P	MS	P
Ostrich carcass	5	0.047	0.2812	17.530	0.1547
Deboning	2	0.504	0.2512	33.160	0.0569
Error	9	0.031		8.275	
Shapiro-Wilk			0.2353		0.0820

df – Degree of freedom

MS – Mean sum of squares

P – Probability value of F-ratio test

Table 4 Mean (\pm standard deviation) minimum pH and time (h) *post mortem* at which the minimum pH values were reached, respectively, within cold-deboned (excised at 24 h *post mortem*) muscles, muscle strips from hot-deboned (20 min to 1 h *post mortem*) muscles at 7 °C, and muscle strips from hot-deboned (20 min to 1 h *post mortem*) muscles at 37 °C

Treatment	Minimum pH	Time (h) to reach minimum pH	Temperature (°C) at minimum pH
Intact muscle	5.85 ± 0.22	2.00 ± 0.00	32.08 ± 4.29
Muscle strips at 7 °C	5.94 ± 0.21	6.42 ± 4.51	7.00 ± 0.50
Muscle strips at 37 °C	5.76 ± 0.13	4.83 ± 3.82	37.00 ± 0.06

Figure 2 illustrates the changes in pH and the decline in muscle temperature for the intact muscle and muscle strips (at 7 °C and at 37 °C) from a single ostrich carcass (representative of the profiles of the other ostriches). As illustrated by the pH and temperature data, there was an initial decrease in pH of the muscle

strip at 37 °C, followed by an increase in pH after a minimum value had been reached. These pH changes were similar to changes in the intact muscle at <4 °C. In general, the pH for intact muscle of *m. gastrocnemius, pars interna* decreased from 1 h *post mortem* to a mean minimum value of 5.85 ± 0.22 at 2 h *post mortem*, indicating a rapid decrease in pH during the first 2 - 3 h *post mortem*. After reaching a minimum value, the pH then increased to a mean value of 6.14 ± 0.19 at 24 h *post mortem*. Pearson & Young (1989) have reported that for chicken, *post mortem* muscle glycogen levels decline to extremely low levels at approximately 2 - 4.5 h *post mortem*. This indicated that the muscles were in full rigor and had reached pH values near those found at 24 h *post mortem* in beef muscles.

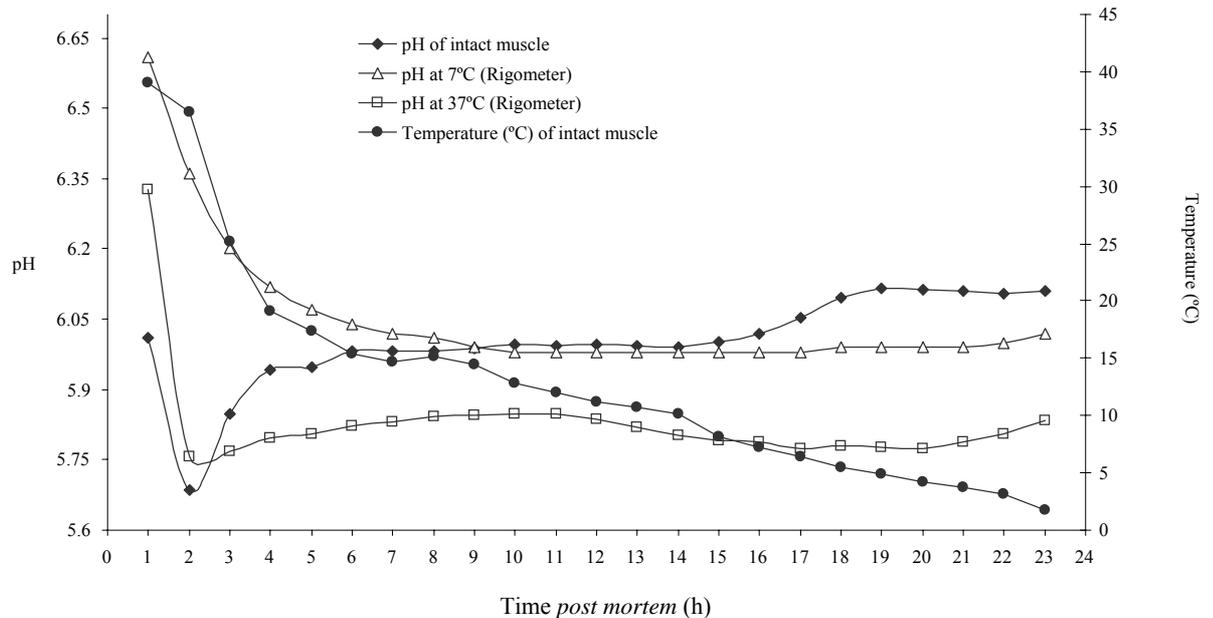


Figure 2 Hourly *post mortem* temperature (°C) and pH values indicating the decline in temperature (●) and pH change (◆) during the first 23 h *post mortem* for intact *m. gastrocnemius, pars interna* refrigerated at <4 °C, as well as for muscle strips from the *m. gastrocnemius, pars interna*, from the same ostrich carcass, maintained at 7 °C (△) and 37 °C (□) in the rigometers.

The intact muscles had a mean muscle temperature of 32.1 ± 4.29 °C at the time when the minimum pH was reached. It was also observed that the minimum pH values in both the intact muscles and the muscle strips at 37 °C were reached at approximately the same time ($P > 0.05$) *post mortem* (Table 4). The mean temperature of the muscles left intact at 1 h *post mortem* was 42.1 ± 7.58 °C, and, since muscle strips were cut from the right muscles from the same ostrich carcasses and placed in the rigometers at 1 h *post mortem*, it was assumed that the mean initial temperature (1 h *post mortem*) of the muscle strips was >35 °C. Thus, muscle strips at 37 °C had a similar fast rate of pH decline during the first 2 h *post mortem* to the intact muscles. Although the muscle strips at 7 °C also had a mean initial temperature of >35 °C, their surface area to volume ratio would have resulted in the muscle strips exhibiting a rapid decline in temperature at 7 °C, causing the rate of pH decrease to be slowed. Therefore, muscle strips at 7 °C reached a minimum pH at 6.4 ± 4.51 h *post mortem*. This clearly indicates that the change in *post mortem* pH is highly temperature dependent.

The high initial (1 h *post mortem*) mean muscle temperature of 42.1 ± 7.58 °C can be explained by outliers, as there were two individual ostrich carcasses in this study with muscle temperatures of 54.7 °C and 43.9 °C, respectively. When these outliers were removed from the data, the mean muscle temperature at 1 h *post mortem* was 37.8 ± 0.98 °C. Blight & Hartley (1965) reported the live body temperature of ostriches to be 38.7 °C. It is hypothesised that the high muscle temperature of these two individual ostrich carcasses at 1 h *post mortem* resulted from the kicking action of the legs while the legs were fixed in the leg clamp during

the stunning procedure (Wotton & Sparray, 2002). The high ambient temperatures in the plucking and skinning halls would also result in the muscle temperature remaining high.

Conclusions

From the results of this study it is concluded that the rate of rigor development, the course of *rigor mortis*, as well as the rate of change in pH in ostrich *m. gastrocnemius, pars interna*, are temperature dependent. These results are in accordance with the results of Hertzman *et al.* (1993). The degree of maximum tension and shortening was also temperature dependent. Maximum tension was higher at 7 °C than at 37 °C, while, in contrast, maximum shortening was significantly higher at 37 °C than at 7 °C. Muscle strips at 37 °C reached a maximum tension value (completion of rigor) within 4.1 ± 3.89 h *post mortem*, while the minimum pH was also reached within 4.8 ± 3.82 h *post mortem* in muscle strips at 37 °C, indicating that full *rigor mortis* in ostrich muscles occurred at the point of minimum pH. The intact muscles reached a minimum pH at approximately 2 h *post mortem*, while muscle temperature was still relatively high (32.1 ± 4.29 °C). It is therefore suggested that ostrich muscles not be hot-deboned within the first 2 h *post mortem*.

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