

# Contamination and disinfection of silicone pacifiers: an *in vitro* study

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## ABSTRACT

**Introduction:** Whilst in use as comforting devices, the nipples of pacifiers are permanently in contact with normal oral flora and saliva, allowing flourishing bacterial biofilms. Effective disinfection will limit contamination, promote oral health and prevent oral infections in children. Studies on pacifier disinfection in South Africa are not well documented.

**Aim:** To investigate *in vitro* disinfection of contaminated pacifiers with alcohol-free oral rinse and microwave.

**Methods:** Seventy two silicone pacifiers were divided into two groups of 36, and contaminated with standardized suspensions of *Candida albicans* and *Streptococcus mutans* respectively. Each group was subdivided into three and disinfected with alcohol-free oral rinse, microwave and sterile distilled water (control), followed by microbiological analysis. Data was analyzed using Kruskal Wallis Anova test.

**Results:** Alcohol-free oral rinse removed *S. mutans* from 42% of pacifiers, whilst microwave removed 33%. Microwave removed *C. albicans* from 83% of pacifiers, but alcohol-free oral rinse removed only 33%, a difference significant at  $p < 0.05$ . All pacifiers treated with sterile water retained bacterial contamination.

**Conclusion:** Microwave was more effective than alcohol-free oral rinse in eliminating *C. albicans* from pacifiers. Microwave and alcohol-free oral rinse were equally effective in eliminating *S. mutans* from pacifiers.

**Key words:** *Candida albicans*, disinfection, microwave, oral rinse, pacifier.

## INTRODUCTION

Pacifiers are commonly used as comforting devices during early childhood.<sup>1</sup> During use, pacifier nipples are permanently in contact with oral normal flora and saliva, allowing a flourishing of bacterial biofilms.<sup>2</sup> These biofilms are important in the formation of plaque, which may lead to dental caries and periodontal disease in children.<sup>3</sup> Several studies have shown that used pacifiers have the ability to retain microorganisms.<sup>1,2</sup> The use of pacifiers has been associated with otitis media,<sup>4</sup> infection by intestinal parasites<sup>5</sup> and yeasts.<sup>1</sup> There is also evidence that these microorganisms can interact with the immune system, leading to allergies,<sup>6</sup> asthma<sup>7</sup> and autoimmune diseases.<sup>8</sup>

However, pacifiers do have their advantages, including soothing infants and affording protection against Sudden Infant Death Syndrome.<sup>9</sup> A recent study showed that microorganisms transferred from mothers sucking pacifiers can reduce the risk of children developing allergies.<sup>10</sup>

Very few studies have investigated disinfection methods for pacifiers. Nelson *et al.*<sup>11</sup> and Chamele *et al.*<sup>12</sup> investigated the effectiveness of alcohol-containing Colgate® PerioGard® Rinse and microwave in removing *S. mutans* from silicone pacifiers. In both studies, PerioGard was found to be as effective as microwave irradiation in eliminating *S. mutans* from pacifiers.<sup>11,12</sup> Da Silva *et al.*<sup>13</sup> showed that microwave was the most effective method when compared with boiling in disinfecting pacifiers.

*S. mutans* is a major cause of dental caries, and is one of the bacteria causing infectious endocarditis in children with congenital heart disease.<sup>14</sup> Early childhood caries (ECC) is one of the most common chronic childhood diseases affecting normal health and well-being. Whilst the overall prevalence of dental caries has reduced worldwide, that of ECC remains high and is currently a WHO concern.<sup>15</sup> In a study investigating the presence of *S. mutans* on latex and silicone pacifiers after use, the bacterium was found to predominate on the silicone pacifiers.<sup>13</sup>

*C. albicans* is a fungus that causes oral thrush in infants and toddlers, and has been reported as a contaminant on pacifiers. In a study of infants up to 8 months of age, the use of a pacifier was found to be highly associated with the frequency of yeast infection, and *C. albicans* was one of the most prevalent species found.<sup>1</sup> Comina *et al.*<sup>2</sup> investigated the microbial contamination of used silicone and latex pacifiers. 80% were found to be predominantly contaminated with *Staphylococcus* and *Candida* species.

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Although there is evidence showing that used pacifiers can retain oral microorganisms<sup>1,2,5</sup> there has been inadequate emphasis worldwide on disinfection methods and their effectiveness in limiting pacifier contamination, including in South Africa. There is therefore a need to investigate disinfection methods for pacifiers to limit their contamination and to promote oral health and prevent oral infections in children. None of the previous studies have tested the efficacy of alcohol-free GUM® Paroex™ Chlorhexidine Gluconate oral rinse in disinfection of pacifiers. The rinse is used to inhibit buildup of plaque thereby reducing gingivitis.

The purpose of the current study was to investigate the *in vitro* disinfection of deliberately contaminated silicone pacifiers using the two methods of alcohol-free GUM® Paroex™ Chlorhexidine Gluconate oral rinse and of microwave.

Significance of study

The knowledge gained from this study would inform the South African community of the importance of regularly disinfecting pacifiers, thus preventing infections such as early childhood caries and oral candidiasis.

MATERIALS AND METHODS

Study population and methodology

The study was conducted at the University of the Witwatersrand, Johannesburg, South Africa. Permission was granted to conduct the study by the Human Research Ethics Committee (Medical) of the University of the

Witwatersrand, Johannesburg (W-CJ-130916-2). Seventy two new silicone pacifiers obtained directly from their packages (Golden Baby, CKT Tek Co. Ltd, New Taipei, Taiwan) were included in the study. Thirty six of these pacifiers were soaked in a standardised inoculum of *C. albicans* (ATCC 90028) at 150x106 cfu/ml for 5 minutes. The remaining 36 pacifiers were soaked in a standardised inoculum of *S. mutants* (NCTC 1044) at 150x106cfu/ml for 5 minutes. The contaminated pacifiers were randomly divided into three groups of 12 each. Group 1 was sprayed three times with GUM® Paroex™ Chlorhexidine Gluconate Oral Rinse (Sunstar Americas, Inc, Ontario, Canada), group 2 was placed in the microwave oven (Sharp SA, Midrand, South Africa) at 750 watts for seven minutes and Group 3 was sprayed three times with sterile distilled water. After disinfection, Group 2 samples were allowed to cool, and Group 1 was rinsed with sterile distilled water for two seconds to remove excess oral rinse.

Microbiological evaluation

After disinfection, all the pacifiers were aseptically suspended in 20ml of sterile phosphate buffered saline (PBS) for two minutes and vortexed for microbial cell detachment. From the initial suspension, dilutions of 10-1, 10-2 and 10-3 were prepared in sterile PBS, and 0.1 ml of each dilution was plated on Mitis salivarius Bacitracin agar (MBA) for the recovery of *S. mutants* and Sabouraud Dextrose agar (SDA) agar for the recovery of *C. albicans*. The SDA plates were incubated at 37°C for 48 hours, and the MBA plates at 37°C (5% CO2) for 48 hours. On conclusion, the numbers of colonies in each plate representing live organisms were counted.

Data analysis

Data were analysed using Kruskal-Wallis ANOVA. Statistical significance was set at the 5% significance level.

RESULTS

Alcohol-free oral rinse removed *S. mutants* from 42% of pacifiers, as compared with microwave which removed *S. mutants* from 33% of pacifiers. Microwave removed *C. albicans* from 83% of pacifiers, as compared with alcohol-free oral rinse, which removed 33%. Sterile water failed to remove either *S. mutants* or *C. albicans* colonies from any of the pacifiers (Figure 1).

Alcohol-free oral rinse and microwave were statistically similar in eliminating *S. mutants* (p>0.05). There was a statistically significant difference between alcohol-free oral

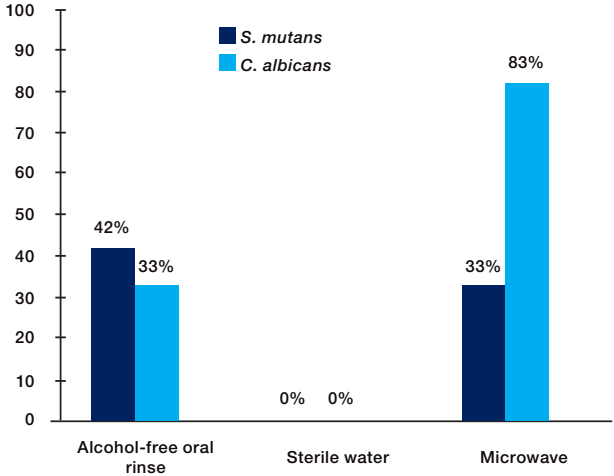


Figure 1: Proportion of pacifiers without contamination after disinfection procedures.

Table 1: Statistical assessment of colony forming units for each microorganism after disinfection with the different methods (Kruskal-Wallis ANOVA)								
Organism tested	Method	N	Mean	Standard deviation	Min	Max	P value	Statistically significant differences
Candida albicans	Sterile distilled water	12	256250.0	25434.85	200000.0	287000.0	0.0001	Sterile distilled water
	Alcohol-free Oral rinse	12	1833.3	2289.63	0.0	7000.0		Alcohol-free Oral rinse
	Microwave	12	3166.7	7505.55	0.0	22000.0		Microwave
Streptococcus mutans	Sterile distilled water	12	250000.0	28556.16	200000.0	287000.0	P<0.05	Alcohol-free Oral rinse
	Alcohol-free Oral rinse	12	1666.7	1874.84	0.0	5000.0		Alcohol-free Oral rinse
	Microwave	12	183333	2289.63	0.0	7000.0		Microwave

rinse and microwave in eliminating *C. albicans* ( $p < 0.05$ ). Statistically significant differences were observed in sterilising efficacy between microwave, alcohol-free oral rinse and sterile water for the removal of both *C. albicans* and *S. mutans* ( $p = 0.0001$ ). (Table 1)

## DISCUSSION

Bacterial biofilms flourish during the continuous contact of pacifier nipples with saliva and oral microorganisms.<sup>2</sup> Disinfection methods for pacifiers have not been adequately addressed.<sup>12</sup> The findings of the current study confirm previous reports that pacifiers can be contaminated by *S. mutans*. There is a need to investigate effective and easy disinfection methods for pacifiers to reduce microbial contamination.

In the current study, alcohol-free oral rinse removed *S. mutans* from 42% of pacifiers, as compared with microwave which showed a success rate of 33% (not significant at  $p > 0.05$ ). This finding differs from those of previous studies, in which it was shown that Periogard containing an alcohol rinse, and microwave irradiation both eliminated *S. mutans* from 100% of pacifiers.<sup>11,12</sup> The presence of alcohol in the Periogard may have increased the disinfecting effect, as alcohol is a known antiseptic.<sup>16</sup> In contrast, a study that investigated the use of mouth rinses with or without alcohol to reduce plaque found both to have similar effects.<sup>17</sup> In a previous study,<sup>13</sup> *S. mutans* has been shown to predominate in silicone pacifiers, and microwave removed this organism from 100% of the pacifiers.<sup>13</sup>

This study revealed that microwave was more effective (83%) than alcohol-free oral rinse (33%) in eliminating *C. albicans* from silicone pacifiers, showing a statistically significant difference ( $p < 0.05$ ) between the disinfecting methods. Da Silva *et al.*<sup>13</sup> found microwave to be as effective as boiling water in disinfecting both latex and silicone pacifiers contaminated with *C. albicans*. Microwave can thus be used for disinfecting silicone pacifiers, as these pacifiers were previously reported to be heat resistant.<sup>12</sup> Microwave has also been found to be effective in the disinfection of other dental materials such as prosthesis<sup>6</sup>, acrylic resins<sup>18</sup> and orthodontic instruments.<sup>19</sup>

*S. mutans* and *C. albicans* colonies remained present in all (100%) pacifiers treated with sterile water, confirming that the use of water to clean pacifiers is not effective in eliminating contamination.<sup>1,2,11</sup> Chamele *et al.* demonstrated that *S. mutans* colonies were still present in 75% of pacifiers treated with sterile water.<sup>12</sup>

*Candida* was found by Comina *et al.* to be one of the most predominant species found in used pacifiers collected from day-care centers.<sup>2</sup> Da Silva *et al.* reported boiling water to be as effective as microwave in removing *C. albicans* from pacifiers.<sup>13</sup>

The limitation of the current study is that it was *in vitro*, carried out on new pacifiers. Different results may be evident if used, worn out pacifiers are assessed in a similar study, for older pacifiers might provide an even more suitable environment for the attachment of microorganisms.

## CONCLUSIONS

Based on the sample size and parameters of this study:

1. Silicone pacifiers can be contaminated *in vitro* with *S. mutans* and *C. albicans*.
2. Microwave was more effective than alcohol-free oral rinse in eliminating *C. albicans* from silicone pacifiers.
3. Microwave and alcohol-free oral rinse were equally effective in eliminating *S. mutans* from silicone pacifiers.

**Conflict of interest:** None declared

## References

1. Mattos-Graner RO, De Moraes AB, Rontani RM, Birman EG. Relation of oral yeast infection in Brazilian infants and use of a pacifier. *J. Dent Child.* 2001; 68: 33–6.
2. Comina E, Marion K, Renaud FN, Dore J, Bergeron E, Freney J. Pacifiers: a microbial reservoir. *Nurs Health Sci.* 2006; 8:216–23.
3. Yonezu T, Yakushiji M. Longitudinal study on influence of prolonged non-nutritive sucking habits on dental caries in Japanese children from 1.5 to 3 years of age. *Bull Tokyo Dent Col* 2008; 49:59–63.
4. Rovers MM, Numans ME, Langenbach E, Grobbee DE, Verheij TJ, Schilder AG. Is pacifier use a risk factor for acute otitis media? A dynamic cohort study. *Fam Pract* 2008; 25:233–6.
5. Pedroso RS, Siqueira RV. A study on protozoan cysts, helminth eggs, and larvae in pacifiers. *J Pediatr* 1997; 73:21–5.
6. Rochow EG. *Silicon and silicones*. Berlin Heidelberg: Springer-Verlag, 1987.
7. Horner AA. Toll-like receptor ligands and atopy: a coin with at least two sides. *J Allergy Clin Immunol* 2006; 117:1133–40.
8. Liu AH. Something old, something new: indoor endotoxin, allergens and asthma. *Paediatr Respir Rev* 2004; 5 Suppl A: S65–71.
9. Moon RY, Yang DC, Tanabe KO, Young HA, Hauck FR. Pacifier use and SIDS: Evidence for a consistently reduced risk. *Matern. Child Health J* 2012; 16: 609–14.
10. Hesselmar B, Sjöberg F, Saalman R, Åberg N, Adlerberth I, Wold AE. Pacifier cleaning practices and risk of allergy development. *Pediatrics* 2013 doi: 10.1542/peds.2012-3345.
11. Nelson-Filho P, Assed S, Raquel S, Luciana S. Disinfection of pacifiers and toothbrushes. *J Pediatric Dent.* 2011; 33:10–13.
12. Chamele J1, Bhat C, Saraf T *et al.* Efficacy of microwaves and chlorhexidine for disinfection of pacifiers and toothbrushes: an *in vitro* study. *J Contemp Dent Pract* 2012; 13:690–4.
13. Da Silva FC, Spolidorio DMP, Zuanon ACC, Godoi RHM. Pacifier disinfection procedure: superficial morphological aspects and microorganisms colonization. *RSBO* 2008; 5:30–3.
14. Topcuoglu N, Bozdogan E, Ozsoy SD *et al.* Prevalence of salivary *Streptococcus mutans* serotype k in children undergoing congenital heart surgery. *J Clin Pediatr Dent* 2013; 38:175–8.
15. Peterson PE. Continuous Improvement of Oral Health in the 21st Century – the approach of the WHO Global Oral Health Programme; World Oral Health Report 2003, WHO/NMH/NPH/ORH/03.
16. Morton H E. Alcohols. In: Bloch S S, editor. *Disinfection, Sterilization, and Preservation*. 3rd ed. Philadelphia, Pa: Lea & Febiger; 1983; 225–39.
17. Leyes BJL, Garcia FM, Gallas TM. Efficacy of chlorhexidine with and without alcohol: a clinical study. *J Periodontol* 2002; 73:317–21.
18. Mima EG, Pavarina AC, Neppelenbroek KH, Vergani CE, Spolidorio DM, Machado AL. Effect of different exposure times on microwave irradiation on the disinfection of a hard chairside relined resin. *J Prosth* 2008; 17:312–7.
19. Yezdani A, Mahalakshmi K, Padmavathy K. Orthodontic instrument sterilization with microwave irradiation. *J Pharm Bioall Sci* 2015;7: Suppl S1:111–5